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(57) Abstract

The present invention provides isolated polypeptides useful in the treatment and prevention of malaria caused by *Plasmodium falciparum* or *P. vivax*. In particular, the polypeptides are derived from the binding domains of the proteins in the DBL family as well as the sialic acid binding protein (SABP) on *P. falciparum* merozoites. The polypeptides may also be derived from the Duffy antigen binding protein (DABP) on *P. vivax* merozoites.

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BINDING DOMAINS FROM PLASMODIUM VIVAX AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING PROTEINS

BACKGROUND OF THE INVENTION.

Malaria infects 200 - 400 million people each year causing 1-2 million deaths, thus remaining one of the most important infectious diseases in the world. Approximately 25 percent of all deaths of children in rural Africa between the ages of one and four years are caused by malaria. Due to the importance of the disease as a worldwide health problem, considerable effort is being expended to identify and develop malaria vaccines.

Malaria in humans is caused by four species of the parasite *Plasmodium: P. falciparum, P. vivax, P. knowlesi* and *P. malariae*. The major cause of malaria in humans is *P. falciparum* which infects 200 million to 400 million people every year, killing 1 to 4 million.

Duffy Antigen Binding Protein (DABP) and Sialic Acid Binding Protein (SABP) are soluble proteins that appear in the culture supernatant after infected erythrocytes release merozoites. Immunochemical data indicate that DABP and SABP which are the respective ligands for the *P. vivax* and *P. falciparum* Duffy and sialic acid receptors on erythrocytes, possess specificities of binding which are identical either in soluble or membrane bound form.

DABP is a 135 kDa protein which binds specifically to Duffy blood group determinants (Wertheimer et al., Exp. Parasitol. 69: 340-350 (1989); Barnwell, et al., J. Exp. Med. 169: 1795-1802 (1989)). Thus, binding of DABP is specific to human Duffy positive erythrocytes. There are four major Duffy phenotypes for human erythrocytes: Fy(a), Fy(b), Fy(ab) and Fy(negative), as defined by the anti-Fy^a and anti-Fy^b sera (Hadley et al., In Red Cell Antigens and Antibodies, G. Garratty, ed. (Arlington, Va.:American Association of Blood Banks) pp. 17-33 (1986)). DABP binds equally to both Fy(a) and Fy(b) erythrocytes which are equally susceptible to invasion by P. vivax; but not to Fy(negative) erythrocytes.

In the case of SABP, a 175kDa protein, binding is specific to the glycophorin sialic acid residues on erythrocytes (Camus and Hadley, *Science* 230:553-556 (1985); Orlandi, *et al.*, *J. Cell Biol.* 116:901-909 (1992)). Thus, neuraminidase treatment (which cleaves off sialic acid residues) render erythrocytes immune to *P. falciparus. invasion*.

The specificities of binding and correlation to invasion by the parasite thus indicate that DABP and SABP are the proteins of *P. vivax* and *P. falciparum* which interact with sialic acids and the Duffy antigen on the erythrocyte. The genes encoding both proteins have been cloned and the DNA and predicted protein sequences have been determined (B. Kim Lee Sim, et al., J. Cell Biol. 111: 1877-1884 (1990); Fang, X., et al., Mol. Biochem Parasitol. 44: 125-132 (1991)).

Despite considerable research efforts worldwide, because of the complexity of the *Plasmodium* parasite and its interaction with its host, it has not been possible to discover a satisfactory solution for prevention or abatement of the blood stage of malaria. Because malaria is a such a large worldwide health <u>problem</u>, there is a need for methods that abate the impact of this disease. The present invention provides effective preventive and therapeutic measures against *Plasmodium* invasion.

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SUMMARY OF THE INVENTION

The present invention provides compositions comprising an isolated DABP binding domain polypeptides and/or isolated SABP binding domain polypeptides. The DABP binding domain polypeptides preferably comprise between about 200 and about 300 amino acid residues while the SABP binding domain polypeptides preferably comprises between about 200 and about 600 amino acid residues. A preferred DABP binding domain polypeptide has about 325 residues of the amino acid sequence found in SEQ ID NO:2. A preferred SABP binding domain polypeptide has about 616 residues of the amino acid sequence of SEQ ID NO:4, encoded by the DNA sequence of SEQ ID NO: 3. The preferred DABP binding domain and SABP binding domain include the cysteine-rich portions of the proteins shown in Figure 1.

The present invention also includes pharmaceutical compositions comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* merozoites in an organism. In addition, isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* may be added

to the pharmaceutical composition.

Also provided are pharmaceutical compositions comprising a pharmaceutically acceptable carrier and an isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* merozoites in an organism. In addition, isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* may be added to the pharmaceutical composition.

Isolated polynucleotides which encode a DABP binding domain polypeptides or SABP binding domain polypeptides are also disclosed. In addition, the present invention includes a recombinant cell comprising the polynucleotide encoding the DABP binding domain polypeptide.

The current invention further includes methods of inducing a protective immune response to Plasmodium merozoites in a patient. The methods comprise administering to the patient an immunologically effective amount of a pharmaceutical composition comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide, an SABP binding domain polypeptide or a combination thereof.

The present disclosure also provides DNA sequences from additional *P. falciparum* genes in the Duffy-binding like (*DBL*) family that have regions conserved with the *P. falciparum* 175 kD and *P. vivax* 135 kD binding proteins.

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DEFINITIONS

As used herein a "DABP binding domain polypeptide" or a "SABP binding domain polypeptide" are polypeptides substantially identical (as defined below) to a sequence from the cysteine-rich, amino-terminal region of the Duffy antigen binding protein (DABP) or sialic acid binding protein (SABP), respectively. Such polypeptides are capable of binding either the Duffy antigen or sialic acid residues on glycophorin. In particular, DABP binding domain polypeptides consist of amino acid residues substantially similar to a sequence of SABP within a binding domain

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containing the cysteine-rich sequence shown in Figure 1. SABP binding domain polypeptides consist of residues substantially similar to a sequence of DABP within a binding domain containing the cysteine-rich sequence shown in Figure 1.

The binding domain polypeptides encoded by the genes of the *DBL* family consist of those residues substantially identical to the sequence of the binding domains of DABP and SABP as defined above. The DBL family comprises sequences with substantial similarity to the conserved regions of the DABP and SABP. These include those sequences reported here as *ebl-1* (SEQ ID NO:5 and SEQ ID NO:6), E31a (SEQ ID NO:7 and SEQ ID NO:8), *var-7* (SEQ. ID. NO:13 and SEQ. ID. NO:14, GenBank Accession No. L42636) and *var-1* (SEQ. ID. NO:15 and SEQ ID NO:16, GenBank Accession No. L40608). The sequence *ebl-2*, (SEQ ID NO:9 and SEQ ID NO:10) represents the binding domains of *var-7*, and Proj3 (SEQ ID NO:11 and SEQ ID NO:12) is the binding domain of *var-1*. The DBL family also includes two other members *var-2* and *var-3* (GenBank Accession No. L40609).

The polypeptides of the invention can consist of the full length binding domain or a fragment thereof. Typically DABP binding domain polypeptides will consist of from about 50 to about 325 residues, preferably between about 75 and 300, more preferably between about 100 and about 250 residues. SABP binding domain polypeptides will consist of from about 50 to about 616 residues, preferably between about 75 and 300, more preferably between about 100 and about 250 residues.

Particularly preferred polypeptides of the invention are those within the binding domain that are conserved between SABP and the *DBL* family. Residues within these conserved domains are shown in Figure 1, below.

Two polynucleotides or polypeptides are said to be "identical" if the sequence of nucleotides or amino acid residues in the two sequences is the same when aligned for maximum correspondence. Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman Adv. Appl. Math. 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch J. Mol. Biol. 48:443 (1970), by the search for similarity method of Pearson and Lipman Proc. Natl. Acad. Sci. (U.S.A.) 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by inspection. term "substantial identity" means that a polypeptide comprises a sequence that has at least 80% sequence identity, preferably 90%, more preferably 95% or more, compared to a reference sequence over a comparison window of about 20 residues to about 600 residues- typically about 50 to about 500 residues usually about 250 to 300 residues. The values of percent identity are determined using the programs above. Particularly preferred peptides of the present invention comprise a sequence in which at least 70% of the cysteine residues conserved in DABP and SABP are present. Additionally, the peptide will comprise a sequence in which at least 50% of the tryptophan residues conserved in DABP and SABP are present. The term substantial similarity is also specifically defined here with respect to those amino acid residues found to be conserved between UABP, SABP and the sequences of the DBL family. These conserved amino acids consist prominently of tryptophan and cysteine residues conserved among all sequences reported here. In addition the conserved amino acid residues include phenylalanine residues which may

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be substituted with tyrosine. These amino acid residues may be determined to be conserved after the sequences have been aligned using methods outlined above by someone skilled in the art.

Another indication that polypeptide sequences are substantially identical is if one protein is immunologically reactive with antibodies raised against the other protein. Thus, the polypeptides of the invention include polypeptides immunologically reactive with antibodies raised against the SABP binding domain, the DABP binding domain or raised against the conserved regions of the DBL family.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. Stringent conditions are sequence dependent and will be different in different circumstances. Generally, stringent conditions are selected to be about 5° C lower than the thermal melting point (Tm) for the specific sequence at a defined ionic strength and pH. The Tm is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typically, stringent conditions will be those in which the salt concentration is about 0.02 molar at pH 7 and the temperature is at least about 60°C.

Nucleotide sequences are also substaintially identical for purposes of this application when the polypeptides which they encode are substantially identical. Thus, where one nucleic acid sequence encodes essentially the same polypeptide as a second nucleic acid sequence, the two nucleic acid sequences are substantially identical, even if they would not hybridize under stringent conditions due to silent substitutions permitted by the genetic code (see, Darnell et al. (1990) Molecular Cell Biology, Second Edition Scientific American Books, W.H. Freeman and Company, New York, NY, for an explanation of codon degeneracy and the genetic code).

The phrases "isolated" or "biologically pure" refer to material which is substantially or essentially free from components which normally accompany it as found in its native state. Thus, the binding domain polypeptides of this invention do not contain materials normally associated with their *in situ* environment, e.g., other proteins from a merozoite membrane. Typically, isolated proteins of the invention are at least about 80% pure, usually at least about 90%, and preferrably at least about 95% as measured by band intensity on a silver stained gel.

Protein purity or homogeneity may be indicated by a number of means well known in the art, such as polyacrylamide gel electrophoresis of a protein sample, followed by visualization upon staining. For certain purposes high resolution will be needed and HPLC or a similar means for purification utilized.

The term "residue" refers to an amino acid (D or L) or amino acid mimetic incorporated in a oligopeptide by an amide bond or amide bond mimetic. An amide bond mimetic of the invention includes peptide backbone modifications well known to those skilled in the art.

BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1 represents an alignment of the predicted amino acid sequences of the DABP binding domain (Vivax) (SEQ ID NO:25), the two homologous SABP domains (SABP F1 (SEQ ID NO:26) and SABP F2 (SEQ ID NO:27)) and the sequenced members of the *DBL* gene family (ebl-1 (SEQ ID NO:38), E31a (SEQ ID NO:39), EBL-2 (SEQ ID NO:30)) and the three homologous Proj3 domains (F1 (SEQ ID NO:31), F2 (SEQ ID NO:32) and F3 (SEQ ID NO:33)).

Figure 2 represents a schematic of the pRE4 cloning vector.

Figure 3 shows primers useful for isolating sequences encoding the conserved motifs of the invention. Primers UNIEBP5 (SEQ ID NO:35) and UNIEBP5A (SEQ ID NO:36) encode the amino acid sequence of SEQ ID NO:34; primers UNIEBP5B (SEQ ID NO:38) and UNIEBP5C (SEQ ID NO:39) encode the amino acid sequence of SEQ ID NO:37; primers UNIEBP3 (SEQ ID NO:41) and UNIEBP3A (SEQ ID NO:42) encode the amino acid sequence of SEQ ID NO:40; and primers UNIEBP3B (SEQ ID NO:44) and UNIEBP3C (SEQ ID NO:45) encode the amino acid sequence of SEQ ID NO:43.

Figure 4 shows the relative position of the E31a ORF on chromosome 7.

Figure 5 shows a map of a *var* gene cluster on chromosome 7. Relative positions of four YACs (PfYEF2, PfYFE6, PfYKF8, PfYED9) are indicated under the chromosome 7 line at the top of the figure. YACs PfYFE6 and PfYKF8 lie entirely within a segment linked to CQR in a genetic cross, whereas YACs PfYED9 and PfYEF2 extend beyond sites (identified by pE53a and pH270.5) that are dissociated from the chloroquine response. The *var* cluster extends over a region of 100-150 kb in PfYED9. Exons and introns of the *var-1*, *var-2* and *var-3* genes within the sequenced 40 kb segment are represented by solid and dotted lines, respectively; arrows show the coding direction. Two more *var* elements outside of the sequenced region, identified by conserved restriction sites and cross-hybridization, are indicated by dashed-lines (*var-2c* and *var-3c*). Bold letters mark repeated restriction sites that suggest a duplication in the *var-2lvar-3* and *var-2clvar-3c* segments. Enzyme recognition sites: A, *Apal*; B, *Bgh*; C, *Cla*; D, *Hind*III; E, *Hae*III; K, *Kpn*I; M, *Bam*HI; P, *Hpal*; S, *Smal*. *Hind*IIII and *Hae*III sites outside of the sequenced region were not mapped. Positions and sizes of inserts from the Dd2 subsegment library are indicated: a, pE280b; b, pB20.3; c, pB600; d, pE21b; e, pB20.24; f, pE32b; h, pE241a; i, pE240a/51d; j, pE33a; k, pB20.23; l, *A*L17BA6; m, pB20.26; n, pB20SU.27; o, p15J2J3. Inserts from the PfYED9 34 kb *Apal-Smal* fragment library: r, pB3; s, p3G11; t, pJVs; u, p2E10; v, pIG3; w, p2E3; x, p2B6; y, PE10; z, pJYr; α, pC5; β, p1A3; γ, p1F6; δ, p3C3; ε, pA2; ζ, p2A9; η, p3C4; θ, pJZn; κ, p3D8.

DESCRIPTION OF THE PREFERRED EMBODIMENT

The binding of merozoites and schizonts to erythrocytes is mediated by specific binding proteins on the surface of the merozoite or schizont and is necessary for erythrocyte invasion. In the case of *P. falciparum*, this binding involves specific interaction between sialic acid glycophorin residues on the erythrocyte and the sialic acid binding protein (SABP) on the surface of the merozoite or schizont. The ability of purified SABP to bind erythrocytes with chemically or enzymatically altered sialic acid residues paralleled the ability of *P. falciparum* to invade these erythrocytes. Furthermore, sialic acid deficient erythrocytes neither bind SABP nor support invasion by *P. falciparum*. The DNA encoding SABP from *P. falciparum* has also been cloned and sequenced.

In *P. vivax*, specific binding to the erythrocytes involves interaction between the Duffy blood group antigen on the erythrocyte and the Duffy antigen binding protein (DABP) on the merozoite. Duffy binding proteins were defined biologically as those soluble proteins that appear in the culture supernatant after the infected erythrocytes release merozoites which bind to human Duffy positive, but not to human Duffy negative erythrocytes. It has been shown that binding of the *P. vivax* DABP protein to Duffy positive erythrocytes is blocked by antisera to the Duffy blood group determinants. Purified Duffy blood group antigens also block the binding to erythrocytes. DABP has also been shown to bind Duffy blood group determinants on Western blots.

Duffy positive blood group determinants on human erythrocytes are essential for invasion of human erythrocytes by *Plasmodium vivax*. Both attachment and reorientation of *P. vivax* merozoites occur equally well on Duffy positive and negative erythrocytes. A junction then forms between the apical end of the merozoite and the Duffy-positive erythrocyte, followed by vacuole formation and entry of the merozoite into the vacuole. Junction formation and merozoite entry into the erythrocyte do not occur on Duffy negative cells, suggesting that the receptor specific for the Duffy determinant is involved in apical junction formation but not initial attachment. The DNA sequences encoding the DABP from *P. vivax* and *P. knowlesi* have been cloned and sequenced.

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P. vivax red cell invasion has an absolute requirement for the Duffy blood group antigen. Isolates of P. falciparum, however, vary in their dependency on sialic acid for invasion. Certain P. falciparum clones have been developed which invade sialic acid deficient erythrocytes at normal rates. This suggests that certain strains of P. falciparum can interact with other ligands on the erythrocyte and so may possess multiple erythrocyte binding proteins with differing specificities.

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A basis for the present invention is the discovery of the binding domains in both DABP and SABP. Comparison of the predicted protein sequences of DABP and SABP reveals an amino-terminal, cysteine-rich region in both proteins with a high degree of similarity between the two proteins. The amino-terminal, cysteine-rich region of DABP contains about 325 amino acids, whereas the amino-terminal, cysteine-rich region of SABP contains about 616 amino acids. This is due to an apparent duplication of the amino-terminal, cysteine-rich region in the SABP protein. The cysteine residues are conserved between the two regions of SABP and DABP, as are the amino acids surrounding the cysteine residues and a number of aromatic amino acid residues in this region. The amino-terminal cysteine rich region and another cysteine-rich region near the carboxyl-terminus show the most similarity between the DABP and SABP proteins. The region of the amino acid sequence between these two cysteine-rich regions show only limited similarity between DABP and SABP.

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Other *P. falciparum* open reading frames and genes with regions that have substantial identity to binding domains of SABP and DABP have been identified. Multiple copies of these sequences exist in the parasite genome, indicating their important activity in host-parasite interactions. A family of these sequences (the *DBL* family) have been cloned from chromosome 7 subsegment libraries that were constructed during genetic studies of the chloroquine resistance locus (Wellems *et. al.*, *PNAS* 88: 3382-3386 (1991)). Certain of these transcripts are known to be from the *var* family of genes that modulate cytoadherence and antigenic variation of *P. falciparum*- infected erythrocytes (*see*, Example 3, below).

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Genes of the *P. falciparum var* family encode 200-350 kD variant surface molecules that determine antigenic and adhesive properties of parasitized erythrocytes. The large repertoire of *var* genes (50-150 copies, having sufficient DNA to account for 2-6% of the haploid genome), the dramatic sequence variation among the gene copies, their variable expression in different parasite lines, the ready detection of DNA rearrangements, and the receptor binding features of the encoded extracellular domains all implicate *var* genes as the major determinants of antigenic variation and cytoadherence in *P. falciparum* malaria.

A second class of *DBL*-encoding transcripts includes single-copy genes such as *ebl-1*. Genetic linkage studies have placed this gene within a region of chromosome 13 that affects invasion of malarial parasites in human red blood cells (Wellems *et al.*, *Cell* 49:633-642 (1987)). Both SABP and *ebl-1* show restriction patterns that are well conserved among different parasite isolates. This conservation of gene structure and the sequence relationships between the *ebl-1* and SABP domains suggest that *ebl-1* encodes a novel erythrocyte binding molecule having receptor properties distinct from those of SABP.

Southern hybridization experiments using probes from these open reading frames have indicated that additional copies of these conserved sequences are located elsewhere in the genome. The largest of the open reading frames on chromosome 7 is 8 kilobases and contains four tandem repeats homologous to the N-terminal, cysteine-rich unit of SABP and DABP.

Figure 1 represents an alignment of the DBL family with the DABP binding domain and two homologous regions of SABP (F₁ and F₂). The DBL family is divided into two sub-families to achieve optimal alignment. Conserved cysteine residues are shown in bold face and conserved aromatic residues are underlined.

The polypeptides of the invention can be used to raise monoclonal antibodies specific for the binding domains of SABP, DABP or the conserved regions in the *DBL* gene family. The antibodies can be used for diagnosis of malarial infection or as therapeutic agents to inhibit binding of merozoites to erythrocytes. The production of monoclonal antibodies against a desired antigen is well known to those of skill in the art and is not reviewed in detail here.

The multitude of techniques available to those skilled in the art for production and manipulation of various immunoglobulin molecules can thus be readily applied to inhibit binding. As used herein, the terms "immunoglobulin" and "antibody" refer to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. Immunoglobulins may exist in a variety of forms besides antibodies, including for example, Fv, Fab, and F(ab)₂, as well as in single chains. For a general review of immunoglobulin structure and function see, Fundamental Immunology, 2d Ed., W.E. Paul ed., Ravens Press, N.Y., (1989).

Antibodies which bind polypeptides of the invention may be produced by a variety of means. The production of non-human monoclonal antibodies, e.g., murine, lagomorpha, equine, etc., is well known and may be accomplished by, for example, immunizing the animal with a preparation containing the polypeptide. Antibody-producing cells obtained from the immunized animals are immortalized and screened, or screened first for the production of antibody which inhibits binding between and meroxoites and erythrocytes and then immortalized.

For a discussion of general procedures of monoclonal antibody production see Harlow and Lane, *Antibodies, A Laboratory Manual* Cold Spring Harbor Publications, N.Y. (1988).

Thus, the present invention allows targeting of protective immune responses or monoclonal antibodies to sequences in the binding domains that are conserved between SABP, DABP and encoded regions of the DBL family. Identification of the binding regions of these proteins facilitates vaccine development because it allows for a focus of effort upon the functional elements of the large molecules. The particular sequences within the binding regions refine the target to critical regions that have been conserved during evolution, and are thus preferred for use as vaccines against the parasite.

The genes of the *DBL* family (which have not previously been sequenced) can be used as markers to detect the presence of the *P. falciparum* parasite in patients. This can be accomplished by means well known to practitioners in the art using tissue or blood from symptomatic patients in PCR reactions with oligonucleotides complementary to portions of the genes of the *DBL* family. Furthermore, sequencing the *DBL* family provides a means for skilled practitioners to generate defined probes to be used as genetic markers in a variety of applications.

Additionally, the present invention defines a conserved motif present in, but not restricted to other members of the subphylum Apicomplexa which participates in host parasite interaction. This motif can be identified in Plasmodium species and other parasitic protozoa by the polymerase chain reaction using the synthetic oligonucleotide primers shown in Figure 3. PCR methods are described in detail below. These primers are designed from regions in the conserved motif showing the highest degree of conservation among DABP, SABP and the DBL family. Figure 3 shows these regions and the consensus amino acid sequences derived from them.

A. General Methods

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Much of the nomenclature and general laboratory procedures required in this application can be found in Sambrook, et al., Molecular Cloning A Laboratory Manual, 2nd Ed., Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989. The manual is hereinafter referred to as "Sambrook, et al., 1989."

The practice of this invention involves the construction of recombinant nucleic acids and the expression of genes in transfected cells. Molecular cloning techniques to achieve these ends are known in the art. A wide variety of cloning and *in vitro* amplification methods suitable for the construction of recombinant nucleic acids are well-known to persons of skill. Examples of these techniques and instructions sufficient to direct persons of skill through many cloning exercises are found in Berger and Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology* volume 152 Academic Press, Inc., San Diego, CA (Berger); and *Current Protocols in Molecular Biology*, F.M. Ausubel *et al.*, eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., (1994 Supplement) (Ausubel).

Examples of techniques sufficient to direct persons of skill through *in vitro* amplification methods, including the polymerase chain reaction (PCR) the ligase chain reaction (LCR), O.B.-replicase amplification and other RNA polymerase mediated techniques are found in Berger, Sambrook et al., 1989, and Ausubel, as well as Mullis *et al.*, (1987) U.S. Patent No. 4,683,202; *PCR Protocols A Guide to Methods and Applications* (Innis *et al.* eds), Academic Press Inc., San Diego, CA, 1990) ("Innis"); Arnheim & Levinson (October 1, 1990) *C&EN* 36-47; *The*

Journal Of NIH Research (1991) 3, 81-94; Kwoh et al. (1989) Proc. Natl. Acad. Sci. USA 86, 1173; Guatelli et al. (1990) Proc. Natl. Acad. Sci. USA 87, 1874; Lomell et al. (1989) J. Clin. Chem 35, 1826; Landegren et al., (1988) Science 241, 1077-1080; Van Brunt (1990) Biotechnology 8, 291-294; Wu and Wallace, (1989) Gene 4, 560; and Barringer et al. (1990) Gene 89, 117. Improved methods of cloning in vitro amplified nucleic acids are described in Wallace et al., U.S. Pat. No. 5,426,039.

The culture of cells used in the present invention, including cell lines and cultured cells from tissue or blood samples is well known in the art. Freshney (*Culture of Animal Cells, a Manual of Basic Technique, third ed.*, Wiley-Liss, New York, NY (1994)) and the references cited therein provides a general guide to the culture of cells.

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DBL genes are optionally bound by antibodies in one of the embodiments of the present invention. Methods of producing polyclonal and monoclonal antibodies are known to those of skill in the art. See, e.g., Coligan (1991) Current Protocols in Immunology Wiley/Greene, NY; and Harlow and Lane (1989) Antibodies: A Laboratory Manual Cold Spring Harbor Press, NY; Stites et al. (eds.) Basic and Clinical Immunology (4th ed.) Lange Medical Publications, Los Altos, CA, and references cited therein; Goding (1986) Monoclonal Antibodies: Principles and Practice (2d ed.) Academic Press, New York, NY; and Kohler and Milstein (1975) Nature 256: 495-497. Other suitable techniques for antibody preparation include selection of libraries of recombinant antibodies in phage or similar vectors. See, Huse et al. (1989) Science 246: 1275-1281; and Ward, et al. (1989) Nature 341: 544-546. Specific Monoclonal and polyclonal antibodies will usually bind with a KD of at least about .1 mM, more usually at least about 1 µM, and most preferably at least about .1 µM or better.

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B. Methods for isolating DNA encoding SABP, DABP and DBL binding regions

The nucleic acid compositions of this invention, whether RNA, cDNA, genomic DNA, or a hybrid of the various combinations, may be isolated from natural sources or may be synthesized in vitro. The nucleic acids claimed may be present in transformed or transfected whole cells, in a transformed or transfected cell lysate, or in a partially purified or substantially pure form.

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Techniques for nucleic acid manipulation of genes encoding the binding domains of the invention, such as subcloning nucleic acid sequences encoding polypeptides into expression vectors, labelling probes, DNA hybridization, and the like are described generally in Sambrook *et al.*, 1989.

Recombinant DNA techniques can be used to produce the binding domain polypeptides. In general, the DNA encoding the SABP and DABP binding domains are first cloned or isolated in a form suitable for ligation into an expression vector. After ligation, the vectors containing the DNA fragments or inserts are introduced into a suitable host cell for expression of the recombinant binding domains. The polypeptides are then isolated from the host cells.

There are various methods of isolating the DNA sequences encoding the SABP, DABP and DBL binding domains. Typically, the DNA is isolated from a genomic or cDNA library using labelled oligonucleotide probes specific for sequences in the DNA. Restriction endonuclease digestion of genomic DNA or cDNA containing the appropriate genes can be used to isolate the DNA encoding the binding domains of these proteins. Since the DNA

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sequences of the SABP and DABP genes are known, a panel of restriction endonucleases can be constructed to give cleavage of the DNA in the desired regions. After restriction endonuclease digestion, DNA encoding SABP binding domain or DABP binding domain is identified by its ability to hybridize with nucleic acid probes, for example on Southern blots, and these DNA regions are isolated by standard methods familiar to those of skill in the art. See Sambrook, et al., 1989.

The polymerase chain reaction can also be used to prepare DABP, SABP DBL binding domain DNA. Polymerase chain reaction technology (PCR) is used to amplify nucleic acid sequences of the DABP and SABP binding domains directly from mRNA, from cDNA, and from genomic libraries or cDNA libraries. The primers shown in Figure 3 are particularly preferred for this process.

Appropriate primers and probes for amplifying the SABP and DABP binding region DNA's are generated from analysis of the DNA sequences. In brief, oligonucleotide primers complementary to the two 3' borders of the DNA region to be amplified are synthesized. The polymerase chain reaction is then carried out using the two primers. See *PCR Protocols: A Guide to Methods and Applications*. (Innis, M, Gelfand, D., Sninsky, J. and White, T., (eds.), Academic Press, San Diego, CA (1990). Primers can be selected to amplify the entire DABP regions or to amplify smaller segments of the DABP and SABP binding domains, as desired.

Oligonucleotides for use as probes are chemically synthesized according to the solid phase phosphoramidite triester method first described by Beaucage, S.L. and Caruthers, M.H., 1981, Tetrahedron Letts., 22(20):1859-1862 using an automated synthesizer, as described in Needham-VanDevanter, D.R., et al. 1984, Nucleic Acids Res., 12:6159-6168. Purification of oligonucleotides is by either native acrylamide gel electrophoresis or by anion-exchange HPLC as described in Pearson, J.D. and Regnier, F.E., 1983, J. Chrom., 255:137-149.

The sequence of the synthetic oligonucleotides can be verified using the chemical degradation method of Maxam, A.M. and Gilbert, 1980, in W., Grossman, L. and Moldave, D., eds. Academic Press, New York, NY, *Methods in Enzymology* 65:499-560.

Other methods known to those of skill in the art may also be used to isolate DNA encoding all or part of the SABP or DABP binding domains. See Sambrook, et al., 1989.

C. Expression of DABP, SABP and DBL Binding Domain Polypeptides

Once binding domain DNAs are isolated and cloned, one may express the desired polypeptides in a recombinantly engineered cell such as bacteria, yeast, insect (especially employing baculoviral vectors), and mammalian cells. It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of the DNA encoding the DABP and SABP binding domains. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of natural or synthetic nucleic acids encoding binding domains will typically be achieved by operably linking the DNA or cDNA to a promoter (which is either constitutive or inducible), followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding the

binding domains. To obtain high level expression of a cloned gene, it is desirable to construct expression plasmids which contain, at the minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator.

1. Expression in Prokaryotes

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Examples of regulatory regions suitable for this purpose in *E. coli* are the promoter and operator region of the *E. coli* tryptophan biosynthetic pathway as described by Yanofsky, C., 1984, J. Bacteriol., 158:1018-1024 and the leftward promoter of phage lambda (P_L) as described by Herskowitz, I. and Hagen, D., 1980, Ann. Rev. Genet., 14:399-445. The inclusion of selection markers in DNA vectors transformed in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol. See Sambrook *et al.*, 1989, for details concerning selection markers for use in *E. coli*.

The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA.

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Expression systems for expressing the DABP and SABP binding domains are available using *E. coli, Bacillus* sp. (Palva, I *et al.*, 1983, Gene 22:229-235; Mosbach, K. *et al.* Nature, 302:543-545 and *Salmonella. E. coli* systems are preferred.

The binding domain polypeptides produced by prokaryote cells may not necessarily fold properly. During purification from *E. coli*, the expressed polypeptides may first be denatured and then renatured. This can be accomplished by solubilizing the bacterially produced proteins in a chaotropic agent such as guanidine HCl and reducing all the cysteine residues with a reducing agent such as beta-mercaptoethanol. The polypeptides are then renatured, either by slow dialysis or by gel filtration. U.S. Patent No. 4,511,503.

Detection of the expressed antigen is achieved by methods known in the art as radioimmunoassays, Western blotting techniques or immunoprecipitation. Purification from *E. coli* can be achieved following procedures described in U.S. Patent No. 4,511,503.

2. Synthesis of SABP, DABP and DBL Binding Domains in Eukaryotes

A variety of eukaryotic expression systems such as yeast, insect cell lines and mammalian cells, are known to those of skill in the art. As explained briefly below, the DABP and SABP binding domains may also be expressed in these eukaryotic systems.

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a. Expression in Yeast

Synthesis of heterologous proteins in yeast is well known and described. *Methods in Yeast Genetics*, Sherman, F., *et al.*, Cold Spring Harbor Laboratory, (1982) is a well recognized work describing the various methods available to produce the binding domains in yeast.

Examples of promoters for use in yeast include GAL1,10 (Johnson, M., and Davies, R.W., 1984, Mol. and Cell. Biol., 4:1440-1448) ADH2 (Russell, D., *et al.* 1983, J. Biol. Chem., 258:2674-2682), PH05 (EMBO J. 6:675-680, 1982), and MFal (Herskowitz, I. and Oshima, Y., 1982, in The Molecular Biology of the Yeast

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Saccharomyces, (eds. Strathern, J.N. Jones, E.W., and Broach, J.R., Cold Spring Harbor, Lab., Cold Spring Harbor, N.Y., pp. 181-209. A multicopy plasmid with a selective marker such as Leu-2, URA-3, Trp-1, and His-3 is also desirable.

A number of yeast expression plasmids like YEp6, YEp13, YEp4 can be used as vectors. A gene of interest can be fused to any of the promoters in various yeast vectors. The above-mentioned plasmids have been fully described in the literature (Botstein, et al., 1979, Gene, 8:17-24; Broach, et al., 1979, Gene, 8:121-133).

Two procedures are used in transforming yeast cells. In one case, yeast cells are first converted into protoplasts using zymolyase, lyticase or glusulase, followed by addition of DNA and polyethylene glycol (PEG). The PEG-treated protoplasts are then regenerated in a 3% agar medium under selective conditions. Details of this procedure are given in the papers by J.D. Beggs, 1978, Nature (London), 275:104-109; and Hinnen, A., et al., 1978, Proc. Natl. Acad. Sci. USA, 75:1929-1933. The second procedure does not involve removal of the cell wall. Instead the cells are treated with lithium chloride or acetate and PEG and put on selective plates (Ito, H., et al., 1983, J. Bact., 153:163-168).

The binding domains can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassays of other standard immunoassay techniques.

b. Expression in Mammalian and Insect Cell Cultures

Illustrative of cell cultures useful for the production of the binding domains are cells of insect or mammalian origin. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. Illustrative examples of mammalian cell lines include VERO and HeLa cells, Chinese hamster overy (CHO) cell lines, W138, BHK, Cos-7 or MDCK cell lines.

As indicated above, the vector, e. g., a plasmid, which is used to transform the host cell, preferably contains DNA sequences to initiate transcription and sequences to control the translation of the antigen gene sequence. These sequences are referred to as expression control sequences. When the host cell is of insect or mammalian origin illustrative expression control sequences are obtained from the SV-40 promoter (Science, 222:524-527, 1983), the CMV I.E. Promoter (Proc. Natl. Acad. Sci. 81:659-663, 1984) or the metallothionein promoter (Nature 296:39-42, 1982). The cloning vector containing the expression control sequences is cleaved using restriction enzymes and adjusted in size as necessary or desirable and ligated with DNA coding for the SABP or DABP polypeptides by means well known in the art.

As with yeast, when higher animal host cells are employed, polyadenlyation or transcription terminator sequences from known mammalian genes need to be incorporated into the vector. An example of a terminator sequence is the polyadenlyation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VPI intron from SV40 (Sprague, J. et al., 1983, J. Virol. 45: 773-781).

Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors. Saveria-Campo, M., 1985, "Bovine Papilloma virus

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DNA a Eukaryotic Cloning Vector* in DNA Cloning Vol. II a Practical Approach Ed. D.M. Glover, IRL Press, Arlington, Virginia pp. 213-238.

The host cells are competent or rendered competent for transformation by various means. There are several well-known methods of introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextran, electroporation and micro-injection of the DNA directly into the cells.

The transformed cells are cultured by means well known in the art. <u>Biochemical Methods in Cell Culture and Virology</u>, Kuchler, R.J., Dowden, Hutchinson and Ross, Inc., (1977). The expressed DABP and SABP binding domain polypeptides are isolated from cells grown as suspensions or as monolayers. The latter are recovered by well known mechanical, chemical or enzymatic means.

c. <u>Expression in recombinant vaccinia virus- or adenovirus-infected cells</u>

In addition to use in recombinant expression systems, the isolated binding domain DNA sequences can also be used to transform viruses that transfect host cells in the patient. Live attenuated viruses, such as vaccinia or adenovirus, are convenient alternatives for vaccines because they are inexpensive to produce and are easily transported and administered. Vaccinia vectors and methods useful in immunization protocols are described, for example, in U.S. Patent No. 4,722,848.

Suitable viruses for use in the present invention include, but are not limited to, pox viruses, such as canarypox and cowpox viruses, and vaccinia viruses, alpha viruses, adenoviruses, and other animal viruses. The recombinant viruses can be produced by methods well known in the art, for example, using homologous recombination or ligating two plasmids. A recombinant canarypox or cowpox virus can be made, for example, by inserting the DNA's encoding the DABP and SABP binding domain polypeptides into plasmids so that they are flanked by viral sequences on both sides. The DNA's encoding the binding domains are then inserted into the virus genome through homologous recombination.

A recombinant adenovirus can be produced, for example, by ligating together two plasmids each containing about 50% of the viral sequence and the DNA sequence encoding erythrocyte binding domain polypeptide. Recombinant RNA viruses such as the alpha virus can be made via a cDNA intermediate using methods known in the art.

In the case of vaccinia virus (for example, strain WR), the DNA sequence encoding the binding domains can be inserted in the genome by a number of methods including homologous recombination using a transfer vector, pTKgpt-OFIS as described in Kaslow, et al., Science 252:1310-1313 (1991).

Alternately the DNA encoding the SABP and DABP binding domains may be inserted into another plasmid designed for producing recombinant vaccinia, such as pGS62, Langford, C.L., et al., 1986, Mol. Cell. Biol. 6:3191-3199. This plasmid consists of a cloning site for insertion of foreign genes, the P7.5 promoter of vaccinia to direct synthesis of the inserted gene, and the vaccinia TK gene flanking both ends of the foreign gene.

Confirmation of production of recombinant virus can be achieved by DNA hybridization using cDNA encoding the DABP and SABP binding domain polypeptides and by immunodetection techniques using antibodies

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specific for the expressed binding domain polypeptides. Virus stocks may be prepared by infection of cells such as HELA S3 spinner cells and harvesting of virus progeny.

The recombinant virus of the present invention can be used to induce anti-SABP and anti-DABP binding domain antibodies in mammals, such as mice or humans. In addition, the recombinant virus can be used to produce the SABP and DABP binding domains by infecting host cells *in vitro*, which in turn express the polypeptide (see section on expression of SABP and DABP binding domains in eukaryotic cells, above).

The present invention also relates to host cells infected with the recombinant virus. The host cells of the present invention are preferably mammalian, such as BSC-1 cells. Host cells infected with the recombinant virus express the DABP and SABP binding domains on their cell surfaces. In addition, membrane extracts of the infected cells induce protective antibodies when used to inoculate or boost previously inoculated mammals.

D. Purification of the SABP, DABP and DBL Binding Domain Polypeptides

The binding domain polypeptides produced by recombinant DNA technology may be purified by standard techniques well known to those of skill in the art. Recombinantly produced binding domain polypeptides can be directly expressed or expressed as a fusion protein. The protein is then purified by a combination of cell lysis (e. g., sonication) and affinity chromatography. For fusion products, subsequent digestion of the fusion protein with an appropriate proteolytic enzyme release the desired SABP and DABP binding domains.

The polypeptides of this invention may be purified to substantial purity by standard techniques well known in the art, including selective precipitation with such substances as ammonium sulfate, column chromatography, immunopurification methods, and others. See, for instance, R. Scopes, *Protein Purification: Principles and Practice*, Springer-Verlag, New York, NY (1982).

E. <u>Production of Binding Domains by protein chemistry techniques</u>

The polypeptides of the invention can be synthetically prepared in a wide variety of ways. For instance polypeptides of relatively short size, can be synthesized in solution or on a solid support in accordance with conventional techniques. Various automatic synthesizers are commercially available and can be used in accordance with known protocols. See, for example, Stewart and Young, *Solid Phase Peptide Synthesis*, 2d. ed., Pierce Chemical Co. (1984).

Alternatively, purified and isolated SABP, DABP or DBL family proteins may be treated with proteolytic enzymes in order to produce the binding domain polypeptides. For example, recombinant DABP and SABP proteins may be used for this purpose. The DABP and SABP protein sequence may then be analyzed to select proteolytic enzymes to be used to generate polypeptides containing desired regions of the DABP and SABP binding domain. The desired polypeptides are then purified by using standard techniques for protein and peptide purification. For a review of standard techniques see, *Methods in Enzymology*, "Guide to Protein Purification", M. Deutscher, ed. Vol. 182 (1990), pages 619-626.

F. Modification of nucleic acid and polypeptide sequences

The nucleotide sequences used to transfect the host cells used for production of recombinant binding domain polypeptides can be modified according to standard techniques to yield binding domain polypeptides,

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with a variety of desired properties. The binding domain polypeptides of the present invention can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the binding domain polypeptides can vary from the naturally-occurring sequence at the primary structure level by amino acid insertions, substitutions, deletions, and the like. These modifications can be used in a number of combinations to produce the final modified protein chain.

The amino acid sequence variants can be prepared with various objectives in mind, including facilitating purification and preparation of the recombinant polypeptides. The modified polypeptides are also useful for modifying plasma half-life, improving therapeutic efficacy, and lessening the severity or occurrence of side effects during therapeutic use. The amino acid sequence variants are usually predetermined variants not found in nature but exhibit the same immunogenic activity as naturally occurring polypeptides. For instance, polypeptide fragments comprising only a portion (usually at least about 60-80%, typically 90-95%) of the primary structure may be produced. For use as vaccines, polypeptide fragments are typically preferred so long as at least one epitope capable of eliciting production of blocking antibodies remains.

In general, modifications of the sequences encoding the binding domain polypeptides may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Giliman and Smith, *Gene* 8:81-97 (1979) and Roberts, S. *et al.*, *Nature* 328:731-734 (1987)). One of ordinary skill will appreciate that the effect of many mutations is difficult to predict. Thus, most modifications are evaluated by routine screening in a suitable assay for the desired characteristic. For instance, changes in the immunological character of the polypeptide can be detected by an appropriate competitive binding assay. Modifications of other properties such as redox or thermal stability, hydrophobicity, susceptibility to proteolysis, or the tendency to aggregate are all assayed according to standard techniques.

G. <u>Diagnostic and Screening Assays</u>

The polypeptides and nucelic acids of the invention can be used in diagnostic applications for the detection of merozoites or nucleic acids in a biological sample. The presence of parasites can be detected using several well recognized specific binding assays based on immunological results. (See U.S. Patents 4,366,241; 4,376,110; 4,517,288; and 4,837,168). For instance, labeled monoclonal antibodies to polypeptides of the invention can be used to detect merozoites in a biological sample. Alternatively, labelled polypeptides of the invention can be used to detect the presence of antibodies to SABP or DABP in a biological sample. For a review of the general procedures in diagnostic immunoassays, see also *Basic and Clinical Immunology* 7th Edition (D. Stites and A. Terr ed.) 1991.

In addition, modified polypeptides, antibodies or other compounds capable of inhibiting the interaction between SABP or DABP and erythrocytes can be assayed for biological activity. For instance, polypeptides can be recombinantly expressed on the surface of cells and the ability of the cells to bind erythrocytes can be measured as described below. Alternatively, peptides or antibodies can tested for the ability to inhibit binding between erythrocytes and merozoites or SABP and DABP.

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Cell-free assays can also be used to measure binding of DABP or SABP polypeptides to isolated Duffy antigen or glycophorin polypeptides. For instance, the erythrocyte proteins can be immobilized on a solid surface and binding of labelled SABP or DABP polypeptides can be measured.

Many assay formats employ labelled assay components. The labelling systems can be in a variety of forms. The label may be coupled directly or indirectly to the desired component of the assay according to methods well known in the art. A wide variety of labels may be used. The component may be labelled by any one of several methods. The most common method of detection is the use of autoradiography with ³H, ¹²⁵l, ³⁵S, ¹⁴C, or ³²P labelled compounds or the like. Non-radioactive labels include ligands which bind to labelled antibodies, fluorophores, chemiluminescent agents, enzymes, and antibodies which can serve as specific binding pair members for a labelled ligand. The choice of label depends on sensitivity required, ease of conjugation with the compound, stability requirements, and available instrumentation.

In addition, the polypeptides of the invention can be assayed using animal models, well known to those of skill in the art. For P falciparum the in vivo models include Actus sp. monkeys or chimpanzees; for P. vivax the in vivo models include Saimiri monkeys.

In the case of the use nucleic acids for diagnostic purposes, standard nucleic hybridization techniques can be used to detect the presence of the genes identified here (e.g., members of the DBL family). If desired, nucleic acids in the sample may first be amplified using standard procedures such as PCR. Diagnostic kits comprising the appropriate primers and probes can also be prepared.

. Н. **DBL** Targeted Therepeutics

DBL polypeptides are expressed on the surface of Plasmodium-infected erythrocytes. As such, they

present ideal targets for therepeutics which target infected erythrocytes. In one preferred embodiement of the present invention, cytotoxic antibodies or antibody fusion proteins with cytotoxic agents are targeted against DBL proteins, killing infected erythrocytes and inhibiting the reproduction of *Plasmodium* in an infected host.

The procedure for attaching a cytotoxic agent to an antibody will vary according to the chemical structure of the agent. Antibodies and cytotoxic agents are typically bound together chemically or, where the antibody and cytotoxic agents are both polypeptides, are optionally synthesized recombinantly as a fusion protein. Polypeptides typically contain variety of functional groups; e.g., carboxylic acid (COOH) or free amine (-NH₂) groups, which are available for reaction with a suitable functional group on either the antibody or the cytotoxic agent.

Alternatively, antibodies or cytotoxic agents are derivitized to attach additional reactive functional groups. The derivatization optionally involves attachment of linker molecules such as those available from Pierce Chemical Company, Rockford Illinois. A "linker", as used herein, is a molecule that is used to join the nucleic acid binding molecule to the receptor ligand. The linker is capable of forming covalent bonds to both the antibody and the cytotoxic agent. Suitable linkers are well known to those of skill in the art and include, but are not limited to, straight or branched-chain carbon linkers, heterocyclic carbon linkers, or peptide linkers. Where the antibody and the cytotoxic agent are polypeptides, the linkers are joined to the constituent amino acids through their side groups (e.g., through a disulfide linkage to cysteine) or to the alpha carbon amino and carboxyl groups of the terminal amino acids.

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A bifunctional linker having one functional group reactive with a group on a particular ligand, and another group reactive with a nucleic acid binding molecule, can be used to form the desired conjugate. Alternatively, derivatization can proceed through chemical treatment of the ligand or nucleic acid binding molecule, e.g., glycol cleavage of the sugar moiety of a glycoprotein with periodate to generate free aldehyde groups. The free aldehyde groups on the glycoprotein may be reacted with free amine or hydrazine groups on an agent to bind the agent thereto (See, e.g., U.S. Patent No. 4,671,958). Procedures for generation of free sulfhydryl groups on polypeptides, are known (See, e.g., U.S. Pat. No. 4,659,839).

Many procedures and linker molecules for attachment of various compounds to proteins are known. See, for example, European Patent Application No. 188,256; U.S. Patent Nos. 4,671,958, 4,659,839, 4,414,148, 4,699,784; 4,680,338; 4,569,789; and 4,589,071; and Borlinghaus *et al. Cancer Res.* 47: 4071-4075 (1987). In particular, production of various antibody conjugates is well-known within the art and can be found, for example in Thorpe *et al., Monoclonal Antibodies in Clinical Medicine,* Academic Press, pp. 168-190 (1982), Waldmann, *Science*, 252: 1657 (1991), and U.S. Patent Nos. 4,545,985 and 4,894,443.

A number of antibodies which bind cell surface receptors have been converted to form suitable for incorporation into fusion proteins, and similar strategies are used to create fusion-protein antibodies which bind DBR polypeptides. see Batra et al., Mol. Cell. Biol., 11: 2200-2205 (1991); Batra et al., Proc. Natl. Acad. Sci. USA, 89: 5867-5871 (1992); Brinkmann, et al. Proc. Natl. Acad. Sci. USA, 88: 8616-8620 (1991); Brinkmann et al., Proc. Natl. Acad. Sci. USA, 87: 1066-1070 (1990); Friedman et al., Cancer Res. 53: 334-339 (1993); Kreitman et al., J. Immunol., 149: 2810-2815 (1992); Nicholls et al., J. Biol. Chem., 268: 5302-5308 (1993); and Wells, et al., Cancer Res., 52: 6310-6317 (1992), respectively).

B. Production of Fusion Proteins

Where the antibody fragment and/or the cytotoxic agents are relatively short polypeptides (i.e., less than about 50 amino acids) they are often synthesized using standard chemical peptide synthesis techniques. Where both molecules are relatively short, a chimeric molecule is optionally synthesized as a single contiguous polypeptide. Alternatively, the ligand and the nucleic acid binding molecule can be synthesized separately and then fused chemically.

Solid phase synthesis in which the C-terminal amino acid of the sequence is attached to an insoluble support followed by sequential addition of the remaining amino acids in the sequence is a preferred method for the chemical synthesis of the ligands of this invention. Techniques for solid phase synthesis are described by Barany and Merrifield, Solid-Phase Peptide Synthesis; pp. 3-284 in The Peptides: Analysis, Synthesis, Biology. Vol. 2: Special Methods in Peptide Synthesis, Part A., Merrifield, et al., J. Am. Chem. Soc., 85: 2149-2156 (1963), and Stewart et al., Solid Phase Peptide Synthesis, 2nd ed. Pierce Chem. Co., Rockford, Ill. (1984).

In a preferred embodiment, the fusion molecules of the invention are synthesized using recombinant nucleic acid methodology. Generally this involves creating a nucleic acid sequence that encodes the receptor-targeted fusion molecule, placing the nucleic acid in an expression cassette under the control of a particular promoter, expressing the protein in a host, isolating the expressed protein and, if required, renaturing the protein. Techniques

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sufficient to guide one of skill through such procedures are found in, e.g., Berger, Sambrook, Ausubel, Innis, and Freshney (all supra).

While the two molecules are often joined directly together, one of skill will appreciate that the molecules may be separated by a peptide spacer consisting of one or more amino acids. Generally the spacer will have no specific biological activity other than to join the proteins or to preserve some minimum distance or other spatial relationship between them. However, the constituent amino acids of the spacer may be selected to influence some property of the molecule such as the folding, net charge, or hydrophobicity.

Once expressed, recombinant fusion proteins can be purified according to standard procedures, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like (see, generally, R. Scopes, *Protein Purification*, Springer-Verlag, N.Y. (1982), Deutscher, *Methods in Enzymology Vol.* 182: Guide to Protein Purification., Academic Press, Inc. N.Y. (1990)). Substantially pure compositions of about 50 to 95% homogeneity are preferred, and 80 to 95% or greater homogeneity are most preferred for use as therepeutic agents.

One of skill in the art will recognize that after chemical synthesis, biological expression, or purification, the fusion molecule may possess a conformation substantially different than the native conformations of the constituent polypeptides. In this case, it is often necessary to denature and reduce the polypeptide and then to cause the polypeptide to re-fold into the preferred conformation. Methods of reducing and denaturing proteins and inducing re-folding are well known to those of skill in the art (See, Debinski *et al. J. Biol. Chem.*, 268: 14065-14070 (1993); Kreitman and Pastan, *Bioconjug. Chem.*, 4: 581-585 (1993); and Buchner, *et al., Anal. Biochem.*, 205: 263-270 (1992).

I. Pharmaceutical compositions comprising binding domain polypeptides

The polypeptides of the invention are useful in therapeutic and prophylactic applications for the treatment of malaria. Pharmaceutical compositions of the invention are suitable for use in a variety of drug delivery systems. Suitable formulations for use in the present invention are found in *Remington's Pharmaceutical Sciences*, Mack Publishing Company, Philadelphia, PA, 17th ed. (1985). For a brief review of methods for drug delivery, see, Langer, *Science* 249:1 527-1533 (1990).

The polypeptides of the present invention can be used in pharmaceutical and vaccine compositions that are useful for administration to mammals, particularly humans. The polypeptides can be administered together in certain circumstances, e.g. where infection by both P. falciparum and P. vivax is likely. Thus, a single pharmaceutical composition can be used for the treatment or prophylaxis of malaria caused by both parasites.

The compositions are suitable for single administrations or a series of administrations. When given as a series, inoculations subsequent to the initial administration are given to boost the immune response and are typically referred to as booster inoculations.

The pharmaceutical compositions of the invention are intended for parenteral, topical, oral or local administration. Preferably, the pharmaceutical compositions are administered parenterally, e.g., intravenously, subcutaneously, intradermally, or intramuscularly. Thus, the invention provides compositions for parenteral

administration that comprise a solution of the agents described above dissolved or suspended in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers may be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be sterilized by conventional, well known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or hyophilized, the hyophilized preparation being combined with a sterile solution prior to administration. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions, such as pH adjusting and buffering agents, tonicity adjusting agents, wetting agents and the like, for example, sodium acetate, sodium lactate, sodium chloride, potassium chloride, calcium chloride, sorbitan monolaurate, triethanolamine oleate, etc.

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For solid compositions, conventional nontoxic solid carriers may be used which include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talcum, cellulose, glucose, sucrose, magnesium carbonate, and the like. For oral administration, a pharmaceutically acceptable nontoxic composition is formed by incorporating any of the normally employed excipients, such as those carriers previously listed, and generally 10-95% of active ingredient and more preferably at a concentration of 25%-75%.

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For aerosol administration, the polypeptides are preferably supplied in finely divided form along with a surfactant and propellant. The surfactant must, of course, be nontoxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed. A carrier can also be included, as desired, as with, e.g., lecithin for intranasal delivery.

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In certain embodiments patients with malaria may be treated with SABP or DABP polypeptides or other specific blocking agents (e.g. monoclonal antibodies) that prevent binding of *Plasmodium* merozoites and schizonts to the erythrocyte surface.

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The amount administered to the patient will vary depending upon what is being administered, the state of the patient and the manner of administration. In therapeutic applications, compositions are administered to a patient already suffering from malaria in an amount sufficient to inhibit spread of the parasite through erythrocytes and thus cure or at least partially arrest the symptoms of the disease and its complications. An amount adequate to accomplish this is defined as "therapeutically effective dose." Amounts effective for this use will depend on the severity of the disease, the particular composition, and the weight and general state of the patient. Generally, the dose will be in the range of about 1mg to about 5gm per day, preferably about 100 mg per day, for a 70 kg patient.

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Alternatively, the polypeptides of the invention can be used prophylactically as vaccines. The vaccines of the invention contain as an active ingredient an immunogenically effective amount of the binding domain polypeptide or of a recombinant virus as described herein. The immune response may include the generation of antibodies; activation of cytotoxic T lymphocytes (CTL) against cells presenting peptides derived from the peptides encoded by the SABP, DABP or DBL sequences of the present invention, or other mechanisms well known in the art.

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See e.g. Paul Fundamental Immunology, Second Edition (Raven Press, New York, NY) for a description of immune response. Useful carriers are well known in the art, and include, for example, thyroglobulin, albumins such as human serum albumin, tetanus toxoid, polyamino acids such as poly(D-lysine:D-glutamic acid), influenza, hepatitis B virus core protein, hepatitis B virus recombinant vaccine. The vaccines can also contain a physiologically tolerable (acceptable) diluent such as water, phosphate buffered saline, or saline, and further typically include an adjuvant. Adjuvants such as incomplete Freund's adjuvant, aluminum phosphate, aluminum hydroxide, or alum are materials well known in the art.

The DNA or RNA encoding the SABP or DABP binding domains and the DBL gene family motifs may be introduced into patients to obtain an immune response to the polypeptides which the nucleic acid encodes. Wolff et. al., *Science* 247: 1465-1468 (1990) which is describes the use of nucleic acids to produce expression of the genes which the nucleic acids encode.

Vaccine compositions containing the polypeptides, nucleic acids or viruses of the invention are administered to a patient to elicit a protective immune response against the polypeptide. A "protective immune response" is one which prevents or inhibits the spread of the parasite through erythrocytes and thus at least partially prevent the symptoms of the disease and its complications. An amount sufficient to accomplish this is defined as an "immunogenically effective dose." Amounts effective for this use will depend on the composition, the manner of administration, the weight and general state of health of the patient, and the judgment of the prescribing physician. For peptide compositions, the general range for the initial immunization (that is for therapeutic or prophylactic administration) is from about $100 \mu g$ to about 1 gm of peptide for a 70 kg patient, followed by boosting dosages of from about $100 \mu g$ to about 1 gm of the polypeptide pursuant to a boosting regimen over weeks to months depending upon the patient's response and condition e.g. by measuring levels of parasite in the patient's blood. For nucleic acids, typically 30-1000ug of nucleic acid is injected into a 70kg patient, more typically about 50-150ug of nucleic acid is injected into a 70kg patient followed by boosting doses as appropriate.

The following examples illustrate preferred embodiments of the invention.

EXAMPLE 1: <u>Identification of the amino-terminal, cysteine-rich region of SABP and DABP as binding domains for crythrocytes</u>

1. Expression of the SABP binding domain polypeptide on the surface of Cos cells.

To demonstrate that the amino-terminal, cysteine-rich region of the SABP protein is the sialic acid binding region, this region of the protein was expressed on the surface of mammalian Cos cells *in vitro*. This DNA sequence is from position 1 to position 1848 of the SABP DNA sequence (SEQ ID No 3). Polymerase chain reaction technology (PCR) was used to amplify this region of the SABP DNA directly from the cloned gene.

Sequences corresponding to restriction endonuclease sites for Pvull or Apal were incorporated into the oligonucleotide sequence of the probes used in PCR amplification in order to facilitate insertion of the PCR amplified regions into the pHE4 vector (see below). The specific oligonucleotides, 5'-ATCGATCAGCTGGGAAGAATACTTCATCT-3'(SEQID NO:17) and 5'-ATCGATGGGCCCCGAAGTTTGTTCATTATT-3'

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(SEQ ID NO:18) were synthesized. These oligonucleotides were used as primers to PCR-amplify the region of the DNA sequence encoding the cysteine-rich amino terminal region of the SABP protein.

PCR conditions were based on the standard described in Saiki, et al., Science 239: 487-491 (1988). Template DNA was provided from cloned fragments of the gene encoding SABP which had been spliced and re-cloned as a single open-reading frame piece.

The vector, pRE4, used for expression in Cos cells is shown in Figure 2. The vector has an SV40 origin of replication, an ampicillin resistance marker and the Herpes simplex virus glycoprotein D gene (HSV glyd) cloned downstream of the Rous sarcoma virus long terminal repeats (RSV LTR). Part of the extracellular domain of the HSV glyd gene was excised using the Pvull and Apal sites in HSV glyd.

As described above, the PCR oligonucleotide primers contained the Pvull or Apal restriction sites. The PCR-amplified DNA fragments obtained above were digested with the restriction enzymes Pvull and Apal and cloned into the Pvull and Apal sites of the vector pRE4. These constructs were designed to express regions of the SABP protein as chimeric proteins with the signal sequence of HSV glyd at the N-terminal end and the transmembrane and cytoplasmic domain of HSV glyd at the C-terminal end. The signal sequence of HSV glyd targets these chimeric proteins to the surface of Cos cells and the transmembrane segment of HSV glyd anchors these chimeric proteins to the Cos cell surface.

Mammalian Cos cells were transfected with the pRE4 constructs containing the PCR-amplified SABP DNA regions, by calcium phosphate precipitation according to standard techniques.

2. Expression of the DABP binding domain polypeptide on the surface of Cos cells.

To demonstrate that the amino-terminal, cysteine-rich region of the DABP protein is the binding domain, this region was expressed on the surface of Cos cells. This region of the DNA sequence from position 1-975 was first PCR-amplified (SEQ ID No 1).

Sequences corresponding to restriction endonuclease sites for Pvull or Apal were incorporated into the oligonucleotide probes used for PCR amplification in order to facilitate subsequent insertion of the amplified DNA into the pRE4 vector, as described above. The oligonucleotides, 5'-TCTCGTCAGCTGACGATCTCTAGTGCTATT-3' (SEQ ID NO:19) and 5'-ACGAGTGGCCCTGTCACAACTTCCTGAGT-3' (SEQ ID NO:20) were synthesized. These oligonucleotides were used as primers to amplify the region of the DABP DNA sequence encoding the cysteine-rich, amino-terminal region of the DABP protein directly from the cloned DABP gene, using the same conditions described above.

The same pRE4 vector described above in the section on expression of SABP regions in Cos cells was also used as a vector for the DABP DNA regions.

3. Binding studies with erythrocytes.

To demonstrate their ability to bind human erythrocytes, the transfected Cos cells expressing binding domains from DABP and SABP were incubated with erythrocytes for two hours at 37°C in culture media (DMEM/10% FBS). The non-adherent erythrocytes were removed with five washes of phosphate-buffered saline and the bound erythrocytes were observed by light microscopy. Cos cells expressing the amino terminal, cysteine-rich

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SABP polypeptides on their surface bound untreated human erythrocytes, but did not bind neuraminidase treated erythrocytes, that is, erythrocytes which lack sialic acid residues on their surface. Cos cells expressing other regions of the SABP protein on their surface did not bind human erythrocytes. These results identified the amino-terminal, cysteine-rich region of SABP as the erythrocyte binding domain and-indicated that the binding of Cos cells expressing these regions to human erythrocytes is specific. Furthermore, the binding of the expressed region to erythrocytes is identical to the binding pattern seen for the authentic SABP- 175 molecule upon binding to erythrocytes.

Similarly, Cos cells expressing the amino-terminal cysteine-rich region of DABP on their surface bound Duffy-positive human erythrocytes, but did not bind Duffy-negative human erythrocytes, that is erythrocytes which lack the Duffy blood group antigen. Cos cells expressing other regions of the DABP protein on their surface did not bind human erythrocytes. These results identified the amino-terminal cysteine rich region of DABP as the erythrocyte binding domain and indicated that the binding of the Cos cells was specific.

EXAMPLE 2: <u>Isolation of polynucleotide sequences in the DBL family</u>

P. falciparum clones and cell line used include the following. P. falciparum clones 3D7, D10, LF4/1, Camp/A1, SL/D6, HB3, 7G8, V1/S, T2/C6, KMWII, ItG2F6, FCR3/A2 and Dd2 have been previously tabulated (Dolan, et al. (1993), Mol. Biochem. Parasitol. 61, 137-142). Line Dd2/NM1 was selected from clone Dd2 for invasion via a sialic acid-independent pathway (Dolan, et al. (1990), J. Clin. Invest. 86, 618-624). All parasites were maintained in vitro by standard methods (Trager, et al. (1976), Science 193, 673-675).

DNA and RNA Isolation and Analysis. DNA was extracted as described (Peterson, et al. (1990), Proc. Natl. Acad. Sci. USA 87, 3018-3022). Endonuclease digestion, agarose gel electrophoresis, and filter hybridizations were performed by standard methods (Sambrook, et al., 1989). All hybridizations were at 56°C (Sambrook, et al., 1989). Blots were washed for 2 min. at room temperature in 2x standard saline/phosphate/EDTA (SSPE) with 0.5% SDS, followed by two higher stringency washes at 50°C in 0.3xSSPE with 0.5% SDS. Parasite chromosomes were embedded in agarose blocks and separated by pulsed field gel electrophoresis (Dolan, et al. (1993), Methods. Mol. Biol. 21, 319-332). RNA was isolated from cultured parasites by LiC1 extraction of Catrimox-14-precipitated RNA (Dahle, et al. (1993), BioTechniques 15, 1102-1105). Agarose gel electrophoresis of total RNA and filter hybridizations were performed by standard methods (Sambrook, et al., (1989).

Oligonucleotide Primers and PCR. Primers specific for E31a used in a RT-PCR to test for expression of this sequence were E31aT2 (5'-AGA-CCT-CAA-TTT-CTA-AG-3') (SEQ ID NO:21) and E31aRev1 (5'-AAT-CGC-GAG-CAT-CAT-CTG-3') (SEQ ID NO:22).

Two primers were used to amplify additional sequences from genes encoding *DBL* domains. These were designed from conserved amino acids encoded in the *DBL* domain of the eba-175 and E31a sequences. After adaptation to incorporate the most frequently-used *P. falciparum* codons, forward primer UNIEBP5' [5'-CC(A/G)-AG(G/A)-CAA-(G/A)AA-(C/T)TA-TG-3'] (SEQ ID NO:23), based upon the amino acid sequence PRHUKLC, and reverse primer UNIEBP3' [5'-CCA-(A/T)C(T/G)-(T/G)A(A/G)-(A/G)AA-TTG-(A/T)GG-3'] (SEQ ID NO:24), based upon the amino acid sequence PQFLRW, were synthesized.

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RT-PCR amplifications were performed as described (Kawasaki, et al. (1990), PCR Protocols, A Guide to Methods and Applications, eds. Innis, M.A., Gelfand, D.H., Sninsku, J.J. & White, T.J. (Academic, San Diego), pp. 21-27). In brief, 0.5 to 1 mg of total RNA was treated with RQ1 DNAse (Promega), phenol/chloroform extracted, and ethanol precipitated. The RNA was then annealed with random oligonucleotide primers and extended with Superscript reverse transcriptase (GIBCO/BRL). PCR cycling conditions were 94°C for 10 sec, 45°C for 15 sec, and 72°C for 45 sec, for 30 cycles. All PCRs were performed in an Idaho Technology air thermal cycler using buffer containing 2 mM Mg2+.

PCR amplification products were separated by use of PCR Purity Plus gels and protocols (AT Biochem. Malvern. PA).

DNA Clones and Hybridization Probes. Clone pE31a was isolated from a genomic library prepared from the region of chromosome 7 linked to chloroquine resistance Walker-Jonah, et al. (1992), Mol. Biochem. Parasitol. 51, 313-320. Clone pS31H (GenBank accession no. L38454), containing an insert encompassing that of pE31a, was cloned from a size-selected Hind III restriction digest of Dd2 genomic DNA.

Clone pEBLe1 was cloned from a RT-PCR of Dd2 cDNA after amplification with primers UNIEBP5' (SEQ ID NO:23) and UNIEBP3' (SEQ ID NO:24). Clone pEBP1.2 (GenBank accession no. L38450), containing an insert encompassing that of pEBLe1, was isolated from a Dd2 cDNA library probed with pEBLe1. *DBL*-encoding sequences of *dbl-nm1-4* (GenBank accession no. L38455) and *dbl-nm1-5* (GenBank accession no. L38453) were amplified by RT-PCR from first strand cDNA of line Dd2/NM using primers UNIEBP5' and UNIEBP3'. Sequencing was performed on double stranded DNA templates by standard protocols for the dideoxynucleotide method. (Sequenase; U.S. Biochemicals).

Sequences related to the E31a sequence were detected with the 3005 bp insert of clone pS31H. The eba-175 gene was detected with a PCR amplified probe consisting of the first 1825 bp of the coding sequence. ebl-1 sequences were detected with the 2098 bp insert of clone pEBP1.2. All probes were comparable in organization, each containing a region encoding at least one DBL domain and varying amounts of flanking sequence.

Homology searches and alignments. Homology searches were performed with BLAST and the Genetics Computer Group program FASTA (Altschul, et al. (1990), J. Mol. Biol. 215, 403-410; Devereux, et al. (1984), Nucleic Acids. Res. 12(1 Pt 1, 387-395). Optimized alignments were produced with MACAW sequence alignment software (Schuler, et al. (1991), Proteins. 9, 180-190).

Multiple P. falciparum sequences encode DBL domains. Positional cloning experiments directed to P. falciparum chromosome 7 identified an ORF (E31a) encoding a DBL domain that is homologous to the domains found in the P. vivax and P. knowlesi DABPs and the P. falciparum SABP. Figure 4 shows the realtive position of the E31a ORF on chromosome 7.

The homology between the *DBL* domains of E31a and the erythrocyte-binding proteins is due to the presence of short motifs of highly conserved amino acids. These well-conserved stretches are separated by non-homologous sequences and by deletions and insertions that vary the size of the domain by greater than 60 aa. The typical *DBL* domain contains 12 or more cysteine residues and has 7 conserved tryptophan residues. Additional

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well conserved amino acids include 4 arginines, 3 aspartates, 9 positions with aliphatic residues (alanine, isoleucine, leucine, or valine) and 4 with aromatic amino acids (tryptophan, phenylalanine, or tyrosine).

Probes spanning the sequence that encodes the E31a *DBL* domain hybridized to multiple fragments within a single restriction digest and yielded bands that varied among parasite lines. The numerous distinct bands from a selection of different parasite DNAs indicated a large number of diverse but related elements. These multiple bands varied among different *P. falciparum* clones, in contrast to the well-conserved, single-copy signal obtained with the *eba-175* probe.

Because of the numerous cross-hybridizing sequences, it seemed likely that many of these related sequences would be on different chromosomes of the parasite. PFG electrophoresis of *P. falciparum* Dd2 chromosomes and hybridization with the E31a probe identified a number of cross-hybridizing sequences on multiple chromosomes. A control hybridization with the *eba-175* probe under identical conditions yielded a single band of hybridization from chromosome 7.

RNA Analysis of *DBL* Elements. Sequences from E31a (pS31H insert) were used to probe RNA blots for corresponding transcripts. No hybridization was detected. Because it was still possible that a message of low abundance was not being detected on the RNA blot, RT-PCR was used as a means of more sensitive detection. For this purpose, cDNA was generated by RT from random primers annealed to DNAse-treated total RNA. E31a-specific oligonucleotides were then used to test for amplification from the cDNA. No amplification of the E31a sequence was obtained, while genomic DNA controls and amplification from cDNA by dihydrofolate reductase/thymidylate synthetase-specific primers yielded the expected bands. A screen of a cDNA library with E31a specific probes also failed to detect any clones hybridizing with the ORF. These results indicate that E31a is either a pseudogene, or is expressed in parasite strains or stages not examined in this work.

A PCR Method to Isolate Sequences Encoding DBL Domains. The identification of short conserved motifs in DBL domains that otherwise have extreme diversity led to a PCR strategy using degenerate oligonucleotide primers designed from conserved amino acid sequences in the DBL domains. Sequences PRRQKLC and PQFLRW were judged most suitable for minimizing degeneracy while allowing amplification of expressed DBL sequences. After these considerations and adjustment for P. falciparum codon usage, primers UNIEBP5' and UNIEBP3' were synthesized.

While some *P. falciparum* lines yielded similar patterns of amplified bands (e. g. Dd2 and MCamp; FCR3/A2 and K-1), no two separate isolates showed identical patterns, reflecting the diversity of the *DBL* domains in the parasite lines. A few bands of the same apparent size were present in many isolates. These included a consistent 490 bp product that was determined to be the *eba-175* gene by its expected size and hybridization to a gene-specific probe. The number of discernible bands probably underestimates the number of amplifiable sequences because of overlapping products of the same size and possible preferential amplification of some sequences over others. Nevertheless, the parasite-specific patterns in the amplified bands may provide a means to quickly type isolates and serves as a measure of parasite diversity in field samples.

To identify *DBL*-encoding sequences in RNA transcripts, the UNIEBP primers were used to amplify first-strand cDNAs generated from DNAse-treated RNA preparations. Amplified products from Dd2, 3D7, HB3 and MCAMP cDNAs had diverse sizes ranging from 400 bp to nearly 1 kb. These included a band at 480-500 bp that was determined to be *eba-175* from its expected size and cross-hybridization to an *eba-175*-specific probe. Other bands were from amplification of different transcripts encoding *DBL* domains. Dd2-NM1 RNA, for example, yielded bands above the *eba-175* product that included two related sequences (*dbl-nm1-4,dbl-nm1-5*). These bands were found to be isolate-specific and to have features consistent with the *var* genes described in Example 3, below. Probes that detect *dbl-nm1-4* and *dbl-nm1-5* hybridized to multiple chromosomes and aligned more closely with E31a than with EBA-175 or DABP.

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The RT-PCR amplifications also yielded a consistent band that encoded a novel *DBL* domain distinct from *eba-175*. A cDNA clone corresponding to this product was isolated by screening a *Agt10 Dd2 cDNA* library with a radiolabeled *ebl-1* probe. Sequence from this and additional overlapping cDNA clones confirmed the conserved motifs of the *DBL* domain. The alignment of the predicted amino acid sequences showed that the *DBL* domain of *ebl-1* is more similar to *eba-175* than to the multicopy genes. There was, however, extensive divergence from *eba-175* and other known genes outside of the amplified region.

In contrast to the multicopy hybridization patterns of dbl-nm1-4 and dbl-nm1-5, the ebl-1 sequence,

like that of *eba-175*, was found to have hybridization patterns consistent with a conserved single-copy gene. Probes specific for *ebl-1* hybridized only to chromosome 13, and restriction analysis with the enzymes *Cla I, EcoRI, HindIII, Hinf I, Nsi I, Rsa I*, and *Spe I*, all yielded bands expected from a single copy sequence. RNA blots probed with *ebl-1*-specific sequences showed several bands of hybridization, however, corresponding to 8-9.5 kb transcripts in mRNA from the Dd2 and 3D7 parasites. The transcripts of different size may result from alternative start and

termination points or from incompletely processed species containing introns.

EXAMPLE 3: Isolation of var genes

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Parasite clones, DNA analysis and Chromosome Mapping. Parasite clones were cultivated by the methods of (Trager, et al. (1976), Science 193, 673-675). DNA was extracted from parasite cultures as described (Peterson, et al. (1988), Proc. Natl. Acad. Sci. USA 85, 9114-9118) except that the DNA was as recoverd by ethanol precipitation rather than spooling. Fingerprint analysis with the pC4.H32 probe was used to confirm DNA preparations (Dolan, et al. (1993), Mol. Biochem. Parasitol. 61, 137-142). Southern blotting to Nytran membranes was recommended by the manufacturer (Schleicher & Schuell, Keene, NH). PFG separation of the 14 P. falciparum chromosomes and chromosome mapping were performed as described (Wellems, et al. (1987), Cell 49, 633-642; Sinnis, et al. (1988): Genomics 3, 287-295).

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RNA isolation. Parasites from 200 ml mixed stage cultures (5-10% parasitemia) were released by saponin lysis as for DNA preparations except that the procedures were performed with ice-cold solutions. RNA was immediately isolated from the parasite pellet by guanidine thiocyanate/phenol-chloroform methods, recovered and treated with RNAase-free DNAse (Creedon, et al. (1994), J. Biol. Chem. 269, 16364-16370. RNA in H₂0 was combined with 2 vol 100% ETOH, distributed into 2 ml vials and frozen as stock at -70°C. RNA was recovered by

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precipitation with 0.1 vol 3M NaOAc. RNA blots were generated and probed as described (Creedon, et al. (1994), J. Biol. Chem. 269, 16364-16370).

YAC isolation, chromosome-segment libraries and cDNA libraries. Overlapping YACs spanning the 300 kb segment of chromosome 7 that contains the CQR locus were obtained from a YAC library of a CQR FCR3 parasite line de Bruin, et al. (1992), Genomics 14, 332-339) by the procedures of Lanzer, et al. (1993), Nature 361, 654-657. Orientation of the YACs and their overlaps were identified with probes obtained from the YAC ends by inverted PCR.

Attempts to construct cosmid libraries and large insert (~10 kb) A libraries from high molecular weight *P. falciparum* genomic DNA yielded only rearranged clones. An alternative approach was therefore taken in which chromosome-segment libraries were constructed that contained small (0.5-5 kb) inserts in plasmid vectors. Plasmid libraries containing *Alul*, *Hinfl*, *Rsal* and *Sspl* inserts in pCDNAII were constructed from Dd2 chromosome 7 restriction fragments purified by pulsed-field gel (PFG) electrophoresis (Wellems, *et al.* (1991), *Proc. Natl. Acad. Sci. USA* 88, 3382-3386). A plasmid library from a 34 kb *Apal-Smal* restriction fragment of YAC PfYED9 was constructed by the same methods. Inserts in the plasmid libraries were generally 0.5-4 kb.

The Agt10 Dd2 cDNA library was prepared under contract by CloneTech Laboratories Inc. (Palo Alto, CA) from the DNAse-treated, polyA+ fraction of Dd2 RNA. The cDNA was generated in two separate reactions using oligodT primers or random primers. Products of these reactions were combined, processed and cloned into the EcoRI site of Agt10. 1.6 x 10⁶ independent recombinants were obtained and amplified.

Isolation of overlapping clones and DNA sequencing. Plasmid clones from the chromosome-segment and YAC-segment libraries were picked at random and their locations were established by restriction mapping. After sequence data from these clones were generated, overlapping clones were isolated in a process of "chromosome walking" by rescreening the libraries with oligonucleotide probes near the ends of sequenced inserts. Sufficient divergence was present among repetitive elements in the sequences to allow distinction of clones and unambiguous assignment of overlaps (generally 50-200 bp).

Sequencing reactions with single-strand M13 DNA (1 μ g) and double-strand plasmid DNA (2-5 μ g) were performed in 96-well polyvinyl chloride U-bottom microassay plates using a Sequenase protocol recommended by United States Biochemical Corp. (Cleveland, OH). Reactions were separated by 8M urea-6% polyacrylamide sequencing gels and exposed to Kodak BioMax MR film. Sequence data from some clones were also obtained by use of an ABI 373A automated DNA sequencer (Applied Biosystems Inc., Foster City, CA). Cycle sequencing reactions were performed using the ABI PRISM DyeDeoxy system.

DNA sequence editing, analyses and display were performed with MacVector software (International Biotechnologies Inc., New Haven, CT), BLAST (Altschul, et al. (1990), J. Mol. Biol. 215, 403-410), Genetics Computer Group programs (Devereux, et al. (1984), Nucleic Acids Res. 12, 387-395) and the DNADRAW package (Shapiro, et al. (1986), Nucleic Acids Res. 14, 65-73) maintained at the National Institutes of Health.

resistance. Four overlapping yeast artificial chromosomes from the *P. falciparum* FCR3 line were obtained that span the 300 kb chromosome segment linked to CQR, a segment located 300-600 kb from the telomere of chromosome

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7. Figure 5 shows the positions of these YACs (PfYEF2, PfYFE6, PfYKF8, PfYED9) relative to the chromosome map. In order to define the structure of this 300 kb segment, we performed comparative hybridizations to search for polymorphisms between parasite lines. Clones were randomly picked from chromosome segment-specific plasmid libraries and their inserts were hybridized against restriction digests of the YAC and parasite DNAS. Over thirty inserts were identified that recognized PfYEF2, PfYFE6 or PfYKF8 and showed a predonderance of single copy sequences with few polymorphisms (AbA, Hinfl, Rsal and Sspl digests), consistent with prior findings that chromosome internal regions are largely conserved and contain a preponderance of single copy sequences. However, fifteen other inserts that recognized PfYED9 showed highly polymorphic sets of repetitive elements in the parasite DNAs. Southern analysis indicated that these polymorphic elements were part of a chromosome hypervariable region contained within the PfYED9 clone.

Mapping and DNA sequencing of the hypervariable region spanned by YAC PfYED9. Single copy sequences detected by pE45b and pH270.5 flank the hypervariable region spanned by PfYED9 (Figure 5). The pE45b and pH270.5 probes were therefore used to assign large restriction fragments on the PfYED9 map and establish enzyme recognition sites as reference points. A detailed restriction map of the PfYED9 hypervariable region was then developed. Fifteen overlapping clones ("a"·"f' and "h"·"o" in Figure 5) were isolated by a chromosome walking approach from Dd2 chromosome subsegment libraries (Wellems *et al.*, *supra*) The inserts yielded 19.1 kb of continuous Dd2 sequence having predicted enzyme recognition sites in perfect accord with the PfYED9 restriction map. Such agreement indicates that the Dd2 and FCR3 sequences in this part of the chromosome are very similar, despite differences elsewhere in the genome that are evident by restriction analysis.

We also obtained genomic sequence data from the 34 kb *Apal-Smal* fragment of PfYED9. Purified PfYED9 DNA was cut with *Smal* to yield a 110 kb fragment, which was then isolated by PFG electrophoresis and digested with *Apal*. The resulting 34 kb *Apal-Smal* band was purified by PFG electrophoresis, digested in four separate reactions by *Alul*, *Hinfl*, *Rsal* or *Sspl* and incorporated into a plasmid (PCDNAII) library. Cloned inserts from the library were checked for hybridization to the PfYED9 34 kb fragment, assigned to the PfYED9 map and sequenced (Figure 5). Overlapping inserts were obtained by the chromosome walking approach except for three gaps ("t", "z", " θ " in Figure 5) which were closed by PCR amplification of PfYED9 DNA using primers from flanking sequences. The clones from PfYED9 ("r"-"z"," γ ", " κ " and " α " + " β " in Figure 5) yielded 22.2 kb of continuous DNA sequence that overlaps the Dd2 sequence at the "f"|" β " junction and has predicted restriction sites that match the PfYED9 map perfectly. The composite sequence from the Dd2 and PfYED9 segments is 40,171 kb.

Structure of a var gene cluster and comparative analysis of predicted amino acid sequences. The 40,171 bp sequence contains three 10-12 kb regions that have related sequences and structure. Each of these regions harbors a pair of ORFS. The first ORF in each pair begins with a consensus ATG start codon preceded by typical P. falciparum non-coding sequence of abundant A+T content. The ORFs of each pair are separated by an intervening AT-rich and non-coding sequence of 0.9 kb to 1.1 kb. Presence of consensus intron-exon splice junction sequences at either end of these intervening sequences and lack of a consistent translation start site in the 3' ORF indicate that the each pair of ORFs belongs to an individual gene having a two exon structure. This has been verified by

comparison of the genomic sequences to the cDNA sequence of an expressed gene (var-7; see subsequent section). The three 10 kb to 12 kb regions thus contain members of a variant gene family which have coding regions of 9.23kb (var-1), 7.99 kb (var-2) and 9.01 kb (var-3). Predicted molecular weights of the encoded proteins are 350 kD, 302 kD and 344 kD. respectively.

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The var genes are flanked by additional members of the var family in PfYED9. Restriction analysis identified two additional genes that are 12-35 kb upstream of the sequenced region and are closely related to var-2 and var-3 (var-2c and Var-3c, Figure 5). The var genes thus have a clustered arrangement in which many individual members are organized in head-to-tail fashion. Between var-1 and var-2 is a 5 kb DNA sequence that harbors a short ORF homologous to that of a repetitive element (rij) suggested to be a transposable element in P. falciparum.

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The deduced protein sequences of the var genes are highly diverse, yet all contain certain conserved motifs and common structural features. Database searches identified 2 to 4 domains within each var sequence that are homologous to cysteine-rich domains of SABP and DABP. In the var sequences, the first domain near the amino-terminus (DBL domain 1) is the most conserved of the DBL domains and has amino acid signatures that differentiate it from subsequent domains (e.g. consensus peptide sequences GAcAp[Y/F]rrL, CTxLARsfadlgdlVrgrdLYLG and VPTYFDYVpqylrwF). Between DBL domains 1 and 2 is another type of conserved domain, a cysteine-rich interdomain region (CIDR) of 300-400 amino acids. The CIDR does not have all the motifs of a DBL domain, but it does have a region at the 3'end which is homologous to the end of the FI DBL domain in SABP. The conservation evident in the sequences of DBL domain I and the CIDR suggest that these regions maintain important structures in the head of the variant molecule.

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DBL domains 2, 3 and 4 (numbering is according to *var-1*, the first sequence completed) have less discriminating signatures than domain 1, and show features of cross-alignment and variation in number that suggest these domains can undergo shuffling and deletion.

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DBL domain 4 is followed by a segment of variable length and a hydrophobic region that is encoded at the end of the first exon (exon 1). In all var sequences this hydrophobic region fits the criteria of a transmembrane segment. The second exon (exon II) encodes a large (45-55 kD) conserved C-terminal sequence that has an acid character (predicted pl = 4.5, vs. 5.9 for the part of the protein upstream of the splice junction) and a cysteine content of < 1% (vs. > 4% upstream). The position of this C-terminal sequence downstream of a single transmembrane segment suggests that it has a cytoplasmic location.

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No consensus signal sequence was detected in the NH₂-terminal region of the predicted *var* ORFs. We note the presence of several motifs in the protein sequences that are known to act as ligands and receptors in the integrin family. These include RGD (*var-1* codons 886-88, 1992-94) and DGEA (*var-1* codons 2111-14). Not all of these motifs occur in each protein sequence and, when they do occur, their positions vary.

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Identification of var transcripts and chromosome expression sites. To identify transcribed var sequences we screened a Agt10 Dd2 cDNA library with var-containing BssHII restriction fragments that had been purified from PfYED9 and radiolabeled by random hexamer priming. This screening yielded 18 clones with inserts that hybridized back to PfYED9. By cross-hybridization studies and DNA sequence analysis the inserts fell into two groups: group

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I inserts that aligned with sequences of *var* exon I (AT240, AT242, AT244, AT284, AT287, AT288, AT295, AT296); and group II inserts that aligned with sequences of *var* exon II (AT140, AT141, AT142, AT145, AT147, AT148, AT150, AT152).

The full ORF of an expressed var gene (var-7) was determined from \(\lambda\)T242 and overlapping cDNA clones that were obtained by a PCR-based walking strategy. The sequence showed that \(\var-7\) has a 6.6 kb ORF containing two \(\textit{DBL}\) domains, a hydrophobic transmembrane sequence and carboxy-terminal region typical of \(\var\) genes (predicted molecular weight 249 kD). Comparison of \(\var-7\) with the \(\var-1\) sequence demonstrated continuity of the alignments at the predicted splice junction between the ORFs of exons I and II. PCR amplification of Dd2 genomic DNA was also performed with primers derived from the two \(\var-7\) exons. Sequence of this \(\var-7\) PCR product confirmed consensus splice sites and a 1 kb intron typical of the \(\var-7\) genes. Transcription of \(\var-7\) was detected as a 7.5 kb band by RNA blot analysis.

Chromosome mapping experiments with a var-7-specific probe localized the var-7 gene to a region that is 600 kb from one end of Dd2 chromosome 12 (chromosome 12 has a length of 2600 kb). No hybridization of the var-7 probe was detected to any other Dd2 chromosome nor to any chromosomes of the HB3, 3D7 or A4 parasites. Other cDNA inserts from the group I clones were also sequenced and examined for chromosome hybridization signals. The AT240 cDNA insert mapped to the var-1/var-2/var-3 cluster on Dd2 chromosome 7 and its sequence matched that of var-3. The AT244, AT284, AT287, AT288, AT295 and AT296 inserts all showed overlapping sequences and yielded the same hybridization patterns. Chromosome sites recognized by these inserts included regions within two Smal fragments from Dd2 chromosome 7 and another from chromosome 9. We note that loss of a cytoadherence phenotype has been correlated with a chromosome 9 deletion in certain P. falciparum lines.

1.8 kb to 2.4 kb RNA transcripts related to var exon II. In addition to the 7.5 kb var-7 band, a broad 1.8 kb to 2.4 kb band was detected on RNA blots after hybridization with a probe that recognizes var exon II. Sequences of eight group II cDNA inserts homologous to exon II were therefore determined and aligned against the var genes. Comparative analysis of the insert sequences showed that all differed from one another in regions of overlap, indicating that transcription of the corresponding RNAs was from different loci. Three of the cDNA sequences (AT140, AT141 and AT148) aligned downstream of the intron/exon II splice junction. However, five other cDNA inserts (AT142, AT145, AT147, AT150 and AT152) had sequences that aligned upstream of the var intron/exon II splice site and included regions homologous to var intron sequences. In the vicinity of the splice junction, consensus splice sites occurred in three of the cDNA sequences (AT142, AT147, AT150) while a fourth sequence (AT145) showed the required AG dinucleotide but not the expected pyrimidine tract of the splice consensus. The part of the fifth sequence (AT152) that aligned with the var intron extended upstream only to the TAG of the splice sequence. All five sequences lacked a consensus start codon preceded by A+T-rich non-coding DNA that is typical of P. falciparum translation start sites.

<u>Isolate-specific var sequences and evidence for DNA recombination in cultivated parasite clones.</u> The diversity of var forms expressed by *P. falciparum* parasites reflects a tremendous repertoire in the var gene family.

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This repertoire is evident in the patterns of restriction polymorphism detected by *var* probes as well as in the detection of *var*-specific sequences that hybridize to some parasite DNAs but not to others. The *var-7* gene expressed by Dd2, for example, is not present in the HB3, 3D7 or A4 genomes. Such *var* diversity suggests that frequent DNA rearrangements underlie the production of antigenically variant types in different parasite strains.

To test for DNA rearrangements in parasites cultivated *in vitro*, we used *var* sequences to probe restricted DNAs from Dd2 lines adapted to neuraminidase-treated erythrocytes. In one rearrangement a novel 35 kb *Bgl* fragment is seen in NM1 DNA probed with the AT142 (group II) insert. In another rearrangement a deletion of a 20 kb *Pst*1 band is evident in NM8 DNA probed with a *var-*7 sequence. Deletion of this 20 kb band was also detected in the Dd2/R8 subclone obtained before neuraminidase selection, indicating that the DNA rearrangement was not produced by selection in neuraminidase-treated erythrocytes.

The above examples are provided to illustrate the invention and other variants of the invention encompassed by the claims will be readily apparent to one of ordinary skill in the art.

-31-

SEQUENCE LISTING

		SEQUENCE LISTING
	(1) GENE	RAL INFORMATION:
5	(i) Secretar	APPLICANT: The United States, As Represented by the
10	(ii)	TITLE OF INVENTION: BINDING DOMAINS FROM PLASMODIUM VIVAX AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING PROTEINS
	(iii)	NUMBER OF SEQUENCES: 45
15	(iv)	CORRESPONDENCE ADDRESS: (A) ADDRESSEE: Knobbe Martens Olson & Bear (B) STREET: 620 Newport Center Drive 16th Floor (C) CITY: Newport Beach (D) STATE: California (E) COUNTRY: US (F) ZIP: 92660
20	(52)	COMPUTER READABLE FORM:
25	(*,	(A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
30	(vi)	CURRENT APPLICATION DATA: (A) APPLICATION NUMBER: (B) FILING DATE: (C) CLASSIFICATION:
35	(vii)	PRIOR APPLICATION DATA (A) APPLICATION NUMBER: US08/487826 (B) FILING DATE: 07-JUN-1996
33	(viii)	ATTORNEY/AGENT INFORMATION: (A) NAME: Israelsen, Ned (B) REGISTRATION NUMBER: 29,655 (C) REFERENCE/DOCKET NUMBER: NIH121.0010PC
40	(ix)	TELECOMMUNICATION INFORMATION: (A) TELEPHONE: (619) 235-8550 (B) TELEFAX: (619) 235-0176
45	(2) INFO	RMATION FOR SEQ ID NO:1:
50	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 4084 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
	(ii)	MOLECULE TYPE: DNA (genomic)
55	(iii)	HYPOTHETICAL: NO
	(vi)	ORIGINAL SOURCE: (A) ORGANISM: Plasmodium vivax
60	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:1:

AAGCTTTTAA AAATAGCAAC AAAATTTCGA AACATTGCCA CAAAAATTTT ATGTTTTACA 60
TATATTTAGA TTCATACAAT TTAGGTGTAC CCTGTTTTTT GATATATGCG CTTAAATTTT 120

TTTTTCGCTC ATATGTTTAG TTATATGTGT AGAACAACTT GCTGAATAAA TTACGTACAC 180 TTTCTGTTCT GAATAATATT ACCACATACA TTTAATTTTA AATACTATGA AAGGAAAAA 240 CCGCTCTTTA TTTGTTCTCC TAGTTTTATT ATTGTTACAC AAGGTATCAT ATAAGGATGA 300 TTTTTCTATC ACACTAATAA ATTATCATGA AGGAAAAAA TATTTAATTA TACTAAAAAG 360 5 AAAATTAGAA AAAGCTAATA ATCGTGATGT TTGCAATTTT TTTCTTCATT TCTCTCAGGT 420 AAATAATGTA TTATTAGAAC GAACAATTGA AACCCTTCTA GAATGCAAAA ATGAATATGT 480 GAAAGGTGAA AATGGTTATA AATTAGCTAA AGGACACCAC TGTGTTGAGG AAGATAACTT 540 AGAACGATGG TTACAAGGAA CCAATGAAAG AAGAAGTGAG GAAAATATAA AATATAAATA 600 TGGAGTAACG GAACTAAAAA TAAAGTATGC GCAAATGAAT GGAAAAAGAA GCAGCCGCAT 660 10 TTTGAAGGAA TCAATTTACG GGGCGCATAA CTTTGGAGGC AACAGTTACA TGGAGGGAAA 720 AGATGGAGGA GATAAAACTG GGGAGGAAAA AGATGGAGAA CATAAAACTG ATAGTAAAAC 780 TGATAACGGG AAAGGTGCAA ACAATTTGGT AATGTTAGAT TATGAGACAT CTAGCAATGG 840 CCAGCCAGCG GGAACCCTTG ATAATGTTCT TGAATTTGTG ACTGGGCATG AGGGAAATTC 900 TCGTAAAAAT TCCTCGAATG GTGGCAATCC TTACGATATT GATCATAAGA AAACGATCTC 960 TAGTGCTATT ATAAATCATG CTTTTCTTCA AAATACTGTA ATGAAAAACT GTAATTATAA 1020 15 GAGAAAACGT CGGGAAAGAG ATTGGGACTG TAACACTAAG AAGGATGTTT GTATACCAGA 1080 TCGAAGATAT CAATTATGTA TGAAGGAACT TACGAATTTG GTAAATAATA CAGACACAA 1140 TTTTCATAGG GATATAACAT TTCGAAAATT ATATTTGAAA AGGAAACTTA TTTATGATGC 1200 TGCAGTAGAG GGCGATTTAT TACTTAAGTT GAATAACTAC AGATATAACA AAGACTTTTG 1260 CAAGGATATA AGATGGAGTT TGGGAGATTT TGGAGATATA ATTATGGGAA CGGATATGGA 1320 20 AGGCATCGGA TATTCCAAAG TAGTGGAAAA TAATTTGCGC AGCATCTTTG GAACTGATGA 1380 AAAGGCCCAA CAGCGTCGTA AACAGTGGTG GAATGAATCT AAAGCACAAA TTTGGACAGC 1440 AATGATGTAC TCAGTTAAAA AAAGATTAAA GGGGAATTTT ATATGGATTT GTAAATTAAA 1500 TGTTGCGGTA AATATAGAAC CGCAGATATA TAGATGGATT CGAGAATGGG GAAGGGATTA 1560 25 CGTGTCAGAA TTGCCCACAG AAGTGCAAAA ACTGAAAGAA AAATGTGATG GAAAAATCAA 1620 TTATACTGAT AAAAAAGTAT GTAAGGTACC ACCATGTCAA AATGCGTGTA AATCATATGA 1680 TCAATGGATA ACCAGAAAAA AAAATCAATG GGATGTTCTG TCAAATAAAT TCATAAGTGT 1740 AAAAAACGCA GAAAAGGTTC AGACGGCAGG TATCGTAACT CCTTATGATA TACTAAAACA 1800 GGAGTTAGAT GAATTTAACG AGGTGGCTTT TGAGAATGAA ATTAACAAAC GTGATGGTGC 1860 ATATATTGAG TTATGCGTTT GTTCCGTTGA AGAGGCTAAA AAAAATACTC AGGAAGTTGT 1920 30 GACAAATGTG GACAATGCTG CTAAATCTCA GGCCACCAAT TCAAATCCGA TAAGTCAGCC 1980 TGTAGATAGT AGTAAAGCGG AGAAGGTTCC AGGAGATTCT ACGCATGGAA ATGTTAACAG 2040 TGGCCAAGAT AGTTCTACCA CAGGTAAAGC TGTTACGGGG GATGGTCAAA ATGGAAATCA 2100 GACACCTGCA GAAAGCGATG TACAGCGAAG TGATATTGCC GAAAGTGTAA GTGCTAAAAA 2160 35 TGTTGATCCG CAGAAATCTG TAAGTAAAAG AAGTGACGAC ACTGCAAGCG TTACAGGTAT 2220 TGCCGAAGCT GGAAAGGAAA ACTTAGGCGC ATCAAATAGT CGACCTTCTG AGTCCACCGT 2280 TGAAGCAAAT AGCCCAGGTG ATGATACTGT GAACAGTGCA TCTATACCTG TAGTGAGTGG 2340 TGAAAACCCA TTGGTAACCC CCTATAATGG TTTGAGGCAT TCGAAAGACA ATAGTGATAG 2400 CGATGGACCT GCGGAATCAA TGGCGAATCC TGATTCAAAT AGTAAAGGTG AGACGGGAAA 2460 GGGGCAAGAT AATGATATGG CGAAGGCTAC TAAAGATAGT AGTAATAGTT CAGATGGTAC 2520 CAGCTCTGCT ACGGGTGATA CTACTGATGC AGTTGATAGG GAAATTAATA AAGGTGTTCC 2580 40 TGAGGATAGG GATAAAACTG TAGGAAGTAA AGATGGAGGG GGGGAAGATA ACTCTGCAAA 2640 TAAGGATGCA GCGACTGTAG TTGGTGAGGA TAGAATTCGT GAGAACAGCG CTGGTGGTAG 2700 CACTAATGAT AGATCAAAAA ATGACACGGA AAAGAACGGG GCCTCTACCC CTGACAGTAA 2760 ACAAAGTGAG GATGCAACTG CGCTAAGTAA AACCGAAAGT TTAGAATCAA CAGAAAGTGG 2820 GGATTTACAA AAGCATGATT TTAAAAGTAA TGATACGCCG AATGAAGAAC CAAATTCTGA 2940 TCAAACTACA GATGCAGAAG GACATGACAG GGATAGCATC AAAAATGATA AAGCAGAAAG 3000 GAGAAAGCAT ATGAATAAAG ATACTTTTAC GAAAAATACA AATAGTCACC ATTTAAATAG 3060 TAATAATAT TIGAGTAATG GAAAATTAGA TATAAAAGAA TACAAATACA GAGATGTCAA 3120 AGCAACAAGG GAAGATATTA TATTAATGTC TTCAGTACGC AAGTGCAACA ATAATATTTC 3180 TTTAGAGTAC TGTAACTCTG TAGAGGACAA AATATCATCG AATACTTGTT CTAGAGAGAA 3240 AAGTAAAAAT TTATGTTGCT CAATATCGGA TTTTTGTTTG AACTATTTTG ACGTGTATTC 3300 TTATGAGTAT CTTAGCTGCA TGAAAAAGGA ATTTGAAGAT CCATCCTACA AGTGCTTTAC 3360 55 GAAAGGGGC TTTAAAGGTA TGCAGAAAAA GATGCTGAAT AGAGAAAGGT GTTGAGTAAA 3420 TTAAAAAGGA ATTAATTTTA GGAATGTTAT AAACATTTTT GTACCCAAAA TTCTTTTGC 3480 AGACAAGACT TACTTTGCCG CGGCGGGAGC GTTGCTGATA CTGCTGTTGT TAATTGCTTC 3540 ANDACTACAA TAACAATTAA AATGAGAAAT GCCTGTTAAT GCACAGTTAA 3660 60 TTCTAACGAT TCCATTTGTG AAGTTTTAAA GAGAGCACAA ATGCATAGTC ATTATGTCCA 3720 TGCATATATA CACATATATG TACGTATATA TAATAAACGC ACACTTTCTT GTTCGTACAG 3780 TTCTGAAGAA GCTACATTTA ATGAGTTTGA AGAATACTGT GATAATATTC ACAGAATCCC 3840 TCTGATGCCT AACAGTAATT CAAATTTCAA GAGCAAAATT CCATTTAAAA AGAAATGTTA 3900 CATCATTTTG CGTTTTTCTT TTTTTCTTT TTTTTCTTT TTTTAGATATT GAACACATGC 3960

-33

AGCCATCAAC CCCCCTGGAT TATTCATGAT GCTACTTTGG TAAGTAAAAG CAATTCTGAT 4020 TGTAGTGCTG ATGTAATTTT AGTCATTTTG CTTGCTGCAA TAAACGAGAA AATATATCAA 4080 GCTT 4084

- 5 (2) INFORMATION FOR SEQ ID NO:2:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1115 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
- 15 (iii) HYPOTHETICAL: NO

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- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Plasmodium vivax
- 20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Lys Gly Lys Asn Arg Ser Leu Phe Val Leu Leu Val Leu Leu Leu 10 Leu His Lys Val Ser Tyr Lys Asp Asp Phe Ser Ile Thr Leu Ile Asn 25 25 Tyr His Glu Gly Lys Lys Tyr Leu Ile Ile Leu Lys Arg Lys Leu Glu 40 45 Lys Ala Asn Asn Arg Asp Val Cys Asn Phe Phe Leu His Phe Ser Gln 30 Val Asn Asn Val Leu Leu Glu Arg Thr Ile Glu Thr Leu Leu Glu Cys 75 70 Lys Asn Glu Tyr Val Lys Gly Glu Asn Gly Tyr Lys Leu Ala Lys Gly 85 90 His His Cys Val Glu Glu Asp Asn Leu Glu Arg Trp Leu Gln Gly Thr 35 100 105 110 Asn Glu Arg Arg Ser Glu Glu Asn Ile Lys Tyr Lys Tyr Gly Val Thr 115 120 125 Glu Leu Lys Ile Lys Tyr Ala Gln Met Asn Gly Lys Arg Ser Ser Arg 135 140 40 . Ile Leu Lys Glu Ser Ile Tyr Gly Ala His Asn Phe Gly Gly Asn Ser 150 155 Tyr Met Glu Gly Lys Asp Gly Gly Asp Lys Thr Gly Glu Glu Lys Asp 165 . 170 175 Gly Glu His Lys Thr Asp Ser Lys Thr Asp Asn Gly Lys Gly Ala Asn 45 180 185 190 Asn Leu Val Met Leu Asp Tyr Glu Thr Ser Ser Asn Gly Gln Pro Ala 200 205 Gly Thr Leu Asp Asn Val Leu Glu Phe Val Thr Gly His Glu Gly Asn 215 220 50 Ser Arg Lys Asn Ser Ser Asn Gly Gly Asn Pro Tyr Asp Ile Asp His 230 235 Lys Lys Thr Ile Ser Ser Ala Ile Ile Asn His Ala Phe Leu Gln Asn 245 250 Thr Val Met Lys Asn Cys Asn Tyr Lys Arg Lys Arg Arg Glu Arg Asp 265 260 Trp Asp Cys Asn Thr Lys Lys Asp Val Cys Ile Pro Asp Arg Arg Tyr 275 280 Gln Leu Cys Met Lys Glu Leu Thr Asn Leu Val Asn Asn Thr Asp Thr 295 300 60 Asn Phe His Arg Asp Ile Thr Phe Arg Lys Leu Tyr Leu Lys Arg Lys 310 315 Leu Ile Tyr Asp Ala Ala Val Glu Gly Asp Leu Leu Leu Lys Leu Asn

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Asn Tyr Arg Tyr Asn Lys Asp Phe Cys Lys Asp Ile Arg Trp Ser Leu

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				340					34					35		
			355					360	0				Glu 36	5		_
5	Tyr	Ser 370		Val	Val	Glu	Asn 375		Leu	Arg	Ser	Ile 38	Phe 0	Gly	Thr	Asp
	Glu 385	Lys	Ala	Gln	Gln	Arg 390	Arg	Lys	Gln	Trp	Trp 395	Asn	Glu	Ser	Lys	Ala
	Gln	Ile	Trp	Thr	Ala 405		Met	Tyr	Ser	Val		Lys	Arg	Leu		
10	Asn	Phe	Ile	Trp 420	Ile		Lys	Leu	Asn 42	Val		Val	Asn	Ile 43	Glu	
	Gln	Ile	Tyr 435	Arg		Ile	Arg	Glu 440		Gly	Arg	Asp	Tyr 44	Val		Glu
15	Leu	Pro 450	Thr		Val	Gln	Lys 455		Lys	Glu	Lys	Cys 46	Asp		Lys	Ile
	Asn 465	Tyr	Thr	Asp	Lys	Lys 470		Cys	Lys	Val	Pro 475	-	Cys	Gln	Asn	Ala 480
		Lys	Ser	Tyr	Asp 485		Trp	Ile	Thr	Arg 49		Lys	Asn	Gln	Trp	Asp
20	Val	Leu	Ser	Asn 500		Phe	Ile	Ser	Val 509		Asn	Ala	Glu	Lys 51	Val	
	Thr	Ala	Gly 515		Val	Thr	Pro	Tyr 520		Ile	Leu	Lys	Gln 52		Ļeu	Asp
25	Glu	Phe 530	Asn	Glu	Val	Ala	Phe 535		Asn	Glu	Ile	Asn 54	Lys 0	Arg	Asp	Gly
	Ala 545	Tyr	Ile	Glu	Leu	Cys 550	Val	Cys	Ser	Val	Glu 555	Glu	Ala	Lys	Lys	Asn 560
					565	•			_	57	0		Lys		57	5
30				580					585	5			Ser	59	0	
	Lys	Val	Pro 595	Gly	Asp	Ser	Thr	His 600		Asn	Val	Asn	Ser 60		Gln	Asp
35	Ser	Ser 610	Thr	Thr	Gly	Lys	Ala 615		Thr	Gly	Asp	Gly 62	Gln 0	Asn	Gly	Asn
	625					630					635		Ile			640
.1				-	645					650	0		Ser	_	65	5
40	Asp	Asp	Thr	Ala 660		Val	Thr	Gly	Ile 669		Glu	Ala	Gly	Lys 67		Asn
			675				_	680)				Val 68	5		
45		690	_	-	_		695	;				70				
	705					710					715		Arg			720
					725					73	0		Ala		73	5
50				740	_			_	749	5		_	Asn	75	0	
			755	_	_			760)		_		Thr 76	5		
55		770	_			_	775	;		_		78	-	_	-	
•	785		_	_	_	790			-		795		Gly	•	_	800
	Asp	Asn	Ser	Ala	Asn 605		Asp	Ala	Ala	Thr 81		Val	Gly	Glu	Asp ĩŝ	
60	Ile	Arg	Glu	Asn 820	Ser	Ala	Gly	Gly	Ser 825		Asn	Asp	Arg	Ser 83		Asn
	_		835	-		_		840)		_		Lys 84	5		
	Asp	Ala	Thr	Ala	Leu	Ser	Lys	Thr	Glu	Ser	Leu	Glu	Ser	Thr	Glu	Ser

850 855 Gly Asp Arg Thr Thr Asn Asp Thr Thr Asn Ser Leu Glu Asn Lys Asn 870 875 Gly Gly Lys Glu Lys Asp Leu Gln Lys His Asp Phe Lys Ser Asn Asp 5 885 890 Thr Pro Asn Glu Glu Pro Asn Ser Asp Gln Thr Thr Asp Ala Glu Gly 900 905 His Asp Arg Asp Ser Ile Lys Asn Asp Lys Ala Glu Arg Arg Lys His 915 920 Met Asn Lys Asp Thr Phe Thr Lys Asn Thr Asn Ser His His Leu Asn 10 935 940 Ser Asn Asn Asn Leu Ser Asn Gly Lys Leu Asp Ile Lys Glu Tyr Lys 950 955 Tyr Arg Asp Val Lys Ala Thr Arg Glu Asp Ile Ile Leu Met Ser Ser 15 965 970 Val Arg Lys Cys Asn Asn Asn Ile Ser Leu Glu Tyr Cys Asn Ser Val 980 985 Glu Asp Lys Ile Ser Ser Asn Thr Cys Ser Arg Glu Lys Ser Lys Asn 995 1000 20 Leu Cys Cys Ser Ile Ser Asp Phe Cys Leu Asn Tyr Phe Asp Val Tyr 1010 1015 1020 Ser Tyr Glu Tyr Leu Ser Cys Met Lys Lys Glu Phe Glu Asp Pro Ser 1025 1030 1035 Tyr Lys Cys Phe Thr Lys Gly Gly Phe Lys Ile Asp Lys Thr Tyr Phe 25 1045 1050 Ala Ala Ala Gly Ala Leu Leu Ile Leu Leu Leu Ile Ala Ser Arg Lys 1060 1065 Met Ile Lys Asn Asp Ser Glu Glu Ala Thr Phe Asn Glu Phe Glu Glu 1075 1080 30 Tyr Cys Asp Asn Ile His Arg Ile Pro Leu Met Pro Asn Asn Ile Glu 1090 1095 His Met Gln Pro Ser Thr Pro Leu Asp Tyr Ser 1110 (2) INFORMATION FOR SEQ ID NO:3: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 4507 base pairs (B) TYPE: nucleic acid 40 . (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) 45 (iii) HYPOTHETICAL: NO (vi) ORIGINAL SOURCE: (A) ORGANISM: Plasmodium falciparum 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3: ' TATATATATA TATATATATA GATAATAACA TATAAATATA TTCAATGTGC ATACAATGAA 60 ATGTAATATT AGTATATT TTTTTGCTTC CTTCTTTGTG TTATATTTTG CAAAAGCTAG 120 GAATGAATAT GATATAAAAG AGAATGAAAA ATTTTTAGAC GTGTATAAAG AAAAATTTAA 180 55 TGAATTAGAT AAAAAGAAAT ATGGAAATGT TCAAAAAACT GATAAGAAAA TATTTACTTT 240 TATAGAAAAT AAATTAGATA TTTTAAATAA TTCAAAATTT AATAAAAGAT GGAAGAGTTA 300 TGGAACTCCA GATAATATAG ATAAAAATAT GTCTTTAATA AATAAACATA ATAATGAAGA 360 AATGTTTAAC AACAATTATC AATCATTTTT ATCGACAAGT TCATTAATAA AGCAAAATAA 420 ATATGTTCCT ATTAACGCTG TACGTGTGTC TAGGATATTA AGTTTCCTGG ATTCTAGAAT 480 TAATAATGGA AGAAATACTT CATCTAATAA CGAAGTTTTA AGTAATTGTA GGGAAAAAAG 540 GAAAGGAATG AAATGGGATT GTAAAAAGAA AAATGATAGA AGCAACTATG TATGTATTCC 600 TGATCGTAGA ATCCAATTAT GCATTGTTAA TCTTAGCATT ATTAAAACAT ATACAAAAGA 660 GACCATGAAG GATCATTTCA TTGAAGCCTC TAAAAAAGAA TCTCAACTTT TGCTTAAAAA 720

AAATGATAAC AAATATAATT CTAAATTTTG TAATGATTTG AAGAATAGTT TTTTAGATTA 780

TGGACATCTT GCTATGGGAA ATGATATGGA TTTTGGAGGT TATTCAACTA AGGCAGAAAA 840 CAAAATTCAA GAAGTTTTTA AAGGGGCTCA TGGGGAAATA AGTGAACATA AAATTAAAAA 900 TTTTAGAAAA GAATGGTGGA ATGAATTTAG AGAGAAACTT TGGGAAGCTA TGTTATCTGA 960 GCATAAAAAT AATATAAATA ATTGTAAAAA TATTCCCCAA GAAGAATTAC AAATTACTCA 1020 ATGGATAAAA GAATGGCATG GAGAATTTTT GCTTGAAAGA GATAATAGAT CAAAATTGCC 1080 AAAAAGTAAA TGTAAAAATA ATACATTATA TGAAGCATGT GAGAAGGAAT GTATTGATCC 1140 ATGTATGAAA TATAGAGATT GGATTATTAG AAGTAAATTT GAATGGCATA CGTTATCGAA 1200 AGAATATGAA ACTCAAAAAG TTCCAAAGGA AAATGCGGAA AATTATTTAA TCAAAATTTC 1260 AGAAAACAAG AATGATGCTA AAGTAAGTTT ATTATTGAAT AATTGTGATG CTGAATATTC 1320 AAAATATTGT GATTGTAAAC ATACTACTAC TCTCGTTAAA AGCGTTTTAA ATGGTAACGA 1380 CAATACAATT AAGGAAAAGC GTGAACATAT TGATTTAGAT GATTTTCTA AATTTGGATG 1440 10 TGATAAAAAT TCCGTTGATA CAAACACAAA GGTGTGGGAA TGTAAAAACC CTTATATATT 1500 ATCCACTAAA GATGTATGTG TACCTCCGAG GAGGCAAGAA TTATGTCTTG GAAACATTGA 1560 TAGAATATAC GATAAAAACC TATTAATGAT AAAAGAGCAT ATTCTTGCTA TTGCAATATA 1620 15 TGAATCAAGA ATATTGAAAC GAAAATATAA GAATAAAGAT GATAAAGAAG TTTGTAAAAT 1680 CATAAATAAA ACTITCGCTG ATATAAGAGA TATTATAGGA GGTACTGATT ATTGGAATGA 1740 TTTGAGCAAT AGAAAATTAG TAGGAAAAAT TAACACAAAT TCAAAATATG TTCACAGGAA 1800 TAAAAAAAT GATAAGCTTT TTCGTGATGA GTGGTGGAAA GTTATTAAAA AAGATGTATG 1860 GAATGTGATA TCATGGGTAT TCAAGGATAA AACTGTTTGT AAAGAAGATG ATATTGAAAA 1920 TATACCACAA TTCTTCAGAT GGTTTAGTGA ATGGGGTGAT GATTATTGCC AGGATAAAAC 1980 AAAAATGATA GAGACTCTGA AGGTTGAATG CAAAGAAAAA CCTTGTGAAG ATGACAATTG 2040 TAAAAGTAAA TGTAATTCAT ATAAAGAATG GATATCAAAA AAAAAAGAAG AGTATAATAA 2100 ACAAGCCAAA CAATACCAAG AATATCAAAA AGGAAATAAT TACAAAATGT ATTCTGAATT 2160 TAAATCTATA AAACCAGAAG TITATTTAAA GAAATACTCG GAAAAATGTT CTAACCTAAA 2220 25 TTTCGAAGAT GAATTTAAGG AAGAATTACA TTCAGATTAT AAAAATAAAT GTACGATGTG 2280 TCCAGAAGTA AAGGATGTAC CAATTTCTAT AATAAGAAAT AATGAACAAA CTTCGCAAGA 2340 AGCAGTTCCT GAGGAAAACA CTGAAATAGC ACACAGAACG GAAACTCCAT CTATCTCTGA 2400 AGGACCAAAA GGAAATGAAC AAAAAGAACG TGATGACGAT AGTTTGAGTA AAATAAGTGT 2460 ATCACCAGAA AATTCAAGAC CTGAAACTGA TGCTAAAGAT ACTTCTAACT TGTTAAAATT 2520 30 AAAAGGAGAT GTTGATATTA GTATGCCTAA AGCAGTTATT GGGAGCAGTC CTAATGATAA 2580 TATAAATGTT ACTGAACAAG GGGATAATAT TTCCGGGGTG AATTCTAAAC CTTTATCTGA 2640 TGATGTACGT CCAGATAAAA AGGAATTAGA AGATCAAAAT AGTGATGAAT CGGAAGAAAC 2700 TGTAGTAAAT CATATATCAA AAAGTCCATC TATAAATAAT GGAGATGATT CAGGCAGTGG 2760 AAGTGCAACA GTGAGTGAAT CTAGTAGTTC AAATACTGGA TTGTCTATTG ATGATGATAG 2820 AAATGGTGAT ACATTTGTTC GAACACAAGA TACAGCAAAT ACTGAAGATG TTATTAGAAA 2880 35 AGAAAATGCT GACAAGGATG AAGATGAAAA AGGCGCAGAT GAAGAAAGAC ATAGTACTTC 2940 TGAAAGCTTA AGTTCACCTG AAGAAAAAAT GTTAACTGAT AATGAAGGAG GAAATAGTTT 3000 AAATCATGAA GAGGTGAAAG AACATACTAG TAATTCTGAT AATGTTCAAC AGTCTGGAGG 3060 AATTGTTAAT ATGAATGTTG AGAAAGAACT AAAAGATACT TTAGAAAATC CTTCTAGTAG 3120 40 CTTGGATGAA GGAAAAGCAC ATGAAGAATT ATCAGAACCA AATCTAAGCA GTGACCAAGA 3180 TATGTCTAAT ACACCTGGAC CTTTGGATAA CACCAGTGAA GAAACTACAG AAAGAATTAG 3240 TAATAATGAA TATAAAGTTA ACGAGAGGGA AGATGAGAGA ACGCTTACTA AGGAATATGA 3300 AGATATTGTT TTGAAAAGTC ATATGAATAG AGAATCAGAC GATGGTGAAT TATATGACGA 3360 AAATTCAGAC TTATCTACTG TAAATGATGA ATCAGAAGAC GCTGAAGCAA AAATGAAAGG 3420 45 AAATGATACA TCTGAAATGT CGCATAATAG TAGTCAACAT ATTGAGAGTG ATCAACAGAA 3480 AAACGATATG AAAACTGTTG GTGATTTGGG AACCACACT GTACAAAACG AAATTAGTGT 3540 TCCTGTTACA GGAGAAATTG ATGAAAAATT AAGGGAAAGT AAAGAATCAA AAATTCATAA 3600 GGCTGAAGAG GAAAGATTAA GTCATACAGA TATACATAAA ATTAATCCTG AAGATAGAAA 3660 TAGTAATACA TTACATTTAA AAGATATAAG AAATGAGGAA AACGAAAGAC ACTTAACTAA 3720 50 TCAAAACATT AATATTAGTC AAGAAAGGGA TITGCAAAAA CATGGATTCC ATACCATGAA 3780 TAATCTACAT GGAGATGGAG TTTCCGAAAG AAGTCAAATT AATCATAGTC ATCATGGAAA 3840 CAGACAAGAT CGGGGGGAA ATTCTGGGAA TGTTTTAAAT ATGAGATCTA ATAATAATAA 3900 TTTTAATAAT ATTCCAAGTA GATATAATTT ATATGATAAA AAATTAGATT TAGATCTTTA 3960 TGAAAACAGA AATGATAGTA CAACAAAGA ATTAATAAAG AAATTAGCAG AAATAAATAA 4020 55 ATGTGAGAAC GAAATTTCTG TAAAATATTG TGACCATATG ATTCATGAAG AAATCCCATT 4080 AAAAACATGC ACTAAAGAAA AAACAAGAAA TCTGTGTTGT GCAGTATCAG ATTACTGTAT 4140 GAGCTATTTT ACATATGATT CAGAGGAATA TTATAATTGT ACGAAAAGGG AATTTGATGA 4200 TCCATCTTAT ACATGTTTCA GAAAGGAGGC TTTTTCAAGT ATGATATTCA AATTTTTAAT 4260 ABCANATIAN ATATATTATT ATTITITATAC TIACABARCT GCARAGTAR CARTABARAR 4320 60 AATTAATTTC TCATTAATTT TTTTTTTCTT TTTTTCTTTT TAGGTATGCC ATATTATGCA 4380 GGAGCAGGTG TGTTATTTAT TATATTGGTT ATTTTAGGTG CTTCACAAGC CAAATATCAA 4440 AGGTTAGAAA AAATAAATAA AAATAAAATT GAGAAGAATG TAAATTAAAT ATAGAATTCG 4500 AGCTCGG

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(2) INFORMATION FOR SEQ ID NO:4:
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(i) SEQUENCE CHARACTERISTICS:
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- (A) LENGTH: 1435 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
 - (vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium falciparum

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Lys Cys Asn Ile Ser Ile Tyr Phe Phe Ala Ser Phe Phe Val Leu 10 20 Tyr Phe Ala Lys Ala Arg Asn Glu Tyr Asp Ile Lys Glu Asn Glu Lys Phe Leu Asp Val Tyr Lys Glu Lys Phe Asn Glu Leu Asp Lys Lys 40 Tyr Gly Asn Val Gln Lys Thr Asp Lys Lys Ile Phe Thr Phe Ile Glu 25 Asn Lys Leu Asp Ile Leu Asn Asn Ser Lys Phe Asn Lys Arg Trp Lys 70 75 Ser Tyr Gly Thr Pro Asp Asn Ile Asp Lys Asn Met Ser Leu Ile Asn 90 30 Lys His Asn Asn Glu Glu Met Phe Asn Asn Asn Tyr Gln Ser Phe Leu 105 110 Ser Thr Ser Ser Leu Ile Lys Gln Asn Lys Tyr Val Pro Ile Asn Ala 120 Val Arg Val Ser Arg Ile Leu Ser Phe Leu Asp Ser Arg Ile Asn Asn 35 135 140 Gly Arg Asn Thr Ser Ser Asn Asn Glu Val Leu Ser Asn Cys Arg Glu 150 155 Lys Arg Lys Gly Met Lys Trp Asp Cys Lys Lys Asn Asp Arg Ser 165 170 175 40 Asn Tyr Val Cys Ile Pro Asp Arg Arg Ile Gln Leu Cys Ile Val Asn 180 185 190 Leu Ser Ile Ile Lys Thr Tyr Thr Lys Glu Thr Met Lys Asp His Phe 200 205 Ile Glu Ala Ser Lys Lys Glu Ser Gln Leu Leu Lys Lys Asn Asp 45 215 220 Asn Lys Tyr Asn Ser Lys Phe Cys Asn Asp Leu Lys Asn Ser Phe Leu 230 235 Asp Tyr Gly His Leu Ala Met Gly Asn Asp Met Asp Phe Gly Gly Tyr 245 ·250 50 Ser Thr Lys Ala Glu Asn Lys Ile Gln Glu Val Phe Lys Gly Ala His 265 Gly Glu Ile Ser Glu His Lys Ile Lys Asn Phe Arg Lys Glu Trp Trp 280 275 Asn Glu Phe Arg Glu Lys Leu Trp Glu Ala Met Leu Ser Glu His Lys 55 295 300 Asn Asn Ile Asn Asn Cys Lys Asn Ile Pro Gln Glu Glu Leu Gln Ile 310 315 Thr Gln Trp Ile Lys Glu Trp His Gly Glu Phe Leu Leu Glu Arg Asp 330 325 60 Asn Arg Ser Lys Leu Pro Lys S r Lys Cys Lys Asn Asn Thr Leu Tyr 345 Glu Ala Cys Glu Lys Glu Cys Ile Asp Pro Cys Met Lys Tyr Arg Asp 360 365 Trp Ile Ile Arg Ser Lys Phe Glu Trp His Thr Leu Ser Lys Glu Tyr

		370					375					380				
	385				Val	390					395					400
5					Lys 405					410					415	Asn
				420					425					430	Thr	Thr
			435		Val			440					445	Lys	Glu	
10		450			Asp		455					460			_	_
	465				Thr	470					475					480
15					Lys 485					490					495	
				500	Ile				505					510		
20			515		Leu			520					525			
20		530			Asn		535					540				
	545				Asp	550					555				_	560
25					Asn 565					570					575	
				580	Arg Ile				585					590	_	
30			595		Thr			600					605	•		
		610			Trp		615		·			620				
	625				Ile	630	-				635					640
35					645 Asn					650					655	
				660	Lys				665					670		_
40			675	-	Gly	,		680	•				685	•	_	
		690			Val		695	•				700			_	
	705				Asp	710					715					720
45					725 Met					730					735	_
				740	Glu				745					750		
50			755		His			760					765			
	Lys	770			Gln		775					780			_	
	785				Glu	790					795					800
55	Ser	Asn	Leu	Leu	805 Lys	Leu	Lys	Gly	Asp	810 Val	Asp	Ile	Ser	Met	815 Pro	Lys
	Ala	Val		820 Gly	Ser	Ser	Pro	Asn	825 Asp	Asn	Ile	Asn	Val	830 Thr	Glu	Gln
60	Gly		Asn	Ile	Ser	Gly		840 Asn	Ser	Lys	Pro		845 Ser	Asp	Asp	Val
	Arg	850 Pro	Asp	Lys	Lys		855 Leu	Glu	Asp	Gln		860 Ser	Asp	Glu	Ser	Glu
	865 Glu	Thr	Val	Val	Asn	870 His	Ile	Ser	Lys	Ser	875 Pro	Ser	Ile	Asn	Asn	880 Gly

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885
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             915
                                920
     Arg Thr Gln Asp Thr Ala Asn Thr Glu Asp Val Ile Arg Lys Glu Asn
                            935
                                               940
     Ala Asp Lys Asp Glu Asp Glu Lys Gly Ala Asp Glu Glu Arg His Ser
                        950
                                            955
     Thr Ser Glu Ser Leu Ser Ser Pro Glu Glu Lys Met Leu Thr Asp Asn
10
                     965
                                        970
     Glu Gly Gly Asn Ser Leu Asn His Glu Glu Val Lys Glu His Thr Ser
                                    985
                                                        990
     Asn Ser Asp Asn Val Gln Gln Ser Gly Gly Ile Val Asn Met Asn Val
15
                               1000
                                                   1005
     Glu Lys Glu Leu Lys Asp Thr Leu Glu Asn Pro Ser Ser Ser Leu Asp
                           1015
                                              1020
     Glu Gly Lys Ala His Glu Glu Leu Ser Glu Pro Asn Leu Ser Ser Asp
                        1030
                                            1035
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     Gln Asp Met Ser Asn Thr Pro Gly Pro Leu Asp Asn Thr Ser Glu Glu
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                                      1050
                                                          1055
     Thr Thr Glu Arg Ile Ser Asn Asn Glu Tyr Lys Val Asn Glu Arg Glu
               1060
                                 1065
     Asp Glu Arg Thr Leu Thr Lys Glu Tyr Glu Asp Ile Val Leu Lys Ser
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                             1080
                                                  1085
     His Met Asn Arg Glu Ser Asp Asp Gly Glu Leu Tyr Asp Glu Asn Ser
                           1095
                                              1100
     Asp Leu Ser Thr Val Asn Asp Glu Ser Glu Asp Ala Glu Ala Lys Met
                        1110
                                            1115
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     Lys Gly Asn Asp Thr Ser Glu Met Ser His Asn Ser Ser Gln His Ile
                    1125
                                       1130
     Glu Ser Asp Gln Gln Lys Asn Asp Met Lys Thr Val Gly Asp Leu Gly
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                                   1145
                                                       1150
     Thr Thr His Val Gln Asn Glu Ile Ser Val Pro Val Thr Gly Glu Ile
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           1155
                               1160
                                                   1165
     Asp Glu Lys Leu Arg Glu Ser Lys Glu Ser Lys Ile His Lys Ala Glu
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                           1175
                                              1180
     Glu Glu Arg Leu Ser His Thr Asp Ile His Lys Ile Asn Pro Glu Asp
                        1190
                                            1195
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     Arg Asn Ser Asn Thr Leu His Leu Lys Asp Ile Arg Asn Glu Glu Asn
                   1205
                                      1210
                                                          1215
     Glu Arg His Leu Thr Asn Gln Asn Ile Asn Ile Ser Gln Glu Arg Asp
                1220
                                  1225
                                                       1230
     Leu Gln Lys His Gly Phe His Thr Met Asn Asn Leu His Gly Asp Gly
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            1235
                               1240
                                                   1245
     Val Ser Glu Arg Ser Gln Ile Asn His Ser His His Gly Asn Arg Gln
                           1255
                                              1260
     Asp Arg Gly Gly Asn Ser Gly Asn Val Leu Asn Met Arg Ser Asn Asn
                        1270
                                           1275
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     Asn Asn Phe Asn Asn Ile Pro Ser Arg Tyr Asn Leu Tyr Asp Lys Lys
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                   1285
                                                          1295
     Leu Asp Leu Asp Leu Tyr Glu Asn Arg Asn Asp Ser Thr Thr Lys Glu
                           1305
              1300
                                                      1310
     Leu Ile Lys Lys Leu Ala Glu Ile Asn Lys Cys Glu Asn Glu Ile Ser
55
           1315
                               1320
                                                  1325
     Val Lys Tyr Cys Asp His Met Ile His Glu Glu Ile Pro Leu Lys Thr
                           1335
                                              1340
     Cys Thr Lys Glu Lys Thr Arg Asn Leu Cys Cys Ala Val Ser Asp Tyr
                                                               1360
                       1350
                                           1355
60
     Cys Met Ser Tyr Phe Thr Tyr Asp Ser Glu Glu Tyr Tyr Asn Cys Thr
                   1365
                                    1370
                                                        1375
     Lys Arg Glu Phe Asp Asp Pro Ser Tyr Thr Cys Phe Arg Lys Glu Ala
                                   1385
     Phe Ser Ser Met Ile Phe Lys Phe Leu Ile Thr Asn Lys Ile Tyr Tyr
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GCCGCTCT

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1400
              1395
                                                        1405
       Tyr Phe Tyr Thr Tyr Lys Thr Ala Lys Val Thr Ile Lys Lys Ile Asn
                              1415
                                                  1420
       Phe Ser Leu Ile Phe Phe Phe Phe Phe Ser Phe
 5
                            1430
                                                 1435
      (2) INFORMATION FOR SEQ ID NO:5:
10
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 2288 base pairs
                (B) TYPE: nucleic acid
                (C) STRANDEDNESS: single
15
                (D) TOPOLOGY: linear
          (ii) MOLECULE TYPE: DNA (genomic)
         (iii) HYPOTHETICAL: NO
20
          (vi) ORIGINAL SOURCE:
                (A) ORGANISM: Plasmodium falciparum
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:
25
     CACTITATGC TTCCGGCTCG TATGTTGTGT GGAATTGTGA GCGGATAACA ATTTCACACA 60
     GGAAACAGCT ATGACCATGA TTACGCCAAG CTCTAATACG ACTCACTATA GGGAAAGCTG 120
     GTACGCCTGC AGGTCCGGTC CGGAATTCAA TAAAATATTT CCAGAAAGGA ATGTGCAAAT 180
     TCACATATCC AATATATTCA AGGAATATAA AGAAAATAAT GTAGATATCA TATTTGGAAC 240
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     GTTGAATTAT GAATATAATA ATTTCTGTAA AGAAAAACCT GAATTAGTAT CTGCTGCCAA 300
     GTATAATCTG AAAGCTCCAA ATGCTAAATC CCCTAGAATA TACAAATCTA AGGAGCATGA 360
     AGAATCAAGT GTGTTTGGTT GCAAAACGAA AATCAGTAAA GTTAAAAAAA AATGGAATTG 420
     TTATAGTAAT AATAAAGTAA CTAAACCTGA AGGTGTATGT GGACCACCAA GAAGGCAACA 480
     ATTATGTCTT GGATATATAT TTTTGATTCG CGACGGTAAC GAGGAAGGAT TAAAAGATCA 540
     TATTAATAAG GCAGCTAATT ATGAGGCAAT GCATTTAAAA GAGAAATATG AGAATGCTGG 600
     TGGTGATAAA ATTTGCAATG CTATATTGGG AAGTTATGCA GATATTGGAG ATATTGTAAG 660
     AGGTTTGGAT GTTTGGAGGG ATATAAATAC TAATAAATTA TCAGAAAAAT TCCAAAAAAT 720
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     GTGGGAAAAA CAAAGGAATT TAATATGGTC TAGTATGGTA AAACACATTC CAAAAGGAAA 840
     AACATGTAAA CGTCATAATA ATTTTGAGAA AATTCCTCAA TTTTTGAGAT GGTTAAAAGA 900
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     ATGGGGTGAT GAATTTTGTG AGGAAATGGG TACGGAAGTC AAGCAATTAG AGAAAATATG 960
     TGAAAATAAA AATTGTTCGG AAAAAAAATG TAAAAATGCA TGTAGTTCCT ATGAAAAATG 1020
     GATAAAGGAA CGAAAAAATG AATATAATTT GCAATCAAAG AAATTTGATA GTGATAAAAA 1080
     ATTAAATAAA AAAAACAATC TTTATAATAA ATTTGAGGAT TCTAAAGCTT ATTTAAGGAG 1140
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     TGAATCAAAA CAGTGCTCAA ATATAGAATT TAATGATGAA ACATTTACAT TTCCTAATAA 1200
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     ATCTTCTGAT TCTATACCAA TTACTCATAT AGAAGCTGAA AAGGGTCAGT CTTCTAATTC 1680
     TAGCGATAAT GATCCTGCAG TAGTAAGTGG TAGAGAATCT AAAGATGTAA ATCTTCATAC 1740
     TTCTGAAAGG ATTAAAGAAA ATGAAGAAGG TGTGATTAAA ACAGATGATA GTTCAAAAAG 1800
     TATTGAAATT TCTAAAATAC CATCTGACCA AAATAATCAT AGTGATTTAT CACAGAATGC 1860
     AAATGAGGAC TCTAATCAAG GGAATAAGGA AACAATAAAT CCTCCTTCTA CAGAAAAAA 1920
     TCTCAAAGAA ATTCATTATA AAACATCTGA TTCTGATGAT CATGGTTCTA AAATTAAAAG 1980
     TGANATTOAN CCANAGGAGT TANCGGAGGA ATCACCTCTT ACTGATAGAA AGACTGAGAG ZU4U
     TGCAGCGATT GGTGATAAAA ATCATGAATC AGTAAAAAGC GCTGATATTT TTCAATCTGA 2100
     GATTCATAAT TCTGATAATA GAGATAGAAT TGTTTCTGAA AGTGTAGTTC AGGATTCTTC 2160
     AGGAAGCTCT ATGAGTACTG AATCTATACG TACTGATAAC AAGGATTTTA AAACAAGTGA 2220
     GGATATTGCA CCTTCTATTA ATGGTCGGAA TTCCCGGGTC GACGAGCTCA CTAGTCGGCG 2280
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(2) INFORMATION FOR SEQ ID NO:6: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 749 amino acids 5 (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein 10 (iii) HYPOTHETICAL: NO (vi) ORIGINAL SOURCE: (A) ORGANISM: Plasmodium falciparum 15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6: Ala Asp Asn Asn Phe Thr Gln Glu Thr Ala Met Thr Met Ile Thr Pro 20 Ser Ser Asn Thr Thr His Tyr Arg Glu Ser Trp Tyr Ala Cys Arg Ser 20 25 Gly Pro Glu Phe Asn Lys Ile Phe Pro Glu Arg Asn Val Gln Ile His 35 Ile Ser Asn Ile Phe Lys Glu Tyr Lys Glu Asn Asn Val Asp Ile Ile 25 55 Phe Gly Thr Leu Asn Tyr Glu Tyr Asn Asn Phe Cys Lys Glu Lys Pro 70 Glu Leu Val Ser Ala Ala Lys Tyr Asn Leu Lys Ala Pro Asn Ala Lys 85 90 30 Ser Pro Arg Ile Tyr Lys Ser Lys Glu His Glu Glu Ser Ser Val Phe 100 105 Gly Cys Lys Thr Lys Ile Ser Lys Val Lys Lys Lys Trp Asn Cys Tyr 120 125 Ser Asn Asn Lys Val Thr Lys Pro Glu Gly Val Cys Gly Pro Pro Arg 35 130 135 Arg Gln Gln Leu Cys Leu Gly Tyr Ile Phe Leu Ile Arg Asp Gly Asn 150 155 Glu Glu Gly Leu Lys Asp His Ile Asn Lys Ala Ala Asn Tyr Glu Ala 165 170 40 Met His Leu Lys Glu Lys Tyr Glu Asn Ala Gly Gly Asp Lys Ile Cys 185 190 Asn Ala Ile Leu Gly Ser Tyr Ala Asp Ile Gly Asp Ile Val Arg Gly 195 200 205 Leu Asp Val Trp Arg Asp Ile Asn Thr Asn Lys Leu Ser Glu Lys Phe 45 210 215 220 Gln Lys Ile Phe Met Gly Gly Gly Asn Ser Arg Lys Lys Gln Asn Asp 230 235 Asn Asn Glu Arg Asn Lys Trp Trp Glu Lys Gln Arg Asn Leu Ile Trp 245 250 50 Ser Ser Met Val Lys His Ile Pro Lys Gly Lys Thr Cys Lys Arg His 260 265 270 Asn Asn Phe Glu Lys Ile Pro Gln Phe Leu Arg Trp Leu Lys Glu Trp 280 Gly Asp Glu Phe Cys Glu Glu Met Gly Thr Glu Val Lys Gln Leu Glu 55 290 295 300 Lys Ile Cys Glu Asn Lys Asn Cys Ser Glu Lys Lys Cys Lys Asn Ala 310 315 Cys Ser Ser Tyr Glu Lys Trp Ile Lys Glu Arg Lys Asn Glu Tyr Asn 325 330 60 Leu Gln Ser Lys Lys Phe Asp Ser Asp Lys Leu Asn Lys Lys Asn 340 345

Asn Leu Tyr Asn Lys Phe Glu Asp Ser Lys Ala Tyr Leu Arg Ser Glu

360 Ser Lys Gln Cys Ser Asn Ile Glu Phe Asn Asp Glu Thr Phe Thr Phe

		370					37	5				31	80			
	Pro 385	Asn	Lys	Tyr	Lys	Glu 390	Ala	Суѕ	Met	Val	Cys 395			Pro	Ser	Ser
5	Ser	Lys	Ala	Leu	Lys 405	Pro	Ile	Lys	Thr	Asn 41		Phe	Pro	Ile		Glu 15
	Ser	Lys	Lys	Ser 420	Glu		Ser	Ser	Leu 42	Thr		Lys	Ser		Asn 30	Thr
	Pro	Asn	Ser 435	Ser	Gly	Gly	Gly	Asn 44	Tyr		Asp	Arg		Ile 45	Ser	Lys
10	Arg	Asp 450	Asp	Val	His	His	Asp 45	Gly	Pro	Lys	Glu				Gly	Glu
	Lys 465	Glu	Val	Pro	Lys	Ile 470	Asp	Ala	Ala	Val	Lys 475	Thr	Glu	Asn	Glu	Phe 480
15	Thr	Ser	Asn	Arg	Asn 485	Asp	Ile	Glu	Gly	Lys 49	Glu	Lys	Ser	Lys	_	Asp 95
	His	Ser	Ser	Pro 500	Val	His	Ser	Lys	Asp 50	Ile		Asn	Glu		Pro 10	Gln
	Arg	Val	Val 515		Glu	Asn	Leu	Pro 52	Lys		Glu	Glu		Met 25	Glu	Ser
20	Ser	Asp 530	Ser	Ile	Pro	Ile	Thr 539	His		Glu	Ala				Gln	Ser
	Ser 545	Asn	Ser	Ser	Asp	Asn 550	Asp	Pro	Ala	Val	Val 555			Arg	Glu	Ser 560
25	Lys	Asp	Val	Asn	Leu 565	His	Thr	Ser	Glu	Arg 57		Lys	Glu	Asn		Glu 75
	Gly	Val	Ile	Lys 580		Asp	Asp	Ser	Ser 58		Ser	Ile	Glu			
	Ile	Pro	Ser 595	Asp	Gln	Asn	Asn	His		Asp	Leu	Ser	Gln 60	Asn	Ala	Asn
30	Glu	Asp 610	Ser	Asn	Gln	Gly	Asn 619		Glu	Thr	Ile	Asn 62	Pro		Ser	Thr
	Glu 625	Lys	Asn	Leu	Lys	Glu 630	Ile	His	Tyr	Lys	Thr 635	Ser	Asp	Ser	Asp	Asp 640
35	His	Gly	Ser	Lys	Ile 645		Ser	Glu	Ile	Glu 65		Lys	Glu	Leu		Glu 55
	Glu			660					66	5				6	70 -	Ξ.
·	Lys		675					68	0 .		•		68	35		
40	His	690					695	5				70	0			
	Asp 705					710					715				_	720
45	Lys				725					73	0				_	Arg 35
	Asn	Ser	Arg	Val 740	Asp	Glu	Let	ı Thi	74.		g Ar	g Pr	o Le	eu		
	(2) INPOR	רידא	CONT	ava	e EO	TD N	m. 7.									

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2606 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:

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- (A) ORGANISM: Plasmodium falciparum
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

```
AGCTCTATTA CGACTCACTA TAGGGAAAGC TGGTACGCCT GCAGGTACCG GTCCGGAATT 60
      CCCGGGTCGA CGAGCTCACT AGTCGGCGGC CGCTCTAGAG GATCCAAGCT TAATAGTGTT 120
     TATACGTCTA TTGGCTTATT TTTAAATAGC TTAAAAAGCG GACCATGTAA AAAGGATAAT 180
      GATAATGCAG AGGATAATAT AGATTTTGGT GATGAAGGTA AAACATTTAA AGAGGCAGAT 240
 5
     AATTGTAAAC CATGTTCTCA ATTTACTGTT GATTGTAAAA ATTGTAATGG TGGTGATACA 300
     AAAGGGAAGT GCAATGGCAG CAATGGCAAA AAGAATGGAA ATGATTATAT TACTGCAAGT 360
     GATATTGAAA ATGGAGGGAA TTCTATTGGA AATATAGATA TGGTTGTTAG TGATAAGGAT 420
     GCAAATGGAT TTAATGGTTT AGACGCTTGT GGAAGTGCAA ATATCTTTAA AGGTATTAGA 480
     AAAGAACAAT GGAAATGTGC TAAAGTATGT GGTTTAGATG TATGTGGTCT TAAAAATGGT 540
10
     AATGGTAGTA TAGATAAAGA TCAAAAACAA ATTATAATTA TTAGAGCATT GCTTAAACGT 600
     TGGGTAGAAT ATTTTTTAGA AGATTATAAT AAAATTAATG CCAAAATTTC ACATTGTACG 660
     AAAAAGGATA ATGAATCCAC ATGTACAAAT GATTGTCCAA ATAAATGTAC ATGTGTAGAA 720
     GAGTGGATAA ATCAGAAAAG GACAGAATGG AAAAATATAA AAAAACATTA CAAAACACAA 780
     AATGAAAATG GTGACAATAA CATGAAATCT TTGGTTACAG ATATTTTGGG TGCCTTGCAA 840
     CCCCAAAGTG ATGTTAACAA AGCTATAAAA CCTTGTAGTG GTTTAACTGC GTTCGAGAGT 900
15
     TTTTGTGGTC TTAATGGCGC TGATAACTCA GAAAAAAAG AAGGTGAAGA TTACGATCTT 960
     GTTCTATGTA TGCTTAAAAA TCTTGAAAAA CAAATTCAGG AGTGCAAAAA GAAACATGGC 1020
     GAAACTAGTG TCGAAAATGG TGGCAAATCA TGTACCCCCC TTGACAACAC CACCCTTGAG 1080
     GAGGAACCCA TAGAAGAGGA AAACCAAGTG GAAGCGCCGA ACATTTGTCC AAAACAAACA 1140
     GTGGAAGATA AAAAAAAAGA GGAAGAAGAA GAAACTTGTA CACCGGCATC ACCAGTACCA 1200
20
     GAAAAACCGG TACCTCATGT GGCACGTTGG CGAACATTTA CACCACCTGA GGTATTCAAG 1260
     ATATGGAGGG GAAGGAGAA TAAAACTACG TGCGAAATAG TGGCAGAAAT GCTTAAAGAT 1320
     AAGAATGGAA GGACTACAGT AGGTGAATGT TATAGAAAAG AAACTTATTC TGAATGGACG 1380
     TGTGATGAAA GTAAGATTAA AATGGGACAG CATGGAGCAT GTATTCCTCC AAGAAGACAA 1440
     AAATTATGTT TACATTATTT AGAAAAAATA ATGACAAATA CAAATGAATT GAAATACGCA 1500
25
     TTTATTAAAT GTGCTGCAGC AGAAACTTTT TTGTTATGGC AAAACTACAA AAAAGATAAG 1560
     AATGGTAATG CAGAAGATCT CGATGAAAAA TTAAAAGGTG GTATTATCCC CGAAGATTTT 1620
     AAACGGCAAA TGTTCTATAC GTTTGCAGAT TATAGAGATA TATGTTTGGG TACGGATATA 1680
     TCATCAAAAA AAGATACAAG TAAAGGTGTA GGTAAAGTAA AATGCAATAT TGATGATGTT 1740
30
     TTTTATAAAA TTAGCAATAG TATTCGTTAC CGTAAAAGTT GGTGGGAAAC AAATGGTCCA 1800
     GTTATATGGG AAGGAATGTT ATGCGCTTTA AGTTATGATA CGAGCCTAAA TAATGTTAAT 1860
     CCGGAAACTC ACAAAAAACT TACCGAAGGC AATAACAACT TTGAGAAAGT CATATTTGGT 1920
     AGTGATAGTA GCACTACTTT GTCCAAATTT TCTGAAAGAC CTCAATTTCT AAGATGGTTG 1980
     ACTGAATGGG GAGAAAATTT CTGCAAAGAA CAAAAAAAGG AGTATAAGGT GTTGTTGGCA 2040
35
     AAATGTAAGG ATTGTGATGT TGATGGTGAT GGTAAATGTA ATGGAAAATG TGTTGCGTGC 2100
     AAAGATCAAT GTAAACAATA TCATAGTTGG ATTGGAATAT GGATAGATAA TTATAAAAAA 2160
     CAAAAAGGAA GATATACTGA GGTTAAAAAA ATACCTCTGT ATAAAGAAGA TAAAGACGTG 2220
     AAAAACTCAG ATGATGCTCG CGATTATTTA AAAACACAAT TACAAAATAT GAAATGTGTA 2280
     AATGGAACTA CTGATGAAAA TTGTGAGTAT AAGTGTATGC ATAAAACCTC ATCCACAAAT 2340
40
     AGTGATATGC CCGAATCGTT GGACGAAAAG CCGGAAAAGG TCAAAGACAA GTGTAATTGT 2400
     GTACCTAATG AATGCAATGC ATTGAGTGTA AGTGGTAGCG GTTTTCCTGA TGGTCAAGCT 2460
     TACGTACGCG TGCATGCGAC GTCATAGCTC TTCTATAGTG TCACCTAAAT TCAATTCACT 2520 GGCCGTCGTT TTACAACGTC GTGACTGGGA AAACCTGGCG TTACCCAACT TAATCGCCTT 2580
     GCAGCACATC CCCCTTTCGC CAGCTG
45
     (2) INFORMATION FOR SEO ID NO:8:
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 921 amino acids
50
                (B) TYPE: amino acid
                (C) STRANDEDNESS: single
```

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

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(vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Lys Leu Asn Ser Val Tyr Thr Ser Ile Gly Leu Phe Leu Asn Ser Leu 1 5 10 15

		Lvs	Ser	Glv	Pro	Cvs	Lvs	Lvs	Asp	Asn	Asn	Δen	λla	Glu	Aen	Acn	Tla
					20					25	,				3	0	
_				35					40)				Asp 4	5		_
5			50					55					6	Asn 0			
		Thr 65	Lys	Gly	Lys	Сув	Asn 70	Gly	Ser	Asn	Gly	Lys 75	Lys	Asn	Gly	Asn	Asp 80
10		Tyr	Ile	Thr	Ala	Ser 85	Asp	Ile	Glu	Asn	Gly 90		Asn	Ser	Ile		Asn 5
		Ile	Asp	Met	Val	Val	Ser	Asp	Lys	Asp 10		Asn	Gly	Phe	_	Gly 10	Leu
				115					12	0					25		
15		Trp	Lys 130		Ala	Lys	Val	Cys 13		Leu	Asp	Val		Gly	Leu	Lys	Asn
		Gly			Ser	Ile				Gln	Lys			10 Ile	Ile	Ile	Arg
		145 Ala	Leu	Leu	Lys	Arq	150 Trp	Val	Glu	Tvr	Phe	155 Leu	Glu	Asp	Tvr	Asn	160
20						165	5				17	0				1	75.
					180	1				18	5			Asn	1	90	
05				195					20	0					05	_	
25			210					21	5				22	His 20			
	,	G1n 225	Asn	Glu	Asn	Gly	Asp 230	Asn	Asn	Met	Lys	Ser 235	Leu	Val	Thr	Asp	Ile 240
30		Leu	Gly	Ala	Leu	Gln	Pro	Gln	Ser	Asp.		Asn	Lys	Ala	Ile		Pro
30		Cys	Ser	Gly	Leu	245 Thr		Phe	Glu	Ser	25 Phe		Gly	Leu	Asn	Gly	55 Ala
					260					26	5			Leu	2'	70	
ar				275					28	0				28	35		
35			290					29!	5				30	Lys 00		_	
		Gly 305	Glu	Thr	Ser	Val	Glu 310	Asn	Gly	Gly	Lys	Ser 315	Cys	Thr	Pro	Leu	Asp 320
40			Thr	Thr	Leu	Glu 325	Glu	Glu	Pro	Ile	Glu 33	Glu	Glu	Asn	Gln		Glu 35
		Ala	Pro	Asn	Ile 340	Cys		Lys	Gln	Thr 34	Val		Asp	Lys		Lys	
		Glu	Glu	Glu 355			Cys ·	Thr	Pro 36	Ala		Pro	Val	Pro 36	Glu	50 Lys	Pro
45		Val	Pro 370		Val	Ala	Arg	Trp 37!	Arg		Phe	Thr	Pro 38	Pro		Val	Phe
		Lys 385	-	Trp	Arg	Gly	Arg 390			Lys				Glu	Ile		
			Met	Leu	Lys	Asp		Asn	Gly	Arg		395 Thr	Val	Gly	Glu		400 Tyr
50		Arg	Lys	Glu	Thr	405 Tyr		Glu	Trp	Thr	41 Cys		Glu	Ser	Lys		15 Lys
					420					42	5			Gln	43	30	_
55				435	•				44	D				44	15		-
33			450	•				455	5				46			-	_
		465	Pne	ше	ьуѕ		A1a 470	Ата	AIA	GIU		Pne 475	Leu	Leu	Trp		Asn 480
60		Tyr	Lys	Lys	Asp			Gly	Asīī	Alā		Asp	ŗen	qaƙ	Glu	ъуs	
		Lys	Gly	Gly	Ile 500		Pro	Glu	Asp	Phe 50	Lys	-	Gln	Met	_	Tyr	
		Phe	Ala	Asp		Arg	Asp	Ile	Cys			Thr	Asp	Ile		l0 Ser	Lys
				515					520		-		_	52	_		-

		530)				53	5				5	40		Asp	
	Val 545	Phe	Tyr	Lys	Ile	Ser 550	Asn	Ser	Ile	Arg	Tyr 555	Arg	Lys	Ser	Trp	Trp 560
5	Glu	Thr	Asn	Gly	Pro 56	Val	Ile	Trp	Glu			Leu	Cys	Ala	Leu	Ser 575
	Tyr	Asp	Thr	Ser 580	Leu	Asn	Asn	Val	Asn 58	Pro	Glu	Thr	His	_	Lys	Leu
10	Thr	Glu	Gly 595	Asn	Asn	Asn	Phe	Glu 60	Lys		Ile	Phe		Ser 05	Asp	Ser
	Ser	Thr 610	Thr	Leu	Ser	Lys	Phe 61	Ser		Arg	Pro		Phe 20	Leu	Arg	Trp
	Leu 625	Thr	Glu	Trp	Gly	Glu 630			Cys	Lys	Glu 635	Gln	Lys	Lys	Glu	Tyr
15	Lys	Val	Leu	Leu	Ala 645	Lys	Cys	Lys	Asp	Cys 65	Asp	Val	qaA	Gly	Asp	Gly 55
	Lys	Cys	Asn	Gly 660	Lys	-	Val	Ala	Cys 66	Lys		Gln	Cys		Gln 70	Tyr
20	His	Ser	Trp 675	Ile		Ile	Trp	Ile 68	Asp		Tyr	Lys		Gln 85	Lys	Gly
	Arg	Tyr 690	Thr	Glu	Val	Lys	Lys 69	Ile		Leu	Tyr				Lys	Asp
	Val 705	Lys	Asn	Ser	Asp	Asp 710	Ala	Arg	Asp	Tyr	Leu 715			Gln	Leu	Gln 720
25					725	•				73	10				Tyr 7	Lys 35
				740					74	5				7	Ser	Leu
30			755					76	0				7	65	Pro	
		770					77	5				78	30	_	Gly	
. -	785					790					795				Gly	800
35					805	i				81	.0				Asp 8	15
				820					82	5				8	Ser 30	
40			835					84	0				84	15	Ile	
		850					855	5				86	50		Ile	_
4.5	865					870					875				Val	880
45					885					89	0			_	Ile 8	9Š
				900					90	5	Lys	Met	Lys		Met 10	Lys
50	Lys	Met	Lys 915	Lys	Arg	Lys	Lys	920		e [.]				•		,

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2101 base pairs
(B) TYPE: nucleic acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- 60 (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO

55

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

5	GGAACAGGGT GATAATAAAG TAGGAGCCTG TGCTCCGTAT AGACGATTAC ATTTATGTGA 60
_	TTATAATTTG GAATCTATAG ACACAACGTC GACGACGCAT AAGTTGTTGT TAGAGGTGTG 120
	TATGGCAGCA AAATACGAAG GAAACTCAAT AAATACACAT TATACACAAC ATCAACGAAC 180
	TAATGAGGAT TCTGCTTCCC AATTATGTAC TGTATTAGCA CGAAGTTTTG CAGATATAGG 240
	TGATATCGTA AGAGGAAAAG ATCTATATCT CGGTTATGAT AATAAAGAAA AAGAACAAAG 300
10	AAAAAAATTA GAACAGAAAT TGAAAGATAT TTTCAAGAAA ATACATAAGG ACGTGATGAA 360
••	GACGAATGGC GCACAAGAAC GCTACATAGA TGATGCCAAA GGAGGAGATT TTTTTCAATT 420
	AAGAGAAGAT TGGTGGACGT CGAATCGAGA AACAGTATGG AAAGCATTAA TATGTCATGC 480
	ACCAAAAGAA GCTAATTATT TTATAAAAAC AGCGTGTAAT GTAGGAAAAG GAACTAATGG 540
	TCAATGCCAT TGCATTGGTG GAGATGTTCC CACATATTTC GATTATGTGC CGCAGTATCT 600
15	TCGCTGGTTC GAGGAATGGG CAGAAGACTT TTGCAGGAAA AAAAAAAAA AACTAGAAAA 660
	TTTGCAAAAA CAGTGTCGTG ATTACGAACA AAATTTATAT TGTAGTGGTA ATGGCTACGA 720
	TTGCACAAAA ACTATATATA AAAAAGGTAA ACTTGTTATA GGTGAACATT GTACAAACTG 780
	TTCTGTTTGG TGTCGTATGT ATGAAACTTG GATAGATAAC CAGAAAAAAG AATTTCTAAA 840
	ACAAAAAAGA AAATACGAAA CAGAAATATC AGGTGGTGGT AGTGGTAAGA GTCCTAAAAG 900
20	GACAAAACGG GCTGCACGTA GTAGTAGTAG TAGTGATGAT AATGGGTATG AAAGTAAATT 960
	TTATAAAAAA CTGAAAGAAG TTGGCTACCA AGATGTCGAT AAATTTTTAA AAATATTAAA 1020
	CAAAGAAGGA ATATGTCAAA AACAACCTCA AGTAGGAAAT GAAAAAGCAG ATAATGTTGA 1080
	TTTTACTAAT GAAAAATATG TAAAAACATT TTCTCGTACA GAAATTTGTG AACCGTGCCC 1140
	ATGGTGTGGA TTGGAAAAAG GTGGTCCACC ATGGAAAGTT AAAGGTGACA AAACCTGCGG 1200
25	AAGTGCAAAA ACAAAGACAT ACGATCCTAA AAATATTACC GATATACCAG TACTCTACCC 1260
	TGATAAATCA CAGCAAAATA TACTAAAAAA ATATAAAAAT TTTTGTGAAA AAGGTGCACC 1320
	TGGTGGTGGT CAAATTAAAA AATGGCAATG TTATTATGAT GAACATAGGC CTAGTAGTAA 1380
	AAATAATAAT AATTGTGTAG AAGGAACATG GGACAAGTTT ACACAAGGTA AACAAACCGT 1440
	TAAGTCCTAT AATGTTTTTT TTTGGGATTG GGTTCATGAT ATGTTACACG ATTCTGTAGA 1500
30	GTGGAAGACA GAACTTAGTA AGTGTATAAA TAATAACACT AATGGCAACA CATGTAGAAA 1560
	CAATAATAAA TGTAAAACAG ATTGTGGTTG TTTTCAAAAA TGGGTTGAAA AAAAACAACA 1620
	AGAATGGATG GCAATAAAAG ACCATTTTGG AAAGCAAACA GATATTGTCC AACAAAAAGG 1680
	TCTTATCGTA TTTAGTCCCT ATGGAGTTCT TGACCTTGTT TTGAAGGGCG GTAATCTGTT 1740
05	GCAAAATATT AAAGATGTTC ATGGAGATAC AGATGACATA AAACACATTA AGAAACTGTT 1800
35	GGATGAGGAA GACGCAGTAG CAGTTGTTCT TGGTGGCAAG GACAATACCA CAATTGATAA 1860
	ATTACTACAA CACGAAAAAG AACAAGCAGA ACAATGCAAA CAAAAGCAGG AAGAATGCGA 1920
	GAAAAAAGCA CAACAAGAAA GTCGTGGTCG CTCCGCCGAA ACCCGCGAAG ACGAAAGGAC 1980
	ACAACAACCT GCTGATAGTG CCGGCGAAGT CGAAGAAGAA GAAGACGACG ACGACTACGA 2040
40	CGAAGACGAC GAAGATGACG ACGTAGTCCA GGACGTAGAT GTAAGTGAAA TAAGAGGTCC 2100
40	G 2101

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 700 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- 50 (ii) MOLECULE TYPE: protein

45

- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: Plasmodium falciparum
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:
- Glu Glu Gly Asp Asn Lys Val Gly Ala Cys Ala Fro Tyr Arg Arg Leu

 1 5 10 15

 His Leu Cys Asp Tyr Asn Leu Glu Ser Ile Asp Thr Thr Ser Thr Thr
 20 25 30

 His Lys Leu Leu Leu Glu Val Cys Met Ala Ala Lys Tyr Glu Gly Asn
 35 40 45

	Ser	Ile	Asn	Thr	His	Tyr	Thr	Gln	His	Gln	Arg			Glu	Asp	Ser
	Ala 65	Ser	Gln	Leu	Cys		Val	Leu	Ala	Arg				Asp	Ile	Ξ.
5		Ile	Val	Arg		70 Lys	Asp	Leu	Tyr		75 Gly		Asp	Asn		
	Lys	Glu	Gln	Arg		Lys	Leu	Glu	Gln 10		Leu	Lys	Asp		_	
10	Lys	Ile	His 115	Lys		Val	Met	Lys 120	Thr		Gly	Ala				Tyr
	Ile	Asp 130	Asp	Ala	Lys	Gly	Gly 135	Asp	-	Phe	Gln	Leu 14	_		Asp	Trp
	Trp			Asn	Arg	Glu 150			Trp	Lys	Ala 155			Cys	His	Ala 160
15		Lys	Glu	Ala	Asn 165	Tyr	Phe	Ile	Lys	Thr	Ala	Cys	Asn	Val	Gly	Lys
	Gly	Thr	Asn	Gly 180		Cys	His	Сув	Ile 18		Gly	Asp	Val	Pro	Thr	
20	Phe	Asp	Tyr 195	Val	Pro	Gln	Tyr	Leu 200		Trp	Phe	Glu	Glu 20	Trp		Glu
		210		Arg			215	5				22	0		_	
	Cys 225	Arg	Asp	Tyr	Glu	Gln 230	Asn	Leu	Tyr	Cys	Ser 235	Gly	Asn	Gly	Tyr	Asp 240
25				Thr	245	,			_	25	0		•	_	25	55
				Cys 260					26	5				27	0	_
30			275					280)	-	_	-	28	5		
		290		Gly			295	;				30	0	=	_	
25	305			Ser		310					315				_	320
35				Leu	325	,				33	0				33	5
				Asn 340					34	5		•		35	0	
40			355					360)	•			36	5		_
		370		Arg			375	•			_	38	0 _		_	
45	385			Gly		390					395					400
43				Thr Pro	405					41	0				41	.5
				420 Glu					42	5			_	43	0	_
50			435					440)		•		44	5		
		450		Gly			455	•				46	0			
55	465			Asn		470					475		_			480
33				Glu	485	,				49	0				49	5
				500 Asn					50	5				51	0	
60			515					520)				52	5		_
		530		Gln			535	,				54	0	-		
	545	пув	vəħ	His	FIIE	550	цys	GIII	TILL	nap	555	val	GIII	GIII	пλг	560

Leu Ile Val Phe Ser Pro Tyr Gly Val Leu Asp Leu Val Leu Lys Gly 565 570 Gly Asn Leu Leu Gln Asn Ile Lys Asp Val His Gly Asp Thr Asp Asp 580 5 Ile Lys His Ile Lys Lys Leu Leu Asp Glu Glu Asp Ala Val Ala Val 600 Val Leu Gly Gly Lys Asp Asn Thr Thr Ile Asp Lys Leu Leu Gln His 615 Glu Lys Glu Gln Ala Glu Gln Cys Lys Gln Lys Gln Glu Glu Cys Glu 10 630 635 Lys Lys Ala Gln Gln Glu Ser Arg Gly Arg Ser Ala Glu Thr Arg Glu 645 650 Asp Glu Arg Thr Gln Gln Pro Ala Asp Ser Ala Gly Glu Val Glu Glu 660 665 Glu Glu Asp Asp Asp Asp Tyr Asp Glu Asp Asp Glu Asp Asp Asp Val 15 680 Val Gln Asp Val Asp Val Ser Glu Ile Arg Gly Pro 690 695 20 (2) INFORMATION FOR SEO ID NO:11: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 8220 base pairs (B) TYPE: nucleic acid 25 (C) STRANDEDNESS: single (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: DNA (genomic)
- 30 (iii) HYPOTHETICAL: NO
 - (vi) ORIGINAL SOURCE: (A) ORGANISM: Plasmodium falciparum
- 35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAAAATGGGG CCCAAGGAGG CTGCAGGTGG GGATGATATT GAGGATGAAA GTGCCAAACA 60 TATGTTTGAT AGGATAGGAA AAGATGTGTA CGATAAAGTA AAAGAGGAAG CTAAAGAACG 120 TGGTAAAGGC TTGCAAGGAC GTTTGTCAGA AGCAAAATTT GAGAAAAATG AAAGCGATCC 180 40 ACAAACACCA GAAGATCCAT GCGATCTTGA TCATAAATAT CATACAAATG TAACTACTAA 240 TGTAATTAAT CCGTGCGCTG ATAGATCTGA CGTGCGTTTT TCCGATGAAT ATGGAGGTCA 300 ATGTACACAT AATAGAATAA AAGATAGTCA ACAGGGTGAT AATAAAGGTG CATGTGCTCC 360 ATATAGGCGA TTGCATGTAT GCGATCAAAA TTTAGAACAG ATAGAGCCTA TAAAAATAAC 420 AAATACTCAT AATTTATTGG TAGATGTGTG TATGGCAGCA AAATTTGAAG GACAATCAAT 480 45 AACACAAGAT TATCCAAAAT ATCAAGCAAC ATATGGTGAT TCTCCTTCTC AAATATGTAC 540 TATGCTGGCA CGAAGTTTTG CGGACATAGG GGACATTGTC AGAGGAAGAG ATTTGTATTT 600 AGGTAATCCA CAAGAAATAA AACAAAGACA ACAATTAGAA AATAATTTGA AAACAATTTT 660 CGGGAAAATA TATGAAAAAT TGAATGGCGC AGAAGCACGC TACGGAAATG ATCCGGAATT 720 TTTTAAATTA CGAGAAGATT GGTGGACTGC TAATCGAGAA ACAGTATGGA AAGCCATCAC 780 50 ATGTAACGCT TGGGGTAATA CATATTTTCA TGCAACGTGC AATAGAGGAG AACGAACTAA 840 AGGTTACTGC CGGTGTAACG ACGACCAAGT TCCCACATAT TTTGATTATG TGCCGCAGTA 900 TCTTCGCTGG TTCGAGGAAT GGGCAGAAGA TTTTTGTAGG AAAAAAATA AAAAAATAAA 960 AGATGTTAAA AGAAATTGTC GTGGAAAAGA TAAAGAGGAT AAGGATCGAT ATTGTAGCCG 1020 TAATGGCTAC GATTGCGAAA AAACTAAACG AGCGATTGGT AAGTTGCGTT ATGGTAAGCA 1080 55 ATGCATTAGC TGTTTGTATG CATGTAATCC TTACGTTGAT TGGATAAATA ACCAAAAAGA 1140 ACAATTTGAC AAACAGAAAA AAAAATATGA TGAAGAAATA AAAAAATATG AAAATGGAGC 1200 ATCAGGTGGT AGTAGGCAAA AACGGGATGC AGGTGGTACA ACTACTACTA ATTATGATGG 1260 ATATGAAAAA AAATTTTATG ACGAACTTAA TAAAAGTGAA TATAGAACCG TTGATAAATT 1320 TITGGAAAAA TTAAGTAATG AAGAAATTG CACAAAAGTT AAAGACGAAG AAGGAGGAAC 1360 60 AATTGATTTT AAAAACGTTA ATAGTGATAG TACTAGTGGT GCTAGTGGCA CTAATGTTGA 1440 AAGTCAAGGA ACATTTTATC GTTCAAAATA TTGCCAACCC TGCCCTTATT GTGGAGTGAA 1500 AAAGGTAAAT AATGGTGGTA GTAGTAATGA ATGGGAAGAG AAAAATAATG GCAAGTGCAA 1560 GAGTGGAAAA CTTTATGAGC CTAAACCCGA CAAAGAAGGT ACTACTATTA CAATCCTTAA 1620 AAGTGGTAAA GGACATGATG ATATTGAAGA AAAATTAAAC AAATTTTGTG ATGAAAAAAA 1680

	TGGTGATACA	ATAAATAGTG	GTGGTAGTGG	TACGGGTGGT	AGTGGTGGTG	GTAACAGTGG	1740
	TAGACAGGAA	TTGTATGAAG	AATGGAAATG	TTATAAAGGT	GAAGATGTAG	TGAAAGTTGG	1800
	ACACGATGAG	GATGACGAGG	AGGATTATGA	AAATGTAAAA	AATGCAGGCG	GATTATGTAT	1860
	ATTAAAAAAC	CAAAAAAAGA	ATAAAGAAGA	AGGTGGAAAT	ACGTCTGAAA	AGGAGCCTGA	1920
5	TGAAATCCAA	AAGACATTCA	ATCCTTTTTT	TTACTATTGG	GTTGCACATA	TGTTAAAAGA	1980
	TTCCATACAT	TGGAAAAAAA	AACTTCAGAG	ATGTTTACAA	AATGGTAACA	GAATAAAATG	2040
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	AGACGAATGG	GGGAAAATAG	TACAACATTT	TAAAACGCAA	AATATTAAAG	GTAGAGGAGG	2160
	TAGTGACAAT	ACGGCAGAAT	TAATCCCATT	TGATCACGAT	TATGTTCTTC	מיידית מיינית מ	2220
10	GCAAGAAGAA	TTTTTGAAAG	GCGATTCCGA	ACACCCTTCC	GAAGAAAAAT	CCCAAAATII	2220
	TCTGGATGCA	GAGGAGGCAG	AGGAACTAAA	ACACCTTCGC	GAAATCATTG	ANACTONAMIAG	2240
	СВОТВОТСВВ	CAACCATCTC	THE CHICKNEY	COTONOTON	CAAAAAAATA	MANGI GANGA	2340
	ATTICTOANC	TACCAAAAAC	1100100100	CGICACIGAA	GAAATTCACG	TAATGGATAA	2400
	AGAGGAAAAA	CARARAGE	ACCEANGCEA	AMOUNTOCOLA	GAGGGCGAAA	AAGATGAGGA	2460
15	ተል አጥር ርአጥርም	ACTOCOCANA	CTCCTTAACGA	AIGIAICGAA	GAGGGGGAAA	ATTTTCGTTA	2520
	CUNTCONIGI	CATCACAAA	GIGGIAACAA	ACGATACCCC	GTTCTTGCGA	ACAAAGTAGC	2580
	CACACCOCATO	CATCACAAGG	CAAAGACACA	ATTGGCTAGT	CGTGCTGGTA	GAAGTGCGTT	2640
	GAGAGGTGAT	ATATCCTTAG	CGCAATITAA	AAATGGTCGT	AACGGAAGTA	CATTGAAAGG	2700
	ACAAATTIGC	AAAATTAACG	AAAACTATTC	CAATGATAGT	CGTGGTAATA	GTGGTGGACC	2760
20	ATGTACAGGC	AAAGATGGAG	ATCACGGAGG	TGTGCGCATG	AGAATAGGAA	CGGAATGGTC	2820
20	AAATATTGAA	GGAAAAAAAC	AAACGTCATA	CAAAAACGTC	TTTTTACCTC	CCCGACGAGA	2880
	ACACATGTGT	ACATCCAATT	TAGAAAATTT	AGATGTTGGT	AGTGTCACTA	AAAATGATAA	2940
	GGCTAGCCAC	TCATTATTGG	GAGATGTTCA	GCTCGCAGCA	AAAACTGATG	CAGCTGAGAT	3000
	AATAAAACGC	TATAAAGATC	AAAATAATAT	ACAACTAACT	GATCCAATAC	AACAAAAAGA	3060
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25	AGGAAGAGAT	ATGTGGGATG	AGGATAAGAG	CTCAACAGAC	ATGGAAACAC	GTTTGATAAC	3180
	CGTATTTAAA	AACATTAAAG	AAAAACATGA	TGGAATCAAA	GACAACCCTA	AATATACCGG	3240
•	TGATGAAAGC	AAAAAGCCCG	CATATAAAAA	ATTACGAGCA	GATTGGTGGG	AAGCAAATAG	3300
	ACATCAAGTG	TGGAGAGCCA	TGAAATGCGC	AACAAAAGGC	ATCATATGTC	CTGGTATGCC	3360
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	TAAATATAAA	GAGGAAATAG	AAAAATGGAA	TGAACAATGG	AGAAAAATAT	CAGATAAATA	3600
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	CGCCCCGATC	ACCCCCTACA	GTACTGCTGC	CGGATATATA	CACCAGGAAA	TAGGATATGG	3840
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40	TGCCTGCAAA	ATAGTGGAGA	AAATACTTGA	GGGTAAGAAT	GGAAGGACTA	CACTACCTCA	4020
	ATGTAATCCA	AAAGAGAGTT	ATCCTGATTG	GGATTGCAAA	AACAATATTC	ACATTACTCA	4140
	TGATGGTGCT	TGTATGCCTC	CAAGGAGACA	AAAACTATGT	מיימידמידמיים	TAGCACATGA	4200
	GAGTCAAACA	GAAAATATAA	AAACAGACGA	TAATTTGAAA	GATGCTTTTA	TTAAAACTCC	4260
	AGCAGCAGAA	ACTITICTIT	CATGGCAATA	TTATAAGAGT	AAGAATGATA	GTGAAGGTAA	4200
45	AATATTAGAT	AGAGGCCTTA	TTCCATCCCA	מממידידידממ	TCCATGATGT	ACACCITATE CC.	4320 4380
	AGATTATAGA	GATATATGTT	TGAACACAGA	מממיייייתמיימיית	AAACAAAATC	ACACGITIGG	4300
	GGCAAAAGAT	AAAATAGGTA	אסיידיידיידיריירי	AAAAGATGGC	ACCADATCTC	TIGINGCIAN	4540
	ATCACGCCAA	GAATGGTGGA	AAACAAATGG	TCCAGAGATT	TCCANACCAN	CINGIOGCII	4560
	CTTAACAAAA	TACCTCACAC	ATACCGATAA	CANANCANA	ATCANANGCA	A CALL VALUE OF THE PARTY OF TH	4600
50	CGATAAAGTC	AACCAATCCC	AIACCGAIAA	CHAMAGAMAA	AT CHAMMACG	ACIACICATA	4620
00	TCAATTTCTA	CCTTCCATICCC	TOTA ATTOCAM	ACA ACA COCCII	TCTCCTCAAC	CTGCTAAACC	4680
	CCANANTATC	ATTANANCATO	CAMCINATION	MOMMOMOTIT	1G1GC1GAAC	GTCAGAAGAA	4740
	GGAAAATATC	MCMAAAGAIG	CATGTAATGA	AATAAATTCT	ACACAACAGT	GTAATGATGC	4800
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5 5	AGAATATAAA	GGATATGAAT	ATAAAGACGG	CGTACAACCG	ATACAGGGGA	ATGAGTATTT	4980
	ACTGCAAAAA	TGTGATAATA	ATAAATGTTC	TTGCATGGAT	GGAAATGTAC	TTTCCGTCTC	5040
	TCCAAAAGAA	AAACCITTTG	GAAAATATGC	CCATAAATAT	CCTGAGAAAT	GTGATTGTTA	5100
	TCAAGGAAAA	CATGTACCTA	GCATACCACC	TCCCCCCCA	CCTGTACAAC	CACAACCGGA	5160
	ĀĠĊĀĊĊĀĀĊĀ	GTAACAGTAG	ACGTTTĞCA G	CATAGTAAAA	ACACTATTTA	AAGACACAAA	5220
60	CAATTTTTCC	GACGCTTGTG	GTCTAAAATA	CGGCAAAACC	GCACCATCCA	GTTGGAAATG	5280
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	TGGTAGTATT	TGTATCCCAC	CCAGGAGGCG	ACGATTATAT	GTGGGGAAAC	TACAGGAGTG	5400
	GGCTACCGCG	CTCCCACAAG	GTGAGGGCGC	CGCGCCGTCC	CACTCACGCG	CCGACGACTT	5460
	GCGCAATGCG	TTCATCCAAT	CTGCTGCAAT	AGAGACTTTT	TTCTTATGGG	ATAGATATAA	5520
				· · -			

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     AGGTGCTGGT ATCTTTGAAG GTATTAGAAA AGATGAATGG AAATGTCGTA ATGTATGTGG 6900
     TTATGTTGTA TGTAAACCGG AAAACGTTAA TGGGGAAGCA AAGGGAAAAC ACATTATACA 6960
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     AATTAGAGCA CTGGTTAAAC GTTGGGTAGA ATATTTTTTT GAAGATTATA ATAAAATAAA 7020
     ACATAAAATT TCACATCGCA TAAAAAATGG TGAAATATCT CCATGTATAA AAAATTGTGT 7080
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     TCTAACCGCC CTGGTGACCT CCACCCTCGC CTGGAGCGTT GGCATCGGTT TTGCTACATT 7800
     CACTTATTTT TATCTAAAGG TAAATGGAAG TATATATATG GGGATGTGGA TGTATGTGGA 7860
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     TGTATGTGAA TGTATGTGGA TGTATGTGGA TGTATGTGTAT GGATATGTAT 7920
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     TATATGGATG TACTTGTATG TGTTTTATAT ATATATTTA TATATATGTA TTTATATTAA 8100
     ΑΑΑΑGΑΑΤΑ ΤΑΑΑΑΑCAAA ΤΙΤΑΤΤΑΑΑΑ ΤGΑΑΑΑΑΑG ΑΑΑΑΑΤGΑΑΑ ΤΑΤΑΑΑΑΑΑ 8160
45
     ΑΑΤΤΤΑΤΤΑΑ ΑΑΤΑΑΑΑΑΑΑ ΑΑΑΑΑΑΑΑΑΑ ΑΑΑΑGGAGAA ΑΑΑΤΤΤΤΤΤΑ ΑΑΑΑΑΤΑΑΤΑ 8220
```

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2710 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO

50

55

60

- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Plasmodium falcinarum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Asn Val Met Val Glu Leu Ala Lys Met Gly Pro Lys Glu Ala Ala Gly

	1				5					10					19	5
				20					25				Phe	30)	
5	Gly	Lys	Asp 35	Val	Tyr	Asp	Lys	Val	Lys	Glu	Glu	Ala	Lys 45	-	Arg	Gly
	Lys	Gly 50	Leu	Gln	Gly	Arg	Leu 55	Ser	Glu	Ala	Lys	Phe 60	Glu	Lys	Asn	Glu
	Ser 65	Asp	Pro	Gln	Thr	Pro	Glu	Asp	Pro	Cys	Asp 75	Leu	Asp	His	Lys	Tyr
10	His	Thr	Asn	Val	Thr 85	Thr	Asn	Val	Ile	Asn 90		Cys	Ala	Asp	Arg	_
				100)				10	5			Thr	11	Asn 0	Arg
15			115					12	0				Cys 12	5		_
		130					135	5				14				
•	Lys 145	Ile	Thr	Asn	Thr	His 150	Asn	Leu	Leu	Val		Val	Cys	Met	Ala	
20		Phe	Glu	Gly	Gln 165	Ser	Ile	Thr	Gln	Asp		Pro	Lys	Tyr	_	
	Thr	Tyr	Gly	Asp 180	Ser		Ser	Gln	Ile 18	Cys		Met	Leu	Ala 19		Ser
25	Phe	Ala	Asp 195	Ile	Gly	Asp	Ile	Val 20	Arg		Arg	Asp	Leu 20	Tyr	Leu	Gly
		210					215	5				22				_
••	225					230					235		Ala			240
30					245	5				25	0		Asp		25	55
				260	•				26	5			Asn	27	0	_
35	Asn	Thr	1yr 275		Hls	Ala	Thr	Cys 280		Arg	Gly	Glu	Arg 28		Lys	Gly
		290					295	5				30	Phe 0	Asp	-	
	Pro 305	Gln	Tyr	Leu	Arg	Trp 310	Phe	Glu	Glu	Trp		Glu	Asp	Phe	Cys	
40		Lys	Asn	Lys	Lys 325	Ile	Lys	Asp	Val	Lys 33		Asn	Cys	Arg	Gly 33	
	Asp	ГÀв	Glu	Asp 340	Lys	Asp	Arg	Tyr	Cys 34	Ser		Asn	Gly	Tyr 35	Asp	
45	Glu	Lys	Thr 355	Lys	Arg	Ala	Ile	Gly 360		Leu	Arg	Tyr	Gly 36	Lys		Cys
		370					375	•				38				
	Gln 385	Lys	Glu	Gln	Phe	Asp 390	Lys	Gln	Lys	Lys	Lys 395	Tyr	Asp	Glu	Glu	
50		Lys	Tyr	Glu	Asn 405	Gly	Ala	Ser	Gly	Gly 410	Ser	Arg	Gln	Lys	Arg 41	
	Ala	Gly	Gly	Thr 420	Thr		Thr	Asn	Tyr 425	Asp		Tyr	Glu	Lys 43	Lys	Phe
55	Tyr	Ąsp	Glu 435	Leu	Asn	Lys	Ser	Glu 440		Arg	Thr	Val	Asp	Lys		Leu
	Glu	Lys 450	Leu	Ser	Asn	Glu	Glu 455		Cys	Thr	Lys	Val 46	Lys 0	Asp	Glu	Glu
•	Gly 465	Gly	Thr	Ile	Asp	Phe 470	Lys	Asn	Val		Ser 475	Asp	Ser	Thr		Gly 480
60					485					Gly 490	Thr		Tyr	_	Ser 49	Lys 5
				500					505	5 .			Val	51	Asn 0	Gly
	Gly	Ser	Ser	Asn	Glu	Trp	Glu	Glu	Lys	Asn	Asn	Gly	Lys	Cys	Lys	Ser

			515	5				52	0			٠	51	25		
		/ Lys 530	,				53	Pro	Asp			54	Thr	Thr		
5	545					550					555	Glu	Glu			560
		Phe			56	5				57	0				ξ.	75
40		Thr		581)				58	5				5.5	90	
10		Glu	595					60	0				60)5		
		Glu 610					61	5				62	0			_
15	625					630					635					640
		Ser			645	5				65	0				65	55
20		Tyr		660)'				66	5				67	70	_
		Lys	675					68	0				68	5	_	_
		Asn 690					695	5				70	0			
25	705					710					715			-		720
		Ile Asp			725					73	0				73	15
		·		740	-1-	vul	cu	GIII	74		Deu	GIII	GIU	75		Leu
30		Gly	755					760)				76	Asn 5	Ser	
	•	Ala 770					775	5				78	0			
35	785	Glu -				790					795					800
		Lys			805	•				81	0				81	.5
	Asp	Leu	Cys	ьеи 820	GIU	шe	His	Glu	Asp 825		Glu	Glu	Glu	Lys 83		Lys
40		Asp	835	Asn	Glu			840	Glu	Gly			84	Arg 5	Tyr	
		Cys 850					855	,				86	0			
45	865	Val				870					875					880
	Arg	Ala	GIA	Arg	Ser 885	Ala	Leu	Arg	Gly	Asp 890		Ser	Leu	Ala		
	Lys	Asn	Gly	Arg 900			Ser	Thr	Leu 905	Lys		Gln	Ile	Cys 91		Ile
50	Asn ·	Glu	Asn 915	Tyr	Ser	Asn	Asp	Ser 920	Arg		Asn	Ser	Gly 92:	Gly	Pro	Cys
	Thr	Gly 930	Lys	Asp	Gly	Asp	His 935		Gly	Val	Arg	Met 94	Arg		Gly	Thr
55	Glu 945	Trp	Ser	Asn	Ile	Glu 950	Gly	Lys	Lys	Gln		Ser	Tyr	Lys		
		Leu	Pro	Pro	Arg 965	Arg	Glu	His	Met	Cys 970		Ser	Asn	Leu	Glu	
	Leu	Asp	Val	Gly 980			Thr	Lys	Asn 285	Asp		Ala	Ser	His		Leu
60	_															
		Gly	995					100	0				100	05		
	Lys	Arg 1010	Tyr)	Lys	Asp	Gln	Asn 101		Ile	Gln	Leu	Thr 102		Pro	Ile	Gln

	01 m	T	3	a 1	a 1		M = 4	~	.			_	_	_		
	102	ьув 5	Asp	GIN	GIU	103		Cys	arg	АТА	Val 103!		Tyr	Ser	Phe	Ala 1040
			Gly	Asp	Ile 104	Ile	Arg	Gly	Arg				Asp	Glu		Lys)55
5	Ser	Ser	Thr	Asp	Met		Thr	Arg	Leu 10	Ile		Val	Phe		Asn 070	Ile
	Lys	Glu	Lys 107	His		Gly	Ile	Lys 10	Asp		Pro	Lys		Thr 85	Gly	Asp
10	Glu	Ser	Lys		Pro	Ala	Tyr	Lys		Leu	Arg		Asp	Trp	Trp	Glu
	Ala 110	Asn		His	Gln		Trp		Ala	Met		Сув	Ala	Thr	Lys	_
			Cys	Pro	Gly 112	1110 Met	Pro	Val	Asp	Asp			Pro	Gln		
15	Arg	Trp	Met	Thr 114	Glu		Ala	Glu	Trp	Tyr		Lys	Ala		Ser .50	.35 Gln
	Glu	Tyr	Asp 115	Lys		Lys	Lys	Ile 116	Cys		Asp	Сув		Ser 65	Lys	Gly
20	Asp	Gly 117	Lys		Thr	Gln	Gly 117	Asp		Asp	Cys	Gly 11	Lys	Cys	Lys	Ala
	Ala 1185		Asp	Lys	Tyr	Lýs 1190	Glu D	Glu	Ile	Glu	Lys 1199	Trp		Glu	Gln	Trp 1200
					120	5	Tyr.			12	10				12	Thr
25				122	0		Arg		12:	25				12	30	_
			123	5			Phe	124	10				12	45		
30		125	0				Lys 125	55				12	60			
	1265	5				1270					1275	;		_	-	1280
35	His				128	5				129	90				12	95
33	Glu			130	0				130	05				13	10	-
	Glu		131	5			Thr	132	20				13	25		
40	Val	133	0				133	5			_	13	4 Õ			
	1345 Gly	5				1350)			_	1355	;		_	-	1360
45					136	5	Ile			13'	70			_	13	75
				138	0		Leu		138	35				13	9Ō	
	Gln		139	5				140	0				14	05		
50	Lys	141)				141	.5			•	14	20			
	1425 Lys	•				1430)				1435			_	_	1440
55	Gln				144	5				145	50				14	55
	Cys			146	0				146	55				14	70	í
•	гуs		147	5				148	30			_	14	85	_	
60		1490)				149	5			_	15	00	_		
	Ser 1505	5				1510)				1515	,	_			1520
	Trp	Lys	Gly	Met	Leu 152		Ala	Leu	Thr	Lys 153		Val	Thr	Asp		Asp 35

	Asn	Lys	Arg	Lys 154		Lys	Asn		Tyr 15		Tyr	Asp	Lys		Asn 550	Gln
	Ser	Gln	Asn 155		Asn	Pro	Ser	Leu 15		Glu	Phe	Ala		Lys	Pro	Gln
5	Phe	Leu 157	Arg	Trp	Met	Ile	Glu 15	Trp		Glu	Glu				Glu	Arg
	Gln 158	Lys	Lys	Glu	Asn	Ile 159	Ile		Asp	Ala	Cys 1599	Asn		Ile	Asn	
10			Gln	Cys	Asn 160	Asp		Lys	His	Arg 16	Cys		Gln	Ala	_	_
	Ala	Tyr	Gln	Glu 162	Tyr		Glu	Asn	Lys 16	Lys		Glu	Phe		Gly	Gln
	Thr	Asn	Asn 163	Phe		Leu	Lys	Ala 164	Asn		Gln	Pro		Asp	Pro	Glu
15	Tyr	Lys 165	Gly		Glu	Tyr	Lys 16	Asp		Val	Gln			Gln	Gly	Asn
· ·	Glu 166	Tyr	Leu	Leu	Gln	Lys 1670	Cys		Asn	Asn	Lys 1679	Cys		Cys	Met	_ = = .
20			Val	Leu	Ser 168	Val		Pro	Lys	Glu 16	Lys		Phe	Gly		
	Ala	His	Lys	Tyr 170	Pro		Lys	Cys	Asp	Cys		Gln	Gly		His	Val
	Pro	Ser	Ile 171	Pro		Pro	Pro	Pro	Pro		Gln	Pro		Pro	Glu	Ala
25	Pro	Thr 173	Val		Val	Asp	Val	Cys		Ile	Val		Thr	25 Leu	Phe	Lys
•	Asp 1749	Thr	Asn	Asn	Phe	Ser 1750	Asp		Cys	Gly	Leu	Lys	40 Tyr	Gly	Lys	
30			Ser	Ser	Trp	Lys		Ile	Pro				Lys	Ser		
00	Gly	Ala	Thr	Thr 178	Gly		Ser	Gly				Gly	Ser		Cys	75 Ile
	Pro	Pro	Arg 179	Arg		Arg	Leu	Tyr 180			Lys	Leu		Glu	90 Trp	Ala
35	Thr	Ala 181	Leu		Gln	Gly	Glu 181	Gly		Ala	Pro	Ser 18	His	05 Ser	Arg	Ala
	Asp 1825	Asp	Leu	Arg	Asn	Ala 1830	Phe		Gln	Ser	Ala 1835	Ala		Glu		
40			Trp	Asp	Arg 184	Tyr		Glu	Glu	Lys 18	Lys	Pro	Gln	Gly	Asp	1840 Gly 555
	Ser	Gln	Gln	Ala 186	Leu		Gln	Leu	Thr 18	Ser		Tyr	Ser		Asp :70	Glu
	Glu	Asp	Pro 187	Pro		Lys	Leu	Leu 188	${\tt Gln}$		Gly	Lys		Pro 85	Pro	Asp
45	Phe	Leu 189	Arg 0	Leu	Met	Phe	Tyr	Thr	Leu	Gly	Asp	Tyr	Arg	Asp	Ile	Ļeu
	Val 1905	His	Gly	Gly	Asn	Thr 1910	Ser	Asp	Ser	Gly	Asn 1915	Thr	Asn	Gly	Ser	Asn 1920
50			Asn	Ile	Val 192	Leu		Ala	Ser	Gly 19:	Asn		Glu	Asp	Met	
	Lys	Ile	Gln	Glu 194	Lys		Glu	Gln	Ile 194	Leu		Lys	Asn	_	Gly 50	Thr
	Pro	Leu	Val 195	Pro		Ser	Ser	Ala 196	Gln		Pro	Asp				Asn
55	Glu	His 197	Ala	-	Ser	Ile	Trp 197	Lys		Met	Ile	Cys 19	Ala		Thr	Tyr
	Thr 1985	Glu	Lys	Asn	Pro	Asp 1990	Thr		Ala	Arg	Gly 1995	Asp		Asn		Ile 2000
60			Acp	qaA	Glu 200	Val		Clu	Lys	Pho 20	Pho		Ser	Thr	Ala	Asp
	Lys	His	Gly	Thr 202	Ala		Thr	Pro	Thr 202	Gly		Tyr	Lys		Gln	15 Tyr
	Asp	Tyr	Glu 203	Lys		Lys	Leu	Glu	Asp		Ser	Gly	Ala	Lys	30 Thr	Pro

	Ser	Ala	Ser	Ser	Asp	Thr	Pro	Leu	Leu	Ser	asp	Phe	Val	Leu	Ara	Pro
		205	0				20	55				2.0	160			
_	206	5				207	0				207	5				Lys 2080
5	Arg	Lys	His	Lys	Leu 208	Ala 35	Gln	Ile	Lys	His 20	Glu 90	Cys	Lys	Val		Glu 95
	Asn	Gly	Gly	Gly 210	Ser	Arg	Arg	Gly	Gly 21	Ile	Thr	Arg	Gln		Ser	Gly
10	Asp	Gly	Glu 211	Ala		Asn	Glu	Met 21	Leu	Pro	Lys	Asn		Gly	Thr	Val
	Pro	Asp 213	Leu		Lys	Pro	Ser 213	Cys		Lys	Pro		Ser	.25 Ser	Tyr	Arg
	Lys 2145	Trp		Glu	Ser	Lys 215	Gly		Glu	Phe	Glu	Lys	40 Gln	Glu	Lys	
15		-	Gln	Gln	Lys	Asp		Cys	Val		2159 Gly	Ser	Asn	Lys		
	Asn	Gly	Phe	Cys	Glu		Leu	Thr			Ser	Lys	Aļa		Asp	.75 Phe
	Leu	Lvs	Thr	218 Leu		Pro	Cvs	Lvs	211		Asn	Val	Glu	21	90	Th~
20			219	5				220	00				22	05	•	
		221	0				221	.5			Thr	22	20	_	_	
	2225	Leu	rys	Phe	Ser	Val 223	Asn 1	Cys	Lys	Lys	Asp 2235		Cys	Asp		
25	Lys		Thr	Asp	Cys 224	Arg		Lys	Asn	Ser 22	Ile		Ala	Thr	Asp.	
	Glu	Asn	Gly	Val	Asp		Thr	Val		Glu	Met	Arg	Val		Ala	55 As p
30	Ser	Lys	Ser	226 Gly		Asn	Gly				Glu	Asn		Cys	70 Arg	Gly
30	Ala	Gly	227 Ile		Glu	Gly	Ile	228 Arg		Asp	Glu		Lys	85 Cys	Arg	Asn
	Val	229 Cys	_	Tyr	Val	Val	229 Cys		Pro	Glu	Asn	23 Val		Gly	Glu	Ala
35	2305 Lys			His	Ile	2310 Ile		Ile	Arq	Ala	2315 Leu	Val	Lvs	Ara	Trn	2320 Val
					232	5				23:	30				23	35
•	Glu			234	0				234	15				23	50	
40	Arg		235	5				236	0				23	6Š		
		2370	0				237	'5				23	80			
	Phe 2385					2390)				2395					2400
45	Ser	Phe	Leu	Glu	Thr	Leu	Ile	Pro	Gln			Asp	Ala	Asn	Ala	Lys
	Asn	Lys	Val	Ile	240 Lys		Ser	Lys			Asn	Ser	Cys		Cys	15 Ser
	Ala	Ser	Ala	242 Asn		Gln	Asn	Lys	242 Asn		Glu	Tyr	Lys	24: As p	30 Ala	Ile
50	Asp		243	5				244	10		•		24	45		
	Lys :	2450)				245	5				24	60			-
FF	2465					2470)				2475					2480
55	Gln '				248	5				249	90				24	95
	Ala	Lys	Lys	Asn 250		Met	Pro	Lys	1le 250		Glu	Asn	Val	Leu 25		Thr
60	Ala	Glī	Gln 251		qaĀ	Glu	Glу	050		vaī	Pro	AĨa ,	GIu 25:	Asn		Glu
	Glu	Pro 2530	Ala		Thr	Asp		Gly		Glu	Thr	Pro 25	Glu		Thr	Pro
	Val :			Pro	Glu	Glu			Val	Pro	Glu			Pro	Pro	Pro
	2545					2550					2555					2560

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Pro Gln Glu Lys Ala Pro Ala Pro Ile Pro Gln Pro Gln Pro Pro Thr 2565 2570 Pro Pro Thr Gln Leu Leu Asp Asn Pro His Val Leu Thr Ala Leu Val 2580 2585 2590 Thr Ser Thr Leu Ala Trp Ser Val Gly Ile Gly Phe Ala Thr Phe Thr 5 2595 2600 2605 Tyr Phe Tyr Leu Lys Val Asn Gly Ser Ile Tyr Met Gly Met Trp Met 2610 2615 Tyr Val Asp Val Cys Glu Cys Met Trp Met Tyr Val Asp Val Cys Gly 10 2625 2630 2635 2640 Cys Val Leu Trp Ile Cys Ile Cys Asp Tyr Val Trp Ile Tyr Ile Tyr 2645 2650 2655 Ile Tyr Ile Cys Leu Cys Ile Cys Val Phe Gly Tyr Ile Tyr Val Tyr 2660 2665 2670 15 Val Tyr Asp Phe Leu Tyr Met Tyr Leu Trp Val Lys Asp Ile Tyr Ile 2675 2680 Trp Met Tyr Leu Tyr Val Phe Tyr Ile Tyr Ile Leu Tyr Ile Cys Ile 2700 Tyr Ile Lys Lys Glu Ile 20

- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 19124 base pairs

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

35 ATCCTTCTAT TTTCGATTTT TTCATTTTTT TCCAGTATTA ATTTATTTAT TTATTTGTGA 120 TATTTTATAA TATATTATTT AAATGTGTAT TTATATATGT GTTTTATTTT TGTTATTAAT 180 40 TTGAATAATC CGAGCGAAAA AAAATATATA ATCTCATATA AAAATTATTT ATAATACAAT 240 ATTATATAGT TTCCTATTAA AATAAATTAA TATAATATAC AATAATATTT CTTGTTATTT 300 TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTAA TTTCTTAATT 360 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAA AAAAAAAAA 420 AAAAAAAAA ATTTATTATA ATATAATAAA AAATATAAAG ACATACGTTC ACTTATTATT 480 45 ATAAATGATT TATTACGATT AAAACATATT GAGATTATAA TAATAATT TAACATAGAA 540 AGAGTTAAGA ATACATTTT TTTTTTTTT TGATATGTAA TTCAACATAT ATATATATAT 600 ATATCTTTTT AATTTAATTA AATAAAATTC CTTATTATTC ATATTGTTTC TTTTATCACA 660 TGTGAAATAT TAAAAATAAT TTTCGATTTT ATCGATATAT TTATGTCGTT TATATACTTA 720 TATAGGTCTT TATAACTATT GATTAATAGA AGGTAATAGC CTAATAATAT AAATACTCGT 780 50 ATTTATAAAT TCATTTATAT ATTTCAAATA TATTTCGATG GTTTATTTTC AAATACAATT 840 AATTAGATTT CTTAAATATT TCTTCATTTA TTCATTTTA TAGCATATAC ATGCACATTA 900 TTTCACACAA CATTTAAGTT GTCATAATGT AACACATTAA ATAATATTT ACTTATATAT 1020 ATATAATTAT TAATTATATA TTAAATAAAA ATGTATTATC GCCTGTATTA TCATAGTATA 1080 55 TATAATGTTG TATAACGCTT CAAAATATAT ATAATAATAT AATTAAAAAT ATATATATAG 1140 TAATTAATTA TTTTGTTATG TTATGTAATA ATGCAATTAA TATAAGATAA AATTCTATAG 1200 AATATTATAA TATGTAAATT ATTAATAAAA TATATTTGTA TAACATACAA GACTAAAGAA 1320 AACTATACAA TCTGGTATCT AATAGTATAT ATATATAATA TCTTTTTTAT TTAATTGTTC 1380 GATTTAGTAT TTTAATAATA AATAAATCTT TTAAAAAACT TCAAAACATT TTTGCATAAA 1500

ATAATATTAA TATTAGTAAC CACCTAGATA AATTAGAGAG AAACGTAGAA CATACCAAAA 1560 AAAATTAGAA CAAAAAGAAT ATTACAAAAA ATAATAAAAT TAAATTATTT CTTTACTATT 1620 AATTTAAAGT TTTTTTCAT ATCATATATT ATGATACACA ATGTTTGTTG TTAAATGTTT 1680

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	TATATACATG	CAATGATATG	TTTCTGTTGG	AATATGTATT	ATATACTTAT	ATGTTCTAAT	1740
	AAATGTATTG	TACACCTTTA	GCAACTATTA	CTACACACAT	TTTTATATAA	TTTATAACAG	1800
	GAAAATATGT	TATATTATTA	CAATATCTTA	ATGTGTTTTT	GCAAAAATAT	AAAAAACAAG	1860
_	AAAATTACAA	TTGTAATTAA	TCGTATGACA	TAAAATTATA	TTATATTAGA	AATTAAAATT	1920
5	CAAAATTATA	AAAAATATGG	AAATGTTTTG	TTATATTATT	TTTTTAAAAA	TTTAATTATT	1980
	TTATTTTATT	ATTTATTTT	TTTTTTTTT	GTGTTCTAAA	TAAAAAGGCA	AATATGATTC	2040
	AAGTAAAAA	TATATATATT	TACATAATGG	CAAAATAATT	GTTTATTATA	TTATATGACT	2100
	ATAATAATAT	TITAGATTAA	ACATATGTAA	TTCATTTAAC	AGAATAAAAT	AAAATATTAT	2160
**	ATATATATAT	TAATTATTAA	GTTATAGATT	TAATAAAAAT	ATATTATACA	TATGAGATTA	2220
10	AAAATGAAAG	TTCACTACAG	TAATATATTA	TTATATGTCG	TCAATTTAAG	TATATTCTTA	2280
	ATATCACGTA	TGCACTAAAT	AATGACAATA	ATAATATATA	TGTAACATTT	TATAATTGAT	2340
	GTAAATAAAA	AAATATACAT	ATATACAAAA	ACATATATGA	TATTTACATT	CTTTTTTATA	2400
	GATAAATATC	CAGAAGAACT	ATTACATCAC	TTCACTTCAT	ATACCAAACA	CGAAAAAAAT	2460
15	CAAACCACIA	GGTTATTATG	CGAATGTGAC	TTATATACGT	CCATTTATGA	TAATGACCCG	2520
15	GAMAIGAIAI	TAGIGATGGA	AAATTTCAAT	AAACAGACAG	AAGAAAGGTT	TCATGAATAC	2580
٠.	AMIGAACGCA	TOCARGARAA	ACGAAAAATA	TGTAAAGAAC	AATGCGAAAA	GGATATACAA	2640
	WWWWIIWIII	TANANGATAN	MATCGAAAAG	GAATTAACAG	AAAAGTTAGA	GGCATTGGAA	2700
	CTCCDDDDDD	CCTCTTTTC X X	ATTACCTACT	AMAMMOCOMO	AAAAATCAGT	AGCAGATAAA TCCATCTTTA	2760
20	CCTTTTATTAC	COLOTITORM	WIGIOGWOOI	ATATIGGGIG	TIGGIGIGAC	TCCTTTTTAT	2820
	ADACCTTTTC	ONOUNTAINGO	TCDAAAACCAA	WINNAINNII	CCCCTAAAAAA	TGCTAGTGAT	2880
	ACTGCTCGTA	TTGATACAGT	TATTTANCCA	ATANTATOLIG	CCGGIAMAAI	GCACACTATA	2940
	AATGGTTCTA	CGTTGGGGAA	ACTTATTACC	CTACAACCTC	TTANCCATCA	CACTACTCTT	3000
	ACTACGGCAC	TATATAATGA	ATATGTAAGC	ATGTGTGTAA	ATACCAACCC	TGTCGAAGAC	3120
25	AAATTAATTT	GTGCTTTTGG	GATGAGAGAC	GGTCTAGTTG	CAGGGCAATA	TGCTTCATCG	3120
	CGAGACGTTA	TAGGATCAAG	TGTAAAAGGA	ATTATTAGAA	AAGCTGCAAA	CGCTGCTTCA	3240
	CAAGCTGCTG	AGACAGCTGC	TAACGAAACT	ACTTCCGGAA	TGATCGAAGC	CGAGTTAAGT	3300
	AAAATAACAT	CTGCAGGTGC	TAATTTACAC	AGTGCAATTA	CTTACTCAGT	AACTGCGATA	3360
	TTGGTTATAG	TTTTGGTTAT	GGTAATTATT	TATTTAATAT	TACGTTATCG	TAGAAAAAA	3420
30	AAAATGAAGA	AAAAATTGCA	ATATATAAAA	TTATTAAAGG	AATAGATATA	CGATGTCGAG	3480
	CTATTAGCGG	TAATTTAAAG	TATTGTGAAT	TTTTCATTTA	ATATGCTATG	ATCATTTGAT	3540
	AATTAATTTT	TTTTTTATAAT	ATTATATTT	TTTATACCTT	GGATTCTTAC	ATTGTTTTAT	3600
	TATTATATGA	TTATTTAATT	ATTATACTTA	TATATATA	TATTTTTACA	TTAAGATATT	3660
05	ATATATGTAT	CTATCTATCT	ATCTATCTAT	ATATATATAT	ATATATATAT	ATTATAATAA	3720
35	TTATTATTAT	TAGATGCATA	TTAGTGATGA	TTATAATAAT	AACCTATTGA	AGAGAATAGA	3780
	ACATAATAAT	ATATTAAATT	AATAGAACTT	CATTTTTATT	GTTATATGTA	TATAAAAATA	3840
	AGAAATTTGA	AAAAGTAATT	TACACATGAT	AATGTATTTT	ATTTTATTTG	TGTTGTTTTA	3900
	TATTTATTTA	TAAAAATTGT	TTAATATAAG	TTGTTATTAT	AATTITITAA	TATGGCACCA	3960
40	TANGETTICE	ATTATACAAA	TATATATTTC	CTCATTAGAA	TCTGAATATT	TATTGTATTA	4020
70	AAATTTTAAA	WINNINIA	AAAAIAICIA	AGATTTTTTC	TAATTTGTTT	AATTTATAAT	4080
	AAGAACCTAT	TACAATACGAI	TANCAN ACTOC	WINNI CHACK	TATATATATG	CATTCATCTT	4140
	TGTAAAAGGA	TACTTCTTAA	VGGCL-Indulated	חייית ביייית מייית מייית	TIMIMMAIMM	CHIAAGAAIG	4200
	TATAATAGAT	ATCTTAACAT	ACAACTTTGC	ממים דיים מים מים מים מים מים מים מים מים מים	TARAGIGITI	GIMMOMIMIA	4200
45	AGAAATATTA	TAAATAATAT	TATAAAAAAT	TAAGCATAAA	TCTCACAAAA	VINITATE STATES	4320
	TATTAATTTA	ATTTTATTTT	ATTGTTCTAA	AATATATTGA	TTATGAGAAT	ልጥ ተልጥጥርጥር	4440
	TCTAATATAA	TTAAGATATT	TCTAATATTA	ATTTATATAT	ATATATTTAA	AAGTATTTTA	4500
	AGAATAATTT	TTTACTTATT	TATTATAATA	TGAAATATGC	ATGGAGTATA	TATAAATATT	4560
	GATGACAAAA	AAAAAACTTT	TAAAATGGAA	AATATGCATA	TAATAAAATA	CTATATAGTA	4620
50	TAATTGGTGA	AATAGTTGTA	ACTTATACAA	ACATGTTGCA	TTCATAATTT	AGAGATTATG	4680
	TAATATTGTT	TATGTATCGT	AATATATATT	AATATAATTG	TTTTTTTAGT	ATGTATGGTA	4740
	TTCTAATAAT	ATATTCATAT	GTAGTCATAG	TGTCAATGAA	TATAAAATAT	GGTATATTTA	4800
	TATTATTGTA	TAAATTAAAT	AAGTAACACA	GAACATTATA	TATAGTAATA	AATAGAAGAA	4860
	ATAATATATT	TTTATGTTAT	ATATTATTAG	TTATTATAAA	GGGGAAAATT	CATAATATTT	4920
5 5	ATGAAAATTT	TTGTATATGA	TATAGTTATA	AGTTAAAAAA	AAAAAAAAAC	AAGAACAAAA	4980
	ATGGAAAGCA	TAAAAAATGT	TACTGTAATA	GGATAAAATA	TATTATATAA	AATGTTTATT	5040
	TTATCTTAAA	AAGGTTCCTA	TTATAACATT	TTAAAAAAATT	TGTCCCATTT	TATAAATAAT	5100
	TAACTACATT	TACATAATGA	AATTTCGATT	TTGTGTTTTT	TTGATGAATA	TTATGGACTA	5160
60	ATTATTTATA	TGTGAATGCG	TTCTATATAA	TAATAATAAT	TITATITAAA	AAAA'I'GAAAA	5220
60	ATAAGAAATA	AATATCCTGA	TTTTGTAGTT	CCAATAGCTT	AATATAATTA	TGGACTCATA	5280
	TATATATTAT	ATATATCTTT	ACAACAAGTA	ATAAGTAAAT	ATTATTTAA	TCTTAATAAG	5340
	GAAAATAAAA	TAAAATAAAA	AAGAATACTG	AATAATAAGT	CATATTATAC	ATTTTTAAA	5400
	AATGTAACAT	MATTACAAAT	ACGTAACATG	TATTATAGAA	ATAATAAGAA	TTTAATATTA	5460
	AGGATAAATA	THAMIATTIA	AAATTATATT	TTTTATGTC	AATTTATGTT	ATATTATATT	5520

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	ATATTAACAT	GATTAGTTTT	TTGAAAAATA	TTTAAATATC	ATATAATAAT	AATAAATTAG	5580
	TTAAAATAAT	AGTATTTCAT	ACAAAATACT	AACTTATAAG	TATATCATAT	AATATTATAT	5640
	ATATATATAT	TTATGTGTTT	TTGATTGGGT	GTATATAAGG	CTATAAGTAT	ATATGGGTTG	5700
	TTCATTATAT	ATTTATATGT	GAATAGATAC	ATATAAGTTA	ATATATTTAT	TTGTGTATAT	5760
5	GTCTGTGTTA	AGATAGATAT	GCATTACAGT	TAAGGGTTAT	VC: Antalalalatata	TTTTTTTTT	5820
	GTACATATAT	ΑΤΑΑΑΑΑΤΑ	GATAACTAAC	AATATGCATA	TTACAACAAT	AATATTTGTA	5020
	ТАХААТАТАТ	ΑΤΑΤΑΤΑΤΑΤ	ΑΤΑΤΑΤΑΔΑΩ	ΔΟΔΥΤΆλλΑΟ	TATACTANTA	GGTAATTAGT	5040
	יימיימייייייייייייייייייייייייייייייייי	Cydeledalabababa	יים מיים איים מיים איים מיים איים איים א	the Designation of the Party 112 W. P. S. S. C.		TCGTTCTTTT	2340
	ע ע לאוי ע לאיני אלאיני	אסתאתאאראא	እጥአጥአ <i>አአአር</i> አ	አጥአጥሮአሮሞአ ጥ	TINCITCITO	AATAAATTTA	6000
10	ጥጥርምአርአ ጥ አጥ	VALUE TAYOUTH	MINIMANACA MINIMANACA	WINICHGIWI WINICHGIWI	TIGGWWIWIW	ATATATATAT	6060
10	TICIACAIAI	MIGCWINIMI	WIWIWIWIWI	WINIWINI	ATATATATAT	TATATATAT	6120
	VIVIGIUIOV	TITIMIMUM	TITITATACA	TGCATITIA	TATATTTTAG	TATATACTTT	6180
	AAAGAIAIIA	TIAATATTTA	TATAGTAGCA	TATATGTATT	TATATTATAA	CAAATATTTT	6240
	CATTIATATA	AATATATAGA	ACATGAACAT	TTTATTAATA	ACTCATATTT	GAATATATAT	6300
45	ATTTATAATG	TGTATTTTTA	CITATTITIT	TATATTATAC	AATAAAATTT	TGAAATTCAT	6360
15	AAAATGCATG	AAATACATAA	AAAAATACAA	CAAAACAAAT	GATAAAAACA	TTTTTTTAA	6420
	TATAATATAA	TATAATATAA	TAATATATT	TTCCTGTTAT	TTATTTATCA	${\tt TTTTTTTTT}$	6480
	GATGCTATAT	ATATTATTAT	ATAATAAATT	ATAATATATA	ACAACAAAAA	TTAATAATAA	6540
	TAATATACTA	CTTTTAATAT	AATACAACAA	TACAAAGAAT	ATGTATCTAT	ATCAATTATA	6600
	TATATATGAA	TATATAAATA	TGATAGATAA	TATAGATAGA	GAGAAACGAA	GAACATATTT	6660
20	GTCTCTTTTG	TTATCTCTAA	TATATATATA	TATATAATAA	ATTAAAATAA	AGTCAAAAA	6720
	AATATACATA	TATTAATGTT	AATAATTAAA	TATATAAACA	CGTTGCATAT	ATACTTTTTT	6780
	ATATGTTTGT	ATTTTCGTAT	TTTTTTTTC	TCATTTATAA	TTTTACTTAA	TAAATAAAAC	6840
	ATAAAAAAAA	TAATATATAT	ATAATTAAAT	AGATAAATAA	AGGAATACAT	AAAATATAAT	6900
	ATTTCTGATT	ATATTTTTT	TTTGTTAGAA	TATTTAAATT	TATTATAAAT	TTATTAATAT	6960
25	ATATATATAT	TTTTTTTAAA	AATATATAAA	ACTAATAATT	ATTATTATAT	ACATATTAAA	7020
	TATTATTTT	TTAACATATA	CATATATTGT	AATATTATAA	TAGTACAACT	ATTAATATAT	7080
	ATATATATAT	ATATACAATA	TTTATATATA	TTGTAATACA	TAAATTATAC	CTTACATATA	7140
	TATATACATT	CACAAAAGTG	TTATTATTCT	TATTCTACCA	TATTATAATA	CTACTGTAAT	7200
	ATACATATAT	ACATACCCCC	ACGTACGTAC	GAAACACCAC	CAAACCATGT	ATCACGTATG	7260
30	TATGTATGCC	ACGATATAAA	CCACGTACCA	CGTATGACAT	AATGTAATGG	TGGAGTTAGC	7320
	AAAAATGGGG	CCCAAGGAGG	CTGCAGGTGG	GGATGATATT	GAGGATGAAA	GTGCCAAACA	7380
	TATGTTTGAT	AGGATAGGAA	AAGATGTGTA	CGATAAAGTA	AAAGAGGAAG	CTAAAGAACG	7440
						AAAGCGATCC	
05	ACAAACACCA						
3 5	TGTAATTAAT						
	ATGTACACAT						
	ATATAGGCGA	TTGCATGTAT	GCGATCAAAA	TTTAGAACAG	ATAGAGCCTA	TAAAAATAAC	7740
	AAATACTCAT	AATTTATTGG	TAGATGTGTG	TATGGCAGCA	AAATTTGAAG	GACAATCAAT	7800
	AACACAAGAT						
40	TATGCTGGCA	CGAAGTTTTG	CGGACATAGG	GGACATTGTC	AGAGGAAGAG	ATTTGTATTT	7920
	AGGTAATCCA	CAAGAAATAA	AACAAAGACA	ACAATTAGAA	AATAATTTGA	AAACAATTTT	7980
	CGGGAAAATA	TATGAAAAAT	TGAATGGCGC	AGAAGCACGC	TACGGAAATG	ATCCGGAATT	8040
	TTTTAAATTA	CGAGAAGATT	GGTGGACTGC	TAATCGAGAA	ACAGTATGGA	AAGCCATCAC	8100
	ATGTAACGCT	TGGGGTAATA	CATATTTTCA	TGCAACGTGC	AATAGAGGAG	AACGAACTAA	8160
45	AGGTTACTGC	CGGTGTAACG	ACGACCAAGT	TCCCACATAT	TTTGATTATG	TGCCGCAGTA	8220
	TCTTCGCTGG	TTCGAGGAAT	GGGCAGAAGA	TTTTTGTAGG	AAAAAAAAA	AAAAAATAAA	8280
	AGATGTTAAA	AGAAATTGTC	GTGGAAAAGA	TAAAGAGGAT	AAGGATCGAT	ATTGTAGCCG	8340
	TAATGGCTAC	GATTGCGAAA	AAACTAAACG	AGCGATTGGT	AAGTTGCGTT	ATGGTAAGCA	8400
	ATGCATTAGC						
50	ACAATTTGAC	AAACAGAAAA	AAAAATATGA	TGAAGAAATA	AAAAAATATG	AAAATGGAGC	8520
	ATCAGGTGGT						
	ATATGAAAAA	AAATTTTATG	ACGAACTTAA	TAAAAGTGAA	TATAGAACCG	TTGATAAATT	8640
	TTTGGAAAAA						
	AATTGATTTT						
55	AAGTCAAGGA						
	AAAGGTAAAT	AATGGTGGTA	GTAGTAATGA	ATGGGAAGAG	AAAAATAATG	GCAAGTGCAA	8880
	GAGTGGAAAA						
	AAGTGGTAAA	GGACATGATG	ATATTGAAGA	AAAATTAAAC	AAATTTTGTG	ATGAAAAAA	9000
	TCCTCATACA	ATA A ATACTC	CTCCTACTCC	TACCCCTCCT	ACTOCTOCTO	CTAACACTGG	9060
60	TAGACAGGAA	TTGTATGAAG	AATGGAAATG	TTATAAAGGT	GAAGATGTAG	TGAAAGTTGG	9120
	ACACGATGAG						
	ATTAAAAAAC	CAAAAAAAGA	ATAAAGAAGA	AGGTGGAAAT	ACGTCTGAAA	AGGAGCCTGA	9240
	TGAAATCCAA						
	TTCCATACAT	TGGAAAAAA	AACTTCAGAG	ATGTTTACAA	AATGGTAACA	GAATAAAATG	9360

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(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3060 amino acids
 - (B) TYPE: amino acid

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- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEO ID NO:14:

Met Val Glu Leu Ala Lys Met Gly Pro Lys Glu Ala Ala Gly Gly Asp 10 50 Asp Ile Glu Asp Glu Ser Ala Lys His Met Phe Asp Arg Ile Gly Lys 25 Asp Val Tyr Asp Lys Val Lys Glu Glu Ala Lys Glu Arg Gly Lys Gly Leu Gln Gly Arg Leu Ser Glu Ala Lys Phe Glu Lys Asn Glu Ser Asp 55 55 Pro Gln Thr Pro Glu Asp Pro Cys Asp Leu Asp His Lys Tyr His Thr 70 Asn Val Thr Thr Asn Val Ile Asn Pro Cys Ala Asp Arg Ser Asp Val 90 60 Arg Phe Ser Asp Glu Tyr Gly Gly Gln Cys Thr His Asn Arg Ile Lys 105 Asp Ser Gln Gln Gly Asp Asn Lys Gly Ala Cys Ala Pro Tyr Arg Arg 120 125 Leu His Val Cys Asp Gln Asn Leu Glu Gln Ile Glu Pro Ile Lys Ile 130 135

	Thr 145	Asn	Thr	His	Asn	Leu 150	Leu	Val	Asp	Val			Ala	Ala	Lys	Phe
			Gln	Ser	Ile 165	Thr		Asp	Тут	Pro			Gln	Ala	Thr	160 Tyr
5				180	Ser	Gln			185	Met	Leu		_	190	Phe	Ala
			195					200					205	Gly	Asn	
10		210		Lys			215					220		_		
	225					230					235					Gly 240
15				Glu Val	245					250					255	
				260					265					270		
			275	Ala				280					285		_	-
20		290		Asp	•		295					300				
	305			Trp		310					315					320
	Asn	Lys	Lys	Ile	Lys 325	Asp	Val	Lys	Arg	Asn 330	Суз	Arg	Gly	Lys	Asp 335	Lys
25				Asp 340					345				_	350	Glu	Lys
•			355	Ala				360					365			
30		370		Ala -			375					380				-
	385			Asp		390					395					400
				Gly	405					410					415	
35				Thr 420					425					430	Tyr	_
			435	Lys				440				_	445			-
40 '		450		Glu			455					460			•	_
	465			Phe		470					475			_		480
				Val	485					490					495	-
45									505					510	-	
			515	Trp				520					525			
50		530		Pro			535					540				
	545			Lys		550					555				_	560
				Lys	565					570					575	Thr
55	Gly			580					585					590		
			595	Tyr				600					605		-	
6 0		610					615					62Ú				Ī
	625			Asn		630					635		-			640
	Glu	Lys	Glu		Asp 645	Glu	Ile	Gln	Lys	Thr 650	Phe	Asn	Pro	Phe	Phe 655	Tyr
65	Tyr	Trp	Val-			Met	Leu	Lys	Asp		Ile	His	Trp	Lys	Lys	Lys

										•						
	Leu	Gln	Arg	660 Cys		Gln	Asn	Gly	665 Asn	Arg	Ile	Lys	Cys	670 Gly	Asn	Asn
_	Lys	Cys	675 Asn	Asn	Asp	Cys	Glu	680 Cys		Lys	Arg	Trp	685 Ile		Gln	Lys
5	Lys	Asp					695					700				-
	705	Gly				710					715					720
10		Asp			725					730					735	-
				740					745					750	-	_
		Ser	755					760					765		_	
15		Glu 770					775		•			780				
	785					790					795					800
		Ile			805					810					815	
20	Cys	Leu	Glu	Ile 820	His	Glu	Asp	Glu	Glu 825	Glu	Glu	Lys	Glu	Lys	Gly	Asp
	Gly	Asn	Glu 835	Суѕ	Ile	Glu	Glu	Gly 840	Glu	Asn	Phe	Arg	Tyr 845	Aşn	Pro	Cys
25	Ser	Gly 850	Glu	Ser	Gly	Asn	Lys 855	Arg	Tyr	Pro	Val	Leu 860	Ala	Asn	Lys	Val
	Ala 865	Tyr	Gln	Met	His	His 870	Lys	Ala	Lys	Thr	Gln 875		Ala	Ser	Arg	Ala 880
	Gly	Arg	Ser	Ala	Leu 885	Arg	Gly	qaA	Ile	Ser 890	Leu	Ala	Gln	Phe	Lys 895	Asn
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	Asp 102	Gln 5	Glu	Ala	Met	Cys 1030	Arg	Ala	Val	Arg	Tyr 1035	Ser	Phe	Ala	Asp	Leu 1040
	Gly	Asp	Ile	Ile	Arg 1045	Gly		Asp	Met	Trp	Asp		Asp	Lys	Ser 105	Ser
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60	Mei	Tim	Glu				Trp	Tyr				Ğln	ser			
	Asp	Lys				Ile	Cys				Met	Ser				Gly
er.	Lys	Cys		_	Gly	Asp		. –		Gly	Lys		_		Ala	Cys
65		1170)				1175	5				1180)			

	Asp 118	Lys 5	Tyr	Lys	Glu	Glu 119	lle 0	Glu	Lys	Trp	Asn 119		Gln	Trp	Arg	Lys 1200
	Ile	Ser	Asp	Lys	Tyr 120	Asn 5	Leu	Leu	Tyr	Leu 121	Gln		Lys	Thr	Thr 121	Ser
5	Thr	Asn	Pro	Gly 122	Arg		Val	Leu	Gly 122	Asp	Asp	Asp	Pro	Asp	Tyr	Gln
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	Ala 126	Ala 5	Pro	Ile	Thr	Pro 127	Tyr 0	Ser	Thr	Ala	Ala 1275	Gly	Tyr	Ile	His	Gln 1280
			Gly		128	5				129	Thr	Gln		_	129	Lys 5
15			Gly	130	0				130	5				131	Ω	-
			Lys 131	5				132	0				132	5		
20		133					133	5				134	O			
	134	5	Cys			135	0				1355	;				1360
25			Val		1369	5				1370	0				137	5
25			Asn	138)				138	5				139	0	
			Gln 1399	5				140	0				140	5		
30		1410					141	5				142	0			
	142	•	Ala			1430	0				1435	,	•			1440
35			Glu		1445	•				1450)				145	5
55			Ser	1460)				1465	5				147	0	
			Asp 1475)				1486	0				148	5		
40		1490					1499	5				1500)		-	
	150	5	Arg			1510)				1515					1520
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			Ile	1540)				1545	5				1550)	
			Asn 1555	;				1560)				1565	5		
50		1570					1575	5				1580)			-
	1585	5	Asn			1590)				1595					1600
55		•	Asn		1605	,				1610)				1615	5
33			Tyr	1620)				1625	;				1630)	
			Val 1635	,				1640)				1645	5		
60		1650					1655	•				1666	;			_
	1665	•	Gln			1670)			,	1675					1680
CE			Ser		1685					1690	ì			_	1695	5
65	rys	Tyr	Pro	GIu	Lys	Cys	Asp	Cys	Tyr	Gln	Gly :	Lys	His	Val	Pro	Ser

				. 170					170					171	.0	
			171	5				172	0				172	5		Thr
5		173	0				173	5				174	0		_	Thr
	174	5				175	0				175	5				Pro 1760
10		Ser			176	5				177	0				177	5
10		Thr		178	0				178	5			_	179	0	
•		Arg	179	5				180	0				180	5		
15		Pro 181	0				181	5				182	0		_	-
	182					183	0				183	5				1840
20		Asp			184	5				185	0				185	5
		Ala Pro		186)				186	5				187	0	_
		Leu	187	5				188	0				188	5		
25		189 Gly	0				189	5				190	0			
	190	5 Ile				1910)				191	5				1920
30		Glu			192	5				193	0				193	5
•		Pro		1940)				194	5				195	n	
35		Glu	1959 Ser	5			Gly	196 Met	0				196	5		
33	Lys	1970 Asn		Asp	Thr	Ser	1979 Ala	5 Arg	Gly	Asp	Glu	198 Asn	0 Lys	Ile	Glu	Lys
	1989 Asp	Asp	Glu	Val	Tyr 2009	1990 Glu		Phe	Phe			Thr	Ala	qaA		_
40	Gly	Thr	Ala	Ser 2020	Thr		Thr	Gly	Thr 202			Thr	Gln			5 Tyr
	Glu	Lys	Val 2035	Lys		Glu	Asp	Thr 204	Ser	Gly	Ala	Lys	Thr 204		Ser	Ala
45		Ser 2050	Asp	Thr			205	Ser	Asp			206	Arg	Pro		_
	Phe 2065	Arg	Tyr	Leu	Glu	Glu 2070	Trp	Gly	Gln	Asn	Phe 2075	Cys	Lys	Lys	Arg	Lys 2080
		Lys			2085	5				2090	Lys)	Val			209	Gly 5
50		Gly		2100)				210	5	•			2110	Asp)	Gly
		Ala	2115	5				2120	0				2125	5		
55		Glu 2130)				2135	5				214	י כ		_	_
	2145					2150)				2155	•			_	2160
តំបំ		Gln			2165	5				2170)				217	5
60		Cys		2180	1				2185	5				2190)	_
		Leu	2195	;				2200)			_	2205	5		
65	ush	Asp 2210		nys	IIIF	rue	Lys 2215		TUX	тĀS	ASP	Cys 2220		PTO	cys	ren

	Lys 222	Phe 5	Ser	Val	Asn	Cys 223		Lys	Asp	Glu	Cys 223		Asn	Ser	Lys	Gly 2240
	Thr	Asp	Суѕ	Arg	Asn 224	Lys 5	Asn	Ser	Ile	Asp 225	Ala		Asp	Ile	Glu 225	Asn
5	Gly	Val	Asp	Ser 226	Thr		Leu	Glu	Met 226	Arg	Val	Ser	Ala	Asp 227	Ser	Lys
	Ser	Gly	Phe 227	Asn		Asp	Gly	Leu 228	Glu	Asn	Ala	Cys	Arg 228	Gly	Ala	Gly
10	Ile	Phe 229	Glu 0	Gly	Ile	Arg	Lys 229	Asp	Glu	Trp	Lys	Cys 230	Arg	Asn	Val	Cys
	230	5.				231	0				231	Gly	Glu		_	Gly 2320
					232	5				233	0				233	Tyr
15		Phe		234	0				234	5				235	Arg	Ile
		Asn	235	5				236	0				236	5		_
20		Asp 237	0				237	5				238	0			_
	238					239	0				2395	5				2400
25		Glu			240	5				241	0				241	5
25		Ile		242	0				242	5	•			243	0	
		Asn	243	5				2440	0				244	5		_
30		Leu 2450)				245	5				2460	0			
	246					247)			•	2475	;				2480
35		Glu			248	5				2490)				249	5
		Asn Glu		2500)				250	5				251	0	
		Ala	2519	5				2520)				252	5		
40		2530 Pro)				253	5				2540	0			
	2545	Lys				2550)				2555	•				2560
45		Gln			256	5.				2570)				257	5
		Leu		2580)				2585	5				259	0	
		Leu	2595	5				2600)				2605	5	_	
50		2610 Gln)				2615	5				2620)			
	2625	Arg				2630)				2635					2640
55		Tyr			2645	5				2650)				265	5
		Ser		2660)				2665	5				2670	o _	
		Asp	2675	5				2680)				2685	5		
60		2690 Val)				2695	•				2700)			
	2705	Thr				2710)				2715			•		2720
65	Ser	Asp	Thr	Gln	2725 Asn		Ile	Gln	Asn '	2730 Asp		Ile	Pro	Ser	273! Ser	5 Lys

				274	0				274	5				275	0	
	Ile	Thr	Asp 275		Glu	Trp	Asn	Gln 276		Lys	Asp	Glu	Phe 276	Ile	Ser	Gln
5	Tyr	Leu 277	Gln 0	Ser	Glu	Pro	Asn 277	Thr		Pro	Asn	Met 278	Leu		Tyr	Asn
	Val 278		Asn	Asn	Thr	His 279		Thr	Thr	Ser	His 279	His		Val	Glu	Glu 2800
	Lys	Pro	Phe	Ile	Met 280	Ser	Ile	His	Asp	Arg 281	Asn	Leu	Phe	Ser	Gly 281	Glu
10	Glu	Tyr	Asn	Tyr 282	Asp 0	Met	Phe	Asn	Ser 282		Asn	Asn	Pro	Ile 283	Asn	Ile
	Ser	Asp	Ser 283	Thr 5	Asn	Ser	Met	Asp 284	Ser	Leu	Thr	Ser	Asn 284	Asn	His	Ser
15 .	Pro	Tyr 285	Asn 0	Asp	Lys	Asn	Asp 285		Tyr	Ser	Gly	Ile 286	Asp		Ile	Asn
	286	5	Leu			2870)				287	5				2880
-			Glu		288	5				289)				289	5
20			Tyr	2900)				290	5				291	0	
			Asn 2915	5				292	3				292	5		
25		293					293	5				294	0	•		
	294	5	Asn			2950)				2955	5				2960
30			His		2965	5			·	2970)				297	5
00			Lys	2980)				2985	5				299	0	
			Thr 2995 Tyr	5				3000)				300	5		
35		3010	Glu				3015	5				3020)			
	302	5	Asn			3030)				3035	5				3040
40		1	Val		3045					3050		,,,,,			305	
				3060)											
	(2) INFO	RMAT	ON E	FOR S	SEQ 1	D NO):15	:								
45	(i) SE((A) 1		CE CH TH: 7		_											
	(B) (C)	STRAN	MDEDI	TESS:	sir					. .						
	(D) 7	ropoi	LOGY:	lir	near											
50									*							
	(ii) MOI										•					

- (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO
- 55 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

	TCCAAGCTGT	TTTTTTTCT	TTTTCTAGTT	TTTCCATTGT	ATATTCGTCA	AATACGTACA	60
	CATATATATA	TATATGTATA	ACATGTGAGT	ATTATTTAT	ACATCACATC	GATTACATTT	120
	TAGCGTTTTT	TTTCCCCAGA	TCACATATAG	TACGACTAAG	AAACAAAATA	ACATCATAAC	180
บิบิ	AAACATAGTG	ATTATCAATA	CATGATATTA	CCACATAATA	TAAAGTATTA	ATTATTA	240
	TTGCATGTTA	GTGATAACTA	CTATATCATA	TACACCACTA	CTAACTATCA	CTACATAGTA	300
	ACAGTAGTAG	TCACAATCAT	AGCATCATGG	TAATATAGAT	TTTCATTTCA	TATCTTCCTT	360
	ATTGTTTGTT	ATACATACAC	TATTAATATG	TATTTATGTT	ATAATGGTAG	ACTATGTTAA	420
	CAATGTATGA	ATGACCATCA	TAAATTAATA	ACAGACGCAT	CAAAACAGTG	TATATGTGTG	480
65	CATTTATGAC	ATAATGTAGT	CGGGAAGCAT	ACAAAAATGG	AGCCAGGAGG	TAGCGGTGGT	540

	CGTGGTAGTG	GCGGTAGTAG	TAGTGGTAAA	GGGAAGAAGG	ATACATCTGA	GTATATTTAT	600
	GTGAGCGATG	CTAAGGATCT	TTTGGATAGA	GTTGGAGAAA	AAGTGTACGA	AGAAAAAGTG	660
	AAAAATGGTG	ATGCTAAAAA	ATATATTGAG	GCGTTGAAAG	GAAATTTGAA	CACAGCAAAT	720
	GGTCGTAGTT	CGGAAACAGC	TAGCAGTATT	GAAACGTGCA	CCCTTGTAAA	AGAATATTAT	780
5	GAGCGTGTTA	ATGGTGATGG	TAAAAGGCAT	CCGTGCAGAA	AAGACGCAAA	AAATGAAGAT	840
	GTAAACCGTT	'TTTCGGATAC	ACTTGGTGGC	CAATGTACAT	ACAATAGGAT	AAAAGATAGT	900
	CAACAGGGTG	ATAATAAAGT	AGGAGCCTGT	GCTCCGTATA	GACGATTACA	TTTATGTGAT	960
	TATAATTTGG	AATCTATAGA	CACAACGTCG	ACGACGCATA	AGTTGTTGTT	AGAGGTGTGT	1020
	ATGGCAGCAA	AATACGAAGG	AAACTCAATA	AATACACATT	ATACACAACA	TCAACGAACT	1080
10	AATGAGGATT	CTGCTTCCCA	ATTATGTACT	GTATTAGCAC	GAAGTTTTGC	AGATATAGGT	1140
	GATATCGTAA	. GAGGAAAAGA	TCTATATCTC	GGTTATGATA	ATAAAGAAAA	AGAACAAAGA	1200
	AAAAAATTAG	AACAGAAATT	GAAAGATATT	TTCAAGAAAA	TACATAAGGA	CGTGATGAAG	1260
	ACGAATGGCG	CACAAGAACG	CTACATAGAT	GATGCCAAAG	GAGGAGATTT	TTTTCAATTA	1320
45	AGAGAAGATT	GGTGGACGTC	GAATCGAGAA	ACAGTATGGA	AAGCATTAAT	ATGTCATGCA	1380
15	CCAAAAGAAG	CTAATTATTT	TATAAAAACA	GCGTGTAATG	TAGGAAAAGG	AACTAATGGT	1440
	CAATGCCATT	GCATTGGTGG	AGATGTTCCC	ACATATTTCG	ATTATGTGCC	GCAGTATCTT	1500
	CGCTGGTTCG	AGGAATGGGC	AGAAGACTTT	TGCAGGAAAA	AAAAAAAAA	ACTAGAAAAT	1560
	TIGCAAAAAC	AGTGTCGTGA	TTACGAACAA	AATTTATATT	GTAGTGGTAA	TGGCTACGAT	1620
20	TGCACAAAAA	CTATATATAA	AAAAGGTAAA	CITGITATAG	GTGAACATTG	TACAAACTGT	1680
20	CARRAGA	GTCGTATGTA	TGAAACTTGG	ATAGATAACC	AGAAAAAAGA	ATTTCTAAAA	1740
	CAMAMAMGAA	AATACGAAAC	AGAAATATCA	GGTGGTGGTA	GTGGTAAGAG	TCCTAAAAGG	1800
	TATAAAACGGG	CTGCACGTAG	TAGTAGTAGT	AGTGATGATA	ATGGGTATGA	AAGTAAATTT	1860
	THIMMMAN	TGAAAGAAGT	A CA A COTTOR A	GATGTCGATA	AATTTTTAAA	AATATTAAAC	1920
25	בארט הארט הארני	TATGTCAAAA AAAAATATGT	ACAMCCICAA	TOTOTOTACA	AAAAAGCAGA	TAATGTTGAT	1980
	TGGTGTGGAT	TGGAAAAAGG	TCCTCCACCA	TCICGIACAG	AAATTTGTGA	ACCGIGCCCA	2040
	AGTGCAAAAA	CAAAGACATA	CCDTCCTDDD	A A TRATTA CCC	AAGGIGACAA	AACCTGCGGA	2100
	GATAAATCAC	AGCAAAATAT	ממממממדים	TATALACCG	THIRCCAGT	ACTUTACCUT	5100
	GGTGGTGGTC	AAATTAAAA	ATGGCAATGT	TATAMAMATI	AACATACCCC	TACTACTA A	2220
30	AATAATAATA	ATTGTGTAGA	AGGAACATGG	CACAACTTTA	CACAACCTAA	THGINGIAMA	2280
	AAGTCCTATA	ATGTTTTTT	TTGGGATTGG	GTTCATCATA	TCTTACACCA	TTCTCTACAC	2400
	TGGAAGACAG	AACTTAGTAA	GTGTATAAAT	AATAACACTA	ATGGCAACAC	ATCTACAAAC	2460
	AATAATAAAT	GTAAAACAGA	TTGTGGTTGT	TTTCAAAAAT	GGGTTGAAAA	AAAACAACAA	2520
	GAATGGATGG	CAATAAAAGA	CCATTTTGGA	AAGCAAACAG	ATATTGTCCA	ACAAAAAGGT	2580
3 5	CTTATCGTAT	TTAGTCCCTA	TGGAGTTCTT	GACCTTGTTT	TGAAGGGCGG	TAATCTGTTG	2640
	CAAAATATTA	AAGATGTTCA	TGGAGATACA	GATGACATAA	AACACATTAA	GAAACTGTTG	2700
	GATGAGGAAG	ACGCAGTAGC	AGTTGTTCTT	GGTGGCAAGG	ACAATACCAC	AATTGATAAA	2760
•	TTACTACAAC	ACGAAAAAGA	ACAAGCAGAA	CAATGCAAAC	AAAAGCAGGA	AGAATGCGAG	2820
40	AAAAAAGCAC	AACAAGAAAG	TCGTGGTCGC	TCCGCCGAAA	CCCGCGAAGA	CGAAAGGACA	2880
40	CAACAACCTG	CTGATAGTGC	CGGCGAAGTC	GAAGAAGAAG	AAGACGACGA	CGACTACGAC	2940
	GAAGACGACG	AAGATGACGA	CGTAGTCCAG	GAGGAGGAAG	AGGGAAAGGA	GGAAGGAACG	3000
	GTCACAGAGG	TAACAGAGGT	AACAGAGGTC	GTGGAAGAGA	CGGTAACAGA	ACAGGAAGGG	3060
	GTGAAGCCAT	GTGACATAGT	GGGCAAACTA	TTTGAGGACG	ACAAAAGTCT	CAAAGAGGCA	3120
45	TGTGGTCTAA	AATACGGTCC	AGGTGGAAAA	GAAAAATTCC	CCAATTGGAA	GTGTGTCACA	3180
40	CCAAGIGGIG	TCAGTACTGC	CACTAGTGGA	AAAGACGGCG	CTATATGTGT	GCCACCCAGG	3240
	ACCCACCTCT	TATACGTAGG CGAGTGAAGC	CACTTICATCA	CAATGGGCAA	GTCGTGGTGG	TGACGAGACC	3300
	ACCOMOGICI	TTGAGTCCGC	THE TEGGEG	ACCIDITATION OF THE PROPERTY O	CAGAAAGTGA	AAAACTACGT	3360
	CACAAAAAAC	CACCAGCAAC	ACAACATCCA	CCCCACTTC	CACTATICACT	CCCACA A CCC	3420
50	TCACCACCGG	GAGAGGACCC	CCAAACACAA	TTACAACAAA	CTCCTCTTAT	ACCCCCCCCAT	3480
	TATIONALCCAC	AAATGTTTTA	TACATTACCA	CACTACAAAA	ACATATTATA	CACTCCCCGAI	3540
	AACGACACAA	GTGACACAAC	TGGTAAACAG	ACACCTAGTA	GTAGTA ATGA	CAGIGGIAGI	3660
	AATATTGTTC	TGGAAGCAAG	TGGTAGTACT	GAGCAGGAGA	ACCACAAAAT	CAACCICAAA	300U
	CAAGCGAAAA	TAAAAAAAT	ТТТАААСССТ	GCCACATCTG	GTGTCCCACC	TCTCDCCDDD	3720 3700
5 5	AATAGTGTCA	AAACCCCCCA	ACAAACCTGG	TGGGAAAACA	TCGCGAAGGA	ТАТСТССААТ	3840
	GCTATGGTAT	GTGCACTAAC	ATATAAAGAA	AATGACGCCA	GAGGCACAAG	TCCCAAAATA	3040
	GAACAGAATA	AGGATTTGAA	AAAGGCACTT	TGGGACGAAG	CCAACAAAAA	CACCCCCATA	3960
	GAGAAATACC	AATACACAAA	TGTCAAACTC	GAAGATGAAA	GTGGTGCCAA	AAGCAACGAC	4020
	ACCATCCAAC	CCCCCACGTT	AAAAAATTTT	GTGGAAATAC	CTACATTTTT	TCGTTGGTTA	4080
<u> BŪ</u>	CATGAGTGGG	CNANCACTTT	ᠴᠮᢗᡆᢆᡆᡆᡊᠫ᠈ᢗ	AGAGCAAAAC	CATTCCCACA	*****	4140
	GAGTGTATGG	ATGAGGATGG	TGAAAAACAA	TATAGTGGGG	ATGGGGAATA	TTGTGAAGAA	4200
	ATTTTTAGTA	AGCAATATAA	TGTTCTCCAG	GATTTAAGTT	CCAGTTGCGC	TAAACCTTGT	4260
	AGATTGTATA	AAACGTGGAT	AGAAAAAAA	AAAACAGAAT	ATGAGAAACA	ACAAAAGGCA	4320
07	TATGAACAAC	AAAAAAGTAA	TTACGAAAAT	GAACAAAAAG	ACAAATGCCA	AACACAAAGT	4380
65	AATAATAATG	CTAATGAATT	TTCTAGAACA	CTAGGAGCGT	CCCCTACAGC	TGCAGAATTT	4440

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TTACAAAAGT TAGGATCATG TAAAAATGAT AATGGATATG AGAATGGAGA GGATAATAAA 4500
     ATAGATTTTA AAAATCCAGA TAAAACATTT AAGGAAGCAC ACAGTTGTGA TCCATGTCCT 4560
     ATAACTGGAG TTAAATGTCA AAATGGTCAT TGTGTGGGTT CTGCTAATGG AAAGGAGTGC 4620
     AAAAACAATA AGATTACTGC AGAAGATATT AAAAATAAGA CAGATCCTAA TGGAAACATA 4680
     GAAATGGTTG TCAGTGATGA CAGTACAAAT ACATTTGAAC ATTTAGGCGA TTGTAAAAGC 4740
     TCAGGTATCT TTAAAGGTAT CAGAAAAGAT GAATGGAAAT GCGCTAATGT ATGTGGTGTA 4800
     GATATATGTA CTCTGGAAAA AAAAATTAAG AATGGGCAAG AAGGTGATAA AAAATATATC 4860
     ACAATGAAAG AATTGCTTAA ACGATGGCTA GAATATTTTT TAGAAGATTA TAATAGAATT 4920
     AGAAAAAAA TAAAGCTATG TACGAAAAAG GAAGATGGAT GCAAATGTAT AAAAGGTTGT 4980
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     ATAGAAAAAT GGGTACAAGA AAAAACGAAA GAATGGCAAA AAATAAACGA TACTTATCTT 5040
     GAACAATATA AAAATGATGA TGGTAATACT TTAACTAATT TTTTGGAGCA ATTCCAATAT 5100
     CGAACTGAAT TTAAAAACGC TATAAAACCT TGTGATGGTT TAGACCAGTT CAAGACTTCG 5160
     TGTGGTCTTA ATAGTACTGA TAATTCACAA AATGGTAATA ATAACGATCT TGTTCTATGT 5220
     TTGCTTAATA AACTTCAAAA AAAAATTAGT GAGTGTAAAG AACAACATAG TGGCCAAACC 5280
     CAAACACCGT GTGATAACTC TTCCCTTAGT GGTAAAGAAT CCACCCTCGT TGAAGACGTT 5340
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     GATGATTATG AGGAACAAAA CCCAGAAAAC AAAGTGGAAC AACCTAAATT TTGTCCAGAT 5400
     ATGAAAGAAC CAAAAAAAGA AAACGATGAA GAAGTAGGCA CTTGTGGCGG AGACGAAGAA 5460
     AAAAAAAAG TGGAAGACAG TGTAATCGAA CAAAAAGAGG AAGAAGCAGC TAGTGCCCCA 5520
     GAGGAATCTC CTCCATTAAC CCCGGAAGCA CCAAAAAAAG AGGAAAATGT GGTACCAAAA 5580
     CCACCACCAC CACCAAAAAA ACGCCGAATC AAAACCCGTA ATGTGTTGGA CCACCCCGCT 5640
20
     GTCATACCCG CCCTCATGTC TTCTACCATC ATGTGGAGTA TTGGCATCGG TTTTGCTGCG 5700 TTCACTTATT TTTATCTAAA GAAAAAAACC AAATCATCTG TTGGAAATTT ATTCCAAATA 5760
     CTGCAAATAC CCAAAAGTGA TTATGATATA CCTACATTGA AATCAAGCAA TCGTTATATA 5820
     CCCTATGCAA GTGATAGACA TAAAGGCAAA ACATATATTT ATATGGAAGG AGATAGCAGT 5880
     GGAGATGAAA AATATGCATT TATGTCTGAT ACTACTGATA TAACTTCATC CGAAAGTGAG 5940
25
     TATGAAGAAT TGGATATTAA TGATATATAT GTACCAGGTA GTCCTAAATA TAAAACATTG 6000
     ATAGAAGTAG TACTTGAACC ATCAAAAAGA GATACACAAA ATGATATACA CAATGATATA 6060
     CCTAGTGATA TACCAAATAG TGACACCA CCACCCATTA CTGATGATGA ATGGAATCAA 6120
     TTGAAAAAG ATTTTATATC TAATATGTTA CAAAATACAC AAAATACGGA ACCAAATATT 6180
30
     TTACATGATA ATGTGGATAA TAATACCCAT CCTACCATGT CACGTCATAA TATGGACCAA 6240
     AAACCTTTTA TTATGTCCAT ACATGATAGA AATTTATTTA GTGGAGAAGA ATACAATTAT 6300
     GATATGTTTA ATAGTGGGAA TAATCCAATA AACATTAGTG ATTCAACAAA TAGTATGGAT 6360
     AGTCTAACAA GTAACAACCA TAGTCCATAT AATGATAAAA ATGATTTATA TAGTGGTATC 6420
     GACCTAATCA ACGACGCACT AAGTGGTAAT CATATTGATA TATATGATGA AATGCTCAAA 6480
     CGAAAAGAAA ATGAATTATT CGGGACGCAA CATCATCCAA AAAATATAAC GTCTAACCGT 6540
35
     GTCGTTACCC AAACAAGTAG TGACGACCCT ATAACCAATC AAATAAATTT GTTCCATAAA 6600
     TGGTTAGATA GGCATAGAGA TATGTGCGAA AAGTGGAAAA ATAATCACGA ACGGTTACCC 6660
     AAATTGAAAG AATTGTGGGA AAATGAGACA CATAGTGGTG ACATAAATAG TGGTATACCT 6720
     AGTGGTAACC ATGTGTTGAA TACTGATGTT TCTATTCAAA TAGATATGGA TAATCCGAAA 6780
40
    ACAATGAATG AATTTACTAA TATGGATACA AACCCCGACA AATCTACTAT GGATACTATA 6840
     TTGGATGATC TAGAAAAATA TAACGAACCC TACTACTATG ATTTTTATAA ACATGATATC 6900
     TATTATGATG TAAATGATGA TAAAGCATCT GAGGATCATA TAAATATGGA TCATAATAAG 6960
     ATGGATAATA ATAATTCGGA TGTCCCCACT AACGTACAAA TTGAAATGAA TGTCATTAAT 7020
     AATCAGGAGT TACTACAAAA TGAATATCCT ATATCGCATA TGTAGGGAAT ATGAAAATAA 7080
     TAGATGTATA TATGTTTTTT TCTTTTTTTG TGTGTGTGCA GTTTATATTT TTTATTTGTA 7140
45
    TATATTTTT TTTTTGTGCA TTTGTCTATT TTTTATTTGT GCTTTATATA TATATATATT 7260
     TTATTCAGCT TGGACTTAAC CAGGCTGAAC TTGCT
```

- 50 (2) INFORMATION FOR SEQ ID NO:16:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2182 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
- 60 (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (v) FRAGMENT TYPE: N-terminal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

	1				5					10					15	Ser
5				Lys 20					25					30	Asp	
			35	Leu				40					45		_	
10		50		Asp			55					60		_		
	65			Asn		70					75					80
15				Val	85					90				_	95	_
15				Cys 100					105			_		110	_	
			115	Leu				120					125	_	_	
20		130		Asp			135				•	140				
	145			Asp Leu		150					155					160
25				Thr	165					170					175	
				180 Leu					185					190	_	
			195	Arg				200					205	-		_
30	•	210		Arg			215					220	_		_	
	225			Lys		230					235					240
3 5				Ala	245					250					255	•
	Trp	Thr	Ser	260 Asn	Arg	Glu	Thr	Val	265 Trp	Lys	Ala	Leu	Ile	270 Cys	His	Ala
40	Pro	Lys	275 Glu	Ala	Asn	Tyr		280 Ile	Lys	Thr	Ala		285 Asn	Val	Gly	Lys
40	Gly 305	290 Thr	Asn	Gly	Gln		295 His	Cys	Ile	Gly		300 Asp	Val	Pro	Thr	- - .
		Asp	Tyr	Val	Pro 325	Gln	Tyr	Leu	Arg	Trp	315 Phe	Glu	Glu	Trp		320 Glu
45	Asp	Phe	Cys	Arg 340		Lys	Lys	Lys	Lys 345		Glu	Asn	Leu	Gln 350	335 Lys	Gln
	Cys	Arg	Asp 355	Tyr	Glu	Gln	Asn	Leu 360		Cys	Ser	Gly	Asn 365		Tyr	Asp
50		370		Thr			375					380	Ile			
	385			Cys		390					395					400
		•		Lys	405					410					415	
55				Gly 420					425					430		
			435	Ser -				440					445		_	
CC	,	45û		Leu			455					46Û				
_	465			Asn		470					475					480
65				Ala	485					490				_	495	_
UJ	Tnr	ьие	ser	Arg	ınr	GIU	тте	cys	GIU	Pro	cys	Pro	Trp	cys	Gly	Leu

Glu Lys Gly Gly Pro Pro Trp Lys Val Lys Gly Asp Lys Thr Cys Gly 515 520 525 525 525 525 525 525 525 525 525 526					500					505					510		
530 Val Leu Tyr Pro Asp Lys Ser Gln Gln Asn 11e Leu Lys Lys Tyr Lys 545 Asn Phe Cys Glu Lys Gly Ala Pro Gly Gly Gly Gln Ile Luy Lys Tyr Tyr Sys 555 Asn Phe Cys Glu Lys Gly Ala Pro Gly Gly Gly Gln Ile Lys Lys Tyr 575 580 Gln Cys Tyr Tyr Asp Glu His Arg Pro Ser Ser Lys Asn Asn Asn Asn Sys 595 Cys Val Glu Gly Thr Trp Asp Lys Phe Thr Gln Gly Lys Gln Thr Val 610 Lys Ser Tyr Asn Val Phe Phe Tpr Asp Trp Val His Asp Met Leu His 610 Asp Ser Val Glu Trp Lys Thr Glu Leu Ser Lys Cys Ile Asn Asn Asn 625 Thr Asn Gly Asn Thr Cys Arg Asn Asn Asn 1635 Cys Lys Thr Asp Cys 645 Thr Asn Gly Asn Thr Cys Arg Asn Asn Asn 1635 Cys Lys Thr Asp Cys 675 Agn Gly Cys Phe Gln Lys Trp Val Glu Lys Lys Gln Gln Gln Trp Met Ala Cys 1665 Leu Ile Val Phe Ser Pro Tyr Gly Val Leu Asp Leu Val Leu Lys Gly 675 Asp Gly Asn Leu Leu Gln Asn 11e Lys Asp Val His Gly Asp Thr Asp Asp 680 Gly Asn Leu Leu Gln Asn 11e Lys Asp Val His Gly Asp Thr Asp Asp 680 Gly Asn Leu Gln Ala Glu Chr Cys Lys Gln Lys Gln Gln Gln Cys Lys Thr Asp Asp 705 The Lys Lys Ala Gln Gln Gln Cys Lys Gln Lys Gln Glu Glu Cys Glu 735 Asp Glu Arg Thr Gln Gln Cys Lys Gln Lys Gln Lys Glu Glu Glu Cys Glu 685 Asp Ser Lys Ala Gln Gln Gln Glu Ser Arg Gly Arg Ser Ala Glu Thr Arg Glu 785 Asp Glu Arg Thr Gln Gln Fro Ala Asp Ser Ala Gly Glu Val Glu Glu Glu Glu Glu Glu Glu Glu Glu Gl		Glu	Lys			Pro	Pro	Trp			Lys	Gly	Asp		Thr	Cys	Gly
Sas	5		530		•			535					540				
10		545					550					555					560
September Sept	10					565					570					575	_
15	10				580					585					590		
Sap Ser Val Glu Trp Jys Thr Glu Leu Ser Lys Cys Tle Asn Asn Asn 625 630 630 635 635 640 640 645 645 65				595					600				_	605			
625	15		610					615					620				
20 Gly Cys Phe Gln Lys Trp Val Glu Lys Lys Gln Gln Gln Gln Trp Met Ala 650 11e Lys Asp His Phe Gly Lys Gln Thr Asp Ile Val Gln Gln Lys Gly 675 Leu Ile Val Phe Ser Pro Tyr Gly Val Leu Asp Leu Val Leu Lys Gly 700 Gly Asn Leu Leu Gln Asn Ile Lys Asp Val His Gly Asp Thr Asp Asp 705 11e Lys His Ile Lys Lys Leu Leu Asp Glu Glu Glu Asp Asp Asp 705 11e Lys His Ile Lys Lys Leu Leu Asp Glu Glu Asp Ala Val Ala Val 725 30 Val Leu Gly Gly Lys Asp Asn Thr Tr Ile Asp Lys Leu Leu Gln His 740 Glu Lys Glu Gln Ala Glu Gln Cys Lys Gln Lys Gln Glu Glu Cys Glu 755 Asp Glu Arg Thr Gln Gln Ser Arg Gly Arg Ser Ala Glu Thr Arg Glu 770 Asp Glu Asp Asp Asp Asp Asp Try Asp Glu Asp Asp Glu Glu Val Glu Glu Glu Glu Glu Cys Glu 785 Glu Glu Asp Asp Asp Asp Try Asp Glu Asp Asp Glu Glu Glu Val Glu Glu Glu 785 Glu Glu Asp Asp Asp Asp Try Asp Glu Asp Asp Asp Asp Val 800 Glu Glu Glu Asp Asp Asp Asp Try Asp Glu Glu Gly Thr Val Thr Glu Val 820 Thr Glu Val Thr Glu Val Glu Glu Glu Glu Try Val Glu Glu Glu Val 827 Thr Glu Val Tr Glu Val Glu Glu Glu Fry Lys Leu Phe Glu Asp Asp Asp Lys Ser 850 Val Lys Pro Cys Asp Ile Val Gly Lys Gry Fro Gry Gly Gly Lys Gly 845 45 46 47 48 48 49 40 41 41 42 45 45 46 47 48 49 40 40 40 41 41 42 45 45 46 46 47 48 48 49 40 40 40 41 41 41 42 43 44 45 46 46 47 48 48 49 40 40 40 40 40 40 40 40 40		625					630					635					640
The Lys Asp His Phe Gly Lys Gln Thr Asp The Val Gln Gln Lys Gly 680 680 685 680 680 685 700 705 70	20					645					650					655	_
Leu Tile Val Phe Ser Pro Tyr Gly Val Leu Asp Leu Val Leu Lys Gly Asp Col Gly Asp Leu Gln Asp Asp Tyr Asp Asp					660					665					670		
Gly Asn Leu Leu Gln Asn Ile Lys Asp Val His Gly Asp Thr Asp Asp 720 716 725 725 730 730 730 730 735	25	Leu			Phe	Ser	Pro			Val	Leu	Asp			Leu	Lys	Gly
Tile Lys His Ile Lys Lys Leu Leu Asp Glu Glu Asp Ala Val Ala Val 775	23			Leu	Leu	Gln			Lys	Asp	Val			Asp	Thr	Asp	
30			Lys	His	Ile			Leu	Leu	Asp			Asp	Ala	Val		
Total Process Total Proces	30	Val	Leu	Gly			Asp	Asn	Thr			Asp	Lys	Leu		Gln	His
Asp Glu Arg Thr Gln Gln Pro Ala Asp Ser Ala Gly Glu Val Glu Glu Ros Ros				755					760					765		_	
785	35		770					775					780				
40 Val Gln Glu	•	785					790					795	_				800
Thr Glu Val Thr Glu Val Val Gly Lys Leu Phe Glu Asp Asp Lys Ser 800	40					805					810					815	
Val Lys Pro Cys Asp II					820					825					830		
Leu Lys Glu Ala Cys Gly Leu Lys Tyr Gly Pro Gly Gly Lys Glu Lys 865				835					840					845			
865 Phe Pro Asn Trp Lys Cys Val Thr Pro Ser Gly Val Ser Thr Ala Thr 895 Ser Gly Lys Asp Gly Ala Ile Cys Val Pro Pro Arg Arg Arg Arg Leu 900 Tyr Val Gly Gly Leu Ser Gln Trp Ala Ser Arg Gly Gly Asp Glu Thr 915 Thr Glu Val Ser Ser Glu Ala Thr Ser Ala Pro Ser Gln Ser Glu Ser 935 Glu Lys Leu Arg Thr Ala Phe 1le Glu Ser Ala Ala Ile Glu Thr Phe 945 Phe Leu Trp His Lys Tyr Lys Glu Glu Lys Lys Pro Pro Ala Thr Gln 965 Asp Gly Ala Gly Leu Gly Val Ser Leu Pro Glu Pro Ser Pro Pro Gly 980 Glu Asp Pro Gln Thr Gln Leu Gln Gln Thr Gly Val Ile Pro Pro Asp 995 Phe Leu Arg Gln Met Phe Tyr Thr Leu Ala Asp Tyr Lys Asp Ile Leu	45		850					855					860	_	_		
50 Ser Gly Lys Asp Gly Ala Ile Cys Val Pro Pro Arg Arg Arg Arg Leu 900		865				Lys	870				Ser	875				Ala	880
Tyr Val Gly Gly Leu Ser Gln Trp Ala Ser Arg Gly Gly Asp Glu Thr 915	50	Ser	Gly	Lys			Ala	Ile	Cys			Pro	Arg	Arg			Leu
Thr Glu Val Ser Ser Glu Ala Thr Ser Ala Pro Ser Gln Ser Glu Ser 930		Tyr	Val			Leu	Ser	Gln			Ser	Arg	Gly			Glu	Thr
Glu Lys Leu Arg Thr Ala Phe Ile Glu Ser Ala Ala Ile Glu Thr Phe 945	55	Thr			Ser	Ser	Glu			Ser	Ala	Pro			Ser	Glu	Ser
965 970 975 Asp Gly Ala Gly Leu Gly val Ser Leu Pro Glu Pro Ser Pro Pro Gly 980 985 990 Glu Asp Pro Gln Thr Gln Leu Gln Gln Thr Gly Val Ile Pro Pro Asp 995 1000 1005 Phe Leu Arg Gln Met Phe Tyr Thr Leu Ala Asp Tyr Lys Asp Ile Leu				Leu	Arg	Thr			Ile	Glu	Ser			Ile	Glu	Thr	
980 985 990 Glu Asp Pro Gln Thr Gln Leu Gln Gln Thr Gly Val Ile Pro Pro Asp 995 1000 1005 Phe Leu Arg Gln Met Phe Tyr Thr Leu Ala Asp Tyr Lys Asp Ile Leu						965					970					975	
995 1000 1005 Phe Leu Arg Gln Met Phe Tyr Thr Leu Ala Asp Tyr Lys Asp Ile Leu	υŪ				980			•	•	985					990		-
				995					1000)				100	5		_
	65	rue			GIU	met	rne			тел	ATS	Asp			Asp	тте	ьeu

	Tyr Ser	Gly Se	r Asn	Asp		Ser	Asp	Thr	Thr 103		Lys	Gln	Thr	
		Ser As	n Asp	Asn		Lys	Asn	Ile 105	Val		Glu	Ala		
5	Ser Thi	Glu Gl			Glu	Lys	Met 106	Lys		Ile	Gln			Ile
,	Lys Lys			Gly	Ala	Thr	Ser		Val	Pro				Lys
10	Asn Ser	Val Ly	s Thr	Pro	Gln 109	Gln		Trp	Trp	Glu 110		Ile	Ala	Lys
	Asp Ile		n Ala	Met 111	Val		Ala	Leu		Tyr	Lys	Glu	Asn	_
	Ala Arg	Gly Th	r Ser	Ala		Île	Glu				Asp	Leu		
15	Ala Leu		p Glu		Asn	Lys				Ile	Glu			Gln
	Tyr Thr	Asn Va	.40 1 Lys	Leu	Glu				Gly	Ala			0 Asn	Asp
20	Thr Ile	1155 Gln Pr	o Pro	Thr				Phe	Val				Thr	Phe
20	Phe Arg		u His	Glu	117! Trp		Asn	Ser	Phe	118 Cys		Glu	Arg	Ala
	1185 Lys Arg	Leu Al	a Gln	119 Ile		His	Glu	Cys	1195 Met		Glu	Asp	Gly	1200 Glu
25	Lys Gln	Tvr Se	120 r Glv	-	Glv	Glu	Tvr	121 Cvs			Tle		121: Ser	
	Gln Tyr	12	20				122	5				123	0	_
	Arg Leu	1235				124	0	•			124	5		
30	125	0			125	5				126	0	_		_
	Gln Gln 1265	:		1270	0				1275	, -				1280
	Lys Asp		128	5				129)				129	5
35	Arg Thr	13	00				1309	5				131	0 _	
	Gly Ser	Cys Ly 1315	s Asn	Asp	Asn	Gly 132		Glu	Asn	Gly	Glu 132		Asn	Lys
40	Ile Asp 133		s Asn	Pro	Asp 1335		Thr	Phe	Lys	Glu 1340		His	Ser	Cys
	Asp Pro 1345			1350	o -		-	_	1355	:	-		-	1360
	Gly Ser		136	5				1370)	_			137	5
45	Asp Ile	13	80				1385	5				1390)	
	Ser Asp	Asp Se 1395	r Thr	Asn	Thr	Phe 1400		His	Leu	Gly	Asp 140	_	Lys	Ser
50	Ser Gly		e Lys	Gly	Ile 1415		Lys	Asp	Glu	Trp 1420		Суѕ	Ala	Asn
	Val Cys 1425	Gly Va	l Asp	Ile 1430		Thr	Leu	Glu	Lys 1435		Ile	Lys	Asn	Gly 1440
	Gln Glu	Gly As	p Lys 144		Tyr	Ile	Thr	Met 1450	Lys		Leu	Leu	Lys 1459	Arg
55	Trp Leu	Glu Ty 14		Leu	Glu	Asp	Tyr 1465	Asn		Ile	Arg	Lys 1470	Lys	
	Lys Leu			Lys	Glu	Asp 1480	Gly		Lys	Cys	Ile 1489	Lys		Cys
00	Ile Glu 149		p Val	Gln	Glu 1495	Lys		Lys	Glu	Trp 1500	Gln		Ile	Asn
	Asp Thr 1505		u Glu	Gln 1510	Tyr		Asn	Asp	Asp 1515	Gly		Thr	Leu	Thr 1520
	Asn Phe	Leu Gl	u Gln 152	Phe		Tyr	Arg	Thr 1530	Glu		Lys	Asn	Ala 1539	Ile
65	Lys Pro	Cys As			Asp	Gln	Phe			Ser	Cys	Gly		

	_			154					154					155	0	
			155	5				156	0				156	5		Cys
5		157	0	Lys			157	5				158	0			
	158	5		Thr		159	0				159	5			_	1600
				Leu	160	5				161	0				161	Pro 5
10				Val 162	0				162	5				163	0	
			163					164	0				164	5		
15		165	0	Val			165	5				166	0			
	166	5		Pro		167	0				167	5				1680
20				Asn	168	5				169	0			_	169	5
				Thr 1700	0				170	5				171	n	
			171					172	0				172	5		
25		1730)	Phe Ile			173	5				174	0			
•	174	•				1750)				1755	5				1760
30				Ser	1769	5				1770	0				177	5
				Tyr 1780)				178	5				1790	0	
			1799	Met 5 Leu				1800	0				180	5		
35		1810)				181	5				1820)			
	1825	•		Leu		1830)				1835	5				1840
\$ 0				Ile	1845	5				1850)				185	5
+0				Pro 1860)				1869	5				1870	ס ֿ	_
			1875	Asn Asn				1880)				1889	5		
15	•	1890)	Gln			1899	5				1900)		_	
	1905	5		Glu		1910)				1915	;				1920
50				Ile	1925	•				1930)			_	1939	5
٠				1940 Ser)				1945	5	•			1950)	
	•		1955			•		1960)				1965	5	_	
55		1970	1	Lys			1975	5				1980) .		_	_
	1985	5		Ile		1990)				1995	-				2000
30				Tìm	2005					2010)				2019	5 ~
			Asp	2020 M et)			Trp	2025 Lys	5			_	2030)	_
: E				Glu	Leu					Thr					Ile	Asn
i5		2050	1				2055	•				2060)			

	Ser Gly Ile Pro Ser Gly Asn His Val Leu Asn Thr Asp Val Ser Ile 2065 2070 2075 2080
	Gln Ile Asp Met Asp Asn Pro Lys Thr Met Asn Glu Phe Thr Asn Met 2085 2090 2095
5	Asp Thr Asn Pro Asp Lys Ser Thr Met Asp Thr Ile Leu Asp Asp Leu 2100 2105 2110
	Glu Lys Tyr Asn Glu Pro Tyr Tyr Tyr Asp Phe Tyr Lys His Asp Ile 2115 2120 2125
10	Tyr Tyr Asp Val Asn Asp Asp Lys Ala Ser Glu Asp His Ile Asn Met 2130 2135 2140
	Asp His Asn Lys Met Asp Asn Asn Ser Asp Val Pro Thr Asn Val 2145 2150 2155 2160
	Gln Ile Glu Met Asn Val Ile Asn Asn Gln Glu Leu Leu Gln Asn Glu 2165 2170 2175
15	Tyr Pro Ile Ser His Met 2180
	(2) INFORMATION FOR SEQ ID NO:17:
20	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs
	(B) TYPE: nucleic acid (C) STRANDEDNESS: single
25	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO
	(iv) ANTISENSE: NO (v) FRAGMENT TYPE:
30	(vi) ORIGINAL SOURCE:
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
35	ATCGATCAGC TGGGAAGAAA TACTTCATCT 30
VO	(2) INFORMATION FOR SEQ ID NO:18:
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs
40	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: single (D) TOPOLOGY: linear
45	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO
	(iv) ANTISENSE: NO
	(v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
	ATCGATGGGC CCCGAAGTTT GTTCATTATT 30
55	(2) INFORMATION FOR SEQ ID NO:19:
33	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 30 base pairs (B) TYPE: nucleic acid
	(C) STRANDEDNESS: single
ōิบิ	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO
	(iv) ANTISENSE: NO
65	(v) FRAGMENT TYPE:

	(vi) ORIGINAL SOURCE:	
_	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
5	TCTCGTCAGC TGACGATCTC TAGTGCTATT	30
	(2) INFORMATION FOR SEQ ID NO:20:	
10	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 30 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
15	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:</pre>	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
	ACGAGTGGGC CCTGTCACAA CTTCCTGAGT	30
25	(2) INFORMATION FOR SEQ ID NO:21:	
30	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 17 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
35	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:	
40	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
	AGACCTCAAT TTCTAAG (2) INFORMATION FOR SEQ ID NO:22:	17
45	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single	•
50	(D) TOPOLOGY: linear	
-	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE:	
55	(vi) ORIGINAL SOURCE:(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
	AATCGCGAGC ATCATCTG	18
60		10
	(2) INFORMATION FOR SEQ ID NO:23:	
65	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 20 base pairs(B) TYPE: nucleic acid	
	·-·	

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(C) STRANDEDNESS: single
             (D) TOPOLOGY: linear
           (ii) MOLECULE TYPE: cDNA
 5
           (iii) HYPOTHETICAL: NO
           (iv) ANTISENSE: NO
           (v) FRAGMENT TYPE:
           (vi) ORIGINAL SOURCE:
10
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:
     CCRAGRAGRC AARAAYTATG
                                                               20
             (2) INFORMATION FOR SEO ID NO:24:
15
           (i) SEQUENCE CHARACTERISTICS:
             (A) LENGTH: 18 base pairs
             (B) TYPE: nucleic acid
             (C) STRANDEDNESS: single
(D) TOPOLOGY: linear
20
           (ii) MOLECULE TYPE: cDNA
           (iii) HYPOTHETICAL: NO
           (iv) ANTISENSE: NO
25
           (v) FRAGMENT TYPE:
           (vi) ORIGINAL SOURCE:
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:
30
     CCAWCKKARR AATTGWGG
                                                               18
             (2) INFORMATION FOR SEQ ID NO:25:
          (i) SEQUENCE CHARACTERISTICS:
35
            (A) LENGTH: 291 amino acids
            (B) TYPE: amino acid
            (C) STRANDEDNESS: single
            (D) TOPOLOGY: linear
40
           (ii) MOLECULE TYPE: peptide
          (iii) HYPOTHETICAL: NO
          (iv) ANTISENSE: NO
          (v) FRAGMENT TYPE: internal
          (vi) ORIGINAL SOURCE:
45
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:
     10
                                                      15
50
     Xaa Xaa Xaa Val Cys Ile Pro Asp Arg Arg Tyr Gln Leu Cys Met Lys
               20
                                 25
                                                  30
     35
                             40
     55
                          55
                                           60
     70
                                        75
     Xaa Asp Phe Cys Lys Asp Ile Arg Trp Ser Leu Gly Asp Phe Gly Asp
                   85
                                    90
CC
     The The Met Gly Thr Asp Met Glu Gly The Gly Tyr Ser Lys Xaa Xaa
               100
                                 105
                                                  110
     Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Thr Asp Glu Lys Ala Gln Gln
                             120
                                               125
     Arg Arg Lys Gln Trp Trp Asn Glu Ser Lys Ala Gln Ile Trp Thr Ala
65
                          135
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	Met 145	Met	Tyr	Ser	Val	Xaa 150	Xaa	Xaa	Xaa	Xaa	Xaa 155	Xaa	Xaa	Xaa	Xaa	Xaa 160
	Cys	Xaa	Xaa	Xaa	Xaa 165	Xaa	Xaa	Xaa	Xaa	Glu 170	Pro	Gln	Ile	Tyr	Arg 175	Trp
5	Ile	Arg	Glu	Trp 180	Gly	Arg	Asp	Tyr	Val 185	Ser	Glu	Leu	Pro	Thr 190	Glu	Val
	Gln	Lys	Leu 195	Lys	Glu	Lys	Cys	Xaa 200	Xaa	Xaa	Xaa	Xaa	Xaa 205	Xaa	Xaa	Xaa
10	Xaa	Xaa 210	Cys	Xaa	Val	Pro	Pro 215	Cys	Gln	Asn	Ala	Cys 220	Lys	Ser	Tyr	Asp
	Gln 225	Trp	Ile	Thr	Arg	Lys 230	Lys	Asn	Xaa	Xaa	Xaa 235	Xaa	Xaa	Xaa	Xaa	Xaa 240
	Xaa	Xaa	Xaa	Xaa	Xaa 245	Xaa	Xaa	Xaa	Xaa	Xaa 250	Xaa	Xaa	Xaa	Xaa	Xaa 255	
15	Xaa	Xaa	Xaa	Xaa 260	Xaa	Xaa	Xaa	Xaa	Xaa 265	Xaa	Xaa	Xaa	Xaa	Xaa 270		Xaa
	Xaa	Xaa	Xaa 275	Xaa	Xaa	Xaa	Xaa	Xaa 280	Xaa	Xaa	Xaa	Xaa	Xaa 285	Xaa	Xaa	Xaa
20	Cys	Xaa 290	Cys									•				

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS: 25
 - (A) LENGTH: 271 amino acids
 (B) TYPE: amino acid

 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (iii) MOLECULE TYPE: peptide (iii) HYPOTHETICAL: NO 30

 - (iv) ANTISENSE: NO
 - (v) FRAGMENT TYPE: internal
 - (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

	Cys 1	Xaa	Xaa	Xaa	Xaa 5	Xaa	Xaa	Xaa	Хаа	Xaa 10	Xaa	Cys	Xaa	Xaa	Xaa 15	Xaa
40				20	Xaa				25					30		-
			35		Xaa			40					45			
45	Xaa	Xaa 50	Xaa	Xaa	Xaa	Xaa	Xaa 55	Xaa	Xaa	Xaa	Xaa	Xaa 60	Xaa	Xaa	Xaa	Xaa
	65				Xaa	70					75		_		-	80
					Tyr 85					90			_		95	
50				100	Thr				105			•		110		
	Xaa	Xaa	Xaa 115	Xaa	Xaa	Xaa	Ser	Glu 120	His	Lys	Ile	Lys	Asn 125	Phe	Arg	Lys
55	Glu	Trp 130	Trp	Asn	Glu	Phe	Arg 135	Glu	Lys	Leu	Trp	Glu 140	Ala	Met	Leu	Ser
	Glu 145	His	Xaa	Xaa	Xaa	Xaa 150	Xaa	Xaa	Cys	Xaa	Xaa 155	Xaa	Xaa	Xaa	Xaa	Glu 160
	Leu	Gln	Ile	Thr	Gln 165	Trp	Iḷe	Lys	Glu	Trp 170	His	Gly	Glu	Phe	Leu 175	Leu
ĐŪ	GIu	Arg	Asp	Asn 180	Arg	ser	Lys	Leu	Pro 185	гàз	Ser	Lys	Cys	Xaa 190		Xaa
	Xaa	Xaa	Xaa 195	Xaa	Xaa	Cys	Xaa	Glu 200	Lys	Glu	Cys	Ile	Asp 205	Pro	Cys	Met
65	Lys	Tyr 210	Arg	Asp	Trp	Ile	Ile 215	Arg	Ser	Lys	Phe	Xaa 220		Xaa	Xaa	Xaa

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225
       230
              235
245
            250
                    255
Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Cys
    260
           265
```

(2) INFORMATION FOR SEQ ID NO:27:

```
10
            (i) SEQUENCE CHARACTERISTICS:
              (A) LENGTH: 277 amino acids
```

- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

15

5

- (ii) MOLECULE TYPE: peptide (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- 20 (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

25	Cys 1	Xaa	Xaa	Xaa	Xaa 5	Xaa	Xaa	Xaa	Xaa	Xaa 10	Xaa	Xaa	Xaa	Xaa	Cys 15	Xaa
	•			20	Xaa				25	Val				30	Arg	_
	Gln	Glu	Leu 35	Cys	Leu	Gly	Asn	Ile 40	Xaa	Xaa	Xaa	Xaa	Xaa 45	Xaa	Xaa	Xaa
30		50			Xaa		55					60				
	65				Xaa	70					75				_	80
3 5					Thr 85					90				. –	95	
	Asp	Tyr	Trp	Asn 100	Asp	Leu	Ser	Asn	Arg 105	Xaa	Xaa	Xaa	Xaa	Xaa 110	Xaa	Xaa
	Xaa	Xaa	Xaa 115	Xaa	Xaa	Xaa	Xaa	Xaa 120	Asn	Lys	Lys	Asn	Asp 125	Lys	Leu	Phe
40		130			Trp		135					140	_			
	145				Xaa	150					155					160
45					Phe 165					170		-		_	175	Ī
	Gln	Asp	Lys	Thr 180	Lys	Met	Ile	Glu	Thr 185	Leu	Lys	Val	Glu	Cys 190	Xaa	Xaa
	Xaa	Xaa	Cys 195	Xaa	Asp	Asp	Asn	Cys 200	Lys	Ser	Lys	Cys	Asn 205	Ser	Tyr	Lys
50	Glu	Trp 210	Ile	Ser	Lys	Lys	Lys 215	Lys	Xaa	Xaa	Xaa	Xaa 220	Xaa	Xaa	Xaa	Xaa
	Xaa 225	Xaa	Xaa	Xaa	Xaa	Xaa 230	Xaa	Xaa	Xaa	Xaa	Xaa 235	Xaa	Xaa	Xaa	Xaa	Xaa 240
55	Xaa	Xaa	Xaa	Xaa	Xaa 245	Xaa	Xaa	Xaa	Xaa	Xaa 250	Xaa	Xaa	Суѕ	Xaa	Xaa 255	Xaa
	Xaa	Xaa	Xaa	Xaa 260	Xaa	Xaa	Xaa	Xaa	Xaa 265	Xaa	Xaa	Xaa	Xaa	Xaa 270		Xaa
	Xaa	Cys	Xaa 275		Cys									3,70		
60																

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 282 amino acids
- (B) TYPE: amino acid 65

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(C) STRANDEDNESS: single
      (D) TOPOLOGY: linear
     (ii) MOLECULE TYPE: peptide
     (iii) HYPOTHETICAL: NO
     (iv) ANTISENSE: NO
     (v) FRAGMENT TYPE: internal
     (vi) ORIGINAL SOURCE:
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Val Cys Gly Pro Pro Arg Arg
Gln Gln Leu Cys Leu Gly Tyr Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                     40
70
                              75
Ala Ile Leu Gly Ser Tyr Ala Asp Ile Gly Asp Ile Val Arg Gly Leu
                           90
Asp Val Trp Arg Asp Ile Asn Thr Asn Xaa Xaa Xaa Xaa Xaa Xaa
         100
                        105
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Lys Lys Gln Asn Asp Asn
                     120
                                    125
Asn Glu Arg Asn Lys Trp Trp Glu Lys Gln Arg Asn Leu Ile Trp Ser
   130
                  135
Ser Met Val Lys His Ile Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa
               150
                              155
Xaa Xaa Xaa Ile Pro Gln Phe Leu Arg Trp Leu Lys Glu Trp Gly
            165
                           170
Asp Glu Phe Cys Glu Glu Met Gly Thr Glu Val Lys Gln Leu Glu Lys
         180
                        185
Ile Cys Xaa Xaa Xaa Cys Xaa Glu Lys Lys Cys Lys Asn Ala Cys
                     200
                                    205
Ser Ser Tyr Glu Lys Trp Ile Lys Glu Arg Lys Asn Xaa Xaa Xaa Xaa
                  215
                                 220
230
                              235
245
                           250
260
                        265
Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
    275
                     280
       (2) INFORMATION FOR SEQ ID NO:29:
    (i) SEQUENCE CHARACTERISTICS:
      (A) LENGTH: 324 amino acids
      (B) TYPE: amino acid
      (C) STRANDEDNESS: single
      (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: peptide
```

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE: internal (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

65 Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa

	1				5					10					15	·
	Xaa	Xaa	Xaa	Xaa 20	Xaa	Xaa	Xaa	Ala	Cys 25	Ile	Pro	Pro	Arg	Arg 30	Gln	Lys
5		Cys	35					40				•	45			
	Xaa	Xaa 50	Xaa	Xaa	Xaa	Xaa	Xaa 55	Xaa	Xaa	Xaa	Xaa	Xaa 60	Xaa	Xaa	Xaa	Xaa
	65	Xaa				70					7 5					80
10		Xaa			85			•		90			_		95	
		Thr		100					105					110		
15		Lys	115		•			120					125			
		Xaa 130					135					140	_	_	_	
00	145	Trp				150					155	_			-	160
20		Xaa			165					170					175	
		Xaa 		180					185					190		
25		Xaa -	195					200				_	205			
		Trp 210	•				215					220			-	-
20	225	Tyr				230					235					240
30		Xaa			245					250		_	_		255	-
		Gln		260					265	_		_		270		
35		Xaa	275					280					285			
		Xaa 290					295					300				
40	305	Xaa			Xaa	Xaa 310	Cys	Xaa	Xaa	Xaa	Xaa 315	Xaa	Xaa	Xaa	Xaa	Cys 320
40	Xaa	Xaa	Xaa	Cys												

(2) INFORMATION FOR SEQ ID NO:30:

45 (i) SEQUENCE CHARACTERISTICS:

50

55

(A) LENGTH: 362 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE: internal

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

-81-

```
Ala Arg Ser Phe Ala Asp Ile Gly Asp Ile Val Arg Gly Lys Asp Leu
             70
                       75
   Tyr Leu Gly Tyr Asp Asn Lys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
5
           85
                     90
   100
                   105
   120
                            125
   Phe Phe Gln Leu Arg Glu Asp Trp Trp Thr Ser Asn Arg Glu Thr Val
10
               135
   Trp Lys Ala Leu Ile Cys His Ala Xaa Xaa Xaa Xaa Xaa Xaa Xaa
             150
                       155
   15
           165
                     170
   180
                   185
   Arg Trp Phe Glu Glu Trp Ala Glu Asp Phe Cys Arg Lys Lys Lys
       195
                 200
                           205
20
   Lys Leu Glu Asn Leu Gln Lys Gln Cys Xaa Xaa Xaa Xaa Xaa Xaa Cys
               215
                         220
   230
                       235
   Thr Asn Cys Ser Val Trp Cys Arg Met Tyr Glu Thr Trp Ile Asp Asn
25
           245
                     250
   260
                   265
   275
                280
                            285
30
   295
                         300
   310
                       315
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
35
           325
                     330
   345
   Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
40
```

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 411 amino acids

(B) TYPE: amino acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

ûῦ Ala Cys Ala Pro Tyr Arg Arg Leu His Val Cys Asp Gln Asn Leu Xaa

```
85
                        90
    Met Leu Ala Arg Ser Phe Ala Asp Ile Gly Asp Ile Val Arg Gly Arg
 5
           100
                       105
                                  110
    Asp Leu Tyr Leu Gly Asn Pro Gln Glu Xaa Xaa Xaa Xaa Xaa Xaa
         115
                     120
                                125
    140
10
    Xaa Xaa Xaa Xaa Xaa Xaa Asn Asp Pro Glu Phe Phe Lys Leu Arg
                150
                           155
    Glu Asp Trp Trp Thr Ala Asn Arg Glu Thr Val Trp Lys Ala Ile Thr
             165
                         170
                                     175
    15
                      185
                                  190
    195
                    200
    Xaa Xaa Xaa Xaa Val Pro Gln Tyr Leu Arg Trp Phe Glu Glu Trp Ala
                              220
    Glu Asp Phe Cys Arg Lys Lys Asn Lys Lys Ile Lys Asp Val Lys Arg
20
                230
                           235
    245
                         250
    25
           260
                       265
                                  270
    Xaa Xaa Xaa Xaa Cys Ile Ser Cys Leu Tyr Ala Cys Asn Pro Tyr
        275
                    280
                                285
    Val Asp Trp Ile Asn Asn Gln Lys Glu Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                  295
                             300
30
    310
                           315
    325
                         330
    35
           340
                      345
    355
                    360
                                365
    375
                            380
40
    390
                           395
    Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
             405
45
         (2) INFORMATION FOR SEQ ID NO:32:
       (i) SEQUENCE CHARACTERISTICS:
         (A) LENGTH: 411 amino acids
         (B) TYPE: amino acid
50
         (C) STRANDEDNESS: single
         (D) TOPOLOGY: linear
       (ii) MOLECULE TYPE: peptide
       (iii) HYPOTHETICAL: NO
55
       (iv) ANTISENSE: NO
       (v) FRAGMENT TYPE: internal
       (vi) ORIGINAL SOURCE:
       (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:
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40
    Xaa Xaa Val Phe Leu Pro Pro Arg Arg Glu His Met Cys Thr Ser Asn
                   55
    5
                70
                             75
    85
                          90
    105
10
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Ala Met Cys Arg Ala Val Arg Tyr
                     120
    Ser Phe Ala Asp Leu Gly Asp Ile Ile Arg Gly Arg Asp Met Trp Asp
                   135
                               140
    15
                150
                             155
    165
                          170
    Xaa Xaa Xaa Xaa Lys Lys Pro Ala Tyr Lys Lys Leu Arg Ala Asp
                       185
20
    Trp Trp Glu Ala Asn Arg His Gln Val Trp Arg Ala Met Lys Cys Ala
         195
                     200
    Thr Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Ile Pro
      210
                   215
                               220
    Gln Arg Leu Arg Trp Met Thr Glu Trp Ala Glu Trp Tyr Cys Lys Ala
25
                230
                             235
   Gln Ser Gln Glu Tyr Asp Lys Leu Lys Lys Ile Cys Xaa Xaa Xaa Xaa
              245
                          250
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Cys Gly
           260
                        265
                                    270
30
   Lys Cys Lys Ala Ala Cys Asp Lys Tyr Lys Glu Glu Ile Glu Lys Trp
        275
                     280
                                  285
   Asn Glu Gln Trp Arg Lys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                  295
                               300
   35
                310
                            315
   325
                          330
   340
                       345
40
   355
                     360
                                  365
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                  375
                               380
   45
                390
                            395
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
              405
         (2) INFORMATION FOR SEQ ID NO:33:
50
       (i) SEQUENCE CHARACTERISTICS:
         (A) LENGTH: 311 amino acids
         (B) TYPE: amino acid
```

- (C) STRANDEDNESS: single
- 55 (D) TOPOLOGY: linear

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- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

```
10
    Xaa Xaa Xaa Xaa Xaa Ala Cys Met Pro Pro Arg Arg Gln Lys Leu
           20
    35
                     40
                                  45
    55
    70
                              75
10
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Gln Phe Leu Arg Ser Met Met
              85
                           90
    Tyr Thr Phe Gly Asp Tyr Arg Asp Ile Cys Leu Asn Thr Asp Ile Ser
           100
                        105
                                     110
    Lys Lys Gln Asn Asp Val Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
15
                     120
         115
                                   125
    Xaa Xaa Xaa Xaa Ser Lys Ser Pro Ser Gly Leu Ser Arg Gln Glu
                   135
                                140
    Trp Trp Lys Thr Asn Gly Pro Glu Ile Trp Lys Gly Met Leu Cys Ala
    145
                150
                             155
20
    165
                           170
    180
                        185
    Xaa Xaa Xaa Xaa Xaa Lys Pro Gln Phe Leu Arg Trp Met Ile Glu
25
         195
                      200
                                   205
    Trp Gly Glu Glu Phe Cys Ala Glu Arg Gln Lys Lys Glu Asn Ile Ile
                   215
                                220
   230
                             235
30
   Lys His Arg Cys Asn Gln Ala Cys Arg Ala Tyr Gln Glu Tyr Val Glu
                          250
                                       255
    260
                        265
   35
         275
                     280
                                  285
    295
                                300
   Xaa Xaa Xaa Cys Xaa Cys
40
         (2) INFORMATION FOR SEQ ID NO:34:
        (i) SEQUENCE CHARACTERISTICS:
         (A) LENGTH: 7 amino acids
45
         (B) TYPE: amino acid(C) STRANDEDNESS: single
         (D) TOPOLOGY: linear
       (ii) MOLECULE TYPE: peptide
50
       (iii) HYPOTHETICAL: NO
       (iv) ANTISENSE: NO
        (v) FRAGMENT TYPE: N-terminal
```

(vi) ORIGINAL SOURCE:

55 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Pro Arg Arg Gln Xaa Leu Cys

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(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
- 65 (C) STRANDEDNESS: single

```
(D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
 5
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:
10
      CCRAGRAGRC AARAAYTATG
                                                                           20
                (2) INFORMATION FOR SEQ ID NO:36:
15
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
20
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
25
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:
      CCSMGSMGSC AGCAGYTSTG
                                                                          20
30
                (2) INFORMATION FOR SEQ ID NO:37:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 7 amino acids
35
               (B) TYPE: amino acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: peptide
40
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE: N-terminal
             (vi) ORIGINAL SOURCE:
45
            (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:
      Phe Ala Asp Xaa Xaa Asp Ile
50
                (2) INFORMATION FOR SEQ ID NO:38:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
55
              (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
            (ii) MOLECULE TYPE: cDNA
            (iii) HYPOTHETICAL: NO
60
            (iv) ANTISENSE: NO
            (v) FRAGMENT TYPE:
            (vi) ORIGINAL SOURCE:
            (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:
```

	IIIGCWGAIW WW3GWGAIAI	20
	(2) INFORMATION FOR SEQ ID NO:39:	
5	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
10	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO	
15	(v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:	
20	TTCGCSGATW WCSGSGACAT (2) INFORMATION FOR SEQ ID NO:40:	
25	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 6 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
30	 (ii) MOLECULE TYPE: peptide (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: N-terminal (vi) ORIGINAL SOURCE: 	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:	
	Pro Gln Phe Xaa Arg Trp 1 5	
40	(2) INFORMATION FOR SEQ ID NO:41:	
45	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 18 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
50	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:</pre>	
55	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:	
	CCAWCKKARR AATTGWGG	18
	(2) INFORMATION FOR SEQ ID NO:42:	
60	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single	
65	(D) TOPOLOGY: linear	

```
(ii) MOLECULE TYPE: cDNA
              (iii) HYPOTHETICAL: NO
              (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
 5
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:
       CCASCKGWAG AWCTGSGG
                                                                           18
10
                (2) INFORMATION FOR SEQ ID NO:43:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 7 amino acids
15
               (B) TYPE: amino acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: peptide
20
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE: N-terminal
             (vi) ORIGINAL SOURCE:
25
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:
      Glu Trp Gly Xaa Xaa Xaa Cys
                        5
30
                (2) INFORMATION FOR SEQ ID NO:44:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
35
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
40
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:
45
      CAAWAWTCWT CWCCCCATTC
                                                                           20
                (2) INFORMATION FOR SEQ ID NO:45:
50
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
55
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
60
             (VI) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:
      CAGWASTCST CSCCCCACTC
                                                                           20
65
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WE CLAIM:

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- A composition comprising a nucleotide sequence of the DBL gene family, wherein said nucleotide sequence is selected from the group consisting of the var-1, var-2, var-3 and var-7 genes.
- 2. The composition of Claim 1, wherein the nucleotide sequence of the *var-1*, *var-2*, *var-3* or *var-7* gene encodes a cysteine-rich domain homologous to a cysteine-rich domain of a Duffy Antigen Binding Protein (DABP) derived from *Plasmodium vivax* and a Sialic Acid Binding Protein (SABP) derived from *Plasmodium talciparum*.
- 3. The composition of Claim 1, wherein the nucleotide sequence of the *var-1*, *var-2*, *var-3* or *var-7* gene encodes a cysteine-rich interdomain region between a first domain and a second domain.
- 4. The composition of Claim 1, wherein the nucleotide sequence is derived from a coding region of SEQ ID NO:13 or SEQ ID NO:15.
 - 5. A composition comprising a polypeptide encoded by a nucleotide sequence of the *DBL* gene family, wherein said polypeptide is encoded by a *var-1*, *var-2*, *var-3* or *var-7* gene.
 - 6. The composition of claim 5, wherein the polypeptide comprises a sequence of amino acid residues homologous to cysteine-rich domains of a Duffy Antigen Binding Protein (DABP) derived from *Plasmodium vivax* and a Sialic Acid Binding Protein (SABP) derived from *Plasmodium falciparum*.
 - 7. The composition of claim 5, wherein the polypeptide comprises a sequence of about 300 to 400 amino acid residues occurring in the cysteine-rich interdomain region between a first domain and a second domain of a polypeptide encoded by the var-1, var-2, var-3 or var-7 gene.
 - 8. The composition of claim 5, wherein the polypeptide comprises a sequence of amino acid residues of SEO ID NO:14 or SEO ID NO:16.
 - 9. The composition of claim 5, wherein the polypeptide comprises a sequence of about 50 to about 325 amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
 - 10. The composition of claim 5, wherein the polypeptide comprises a sequence of about 75 to about 300 amino acid residues of SEO ID NO:14 or SEO ID NO:16.
 - 11. The composition of claim 5, wherein the polypeptide comprises a sequence of about 100 to about 250 amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
 - 12. The composition of claim 5, further comprising a pharmaceutically acceptable carrier and an isolated Duffy Antigen Binding Protein (DABP) binding domain polypeptide, a Sialic Acid Binding Protein (SABP) binding domain polypeptide, or a combination thereof, in an amount sufficient to induce a protective immune response to *Plasmodium* merozoites in a mammal.
 - 13. The composition of any of the preceding claims for use in inducing a protective immune response to *Plasmodium* merozoites in a mammal.
 - 14. Use of the composition of any one of claims 1-12 in the preparation of a medicament for inducing a protective immune response to *Plasmodium* merozoites in a mammal.
- 35 15. A method of inducing a protective immune response to *Plasmodium* merozoites in a mammal, comprising administering to a mammal an immunologically effective amount of a pharmaceutical composition

comprising a pharmaceutically acceptable carrier and an isolated cysteine-rich polypeptide encoded by a *var* gene selected from the group of genes consisting of *var-1*, *var-2*, *var-3* and *var-7* genes.

16. The method of claim 15, further comprising administering to said mammal an immunologically effective amount of a Duffy Antigen Binding Protein (DABP) binding domain polypeptide, a Sialic Acid Binding Protein (SABP) binding domain polypeptide, or a combination thereof.

GTDMEGIGYSK-X ₁₁ -GNDMDFGGYST-X ₁ 7-GTDYWNDLSNR-X ₁₅ -GLDVWRDINTN-X ₁₇ -	GKDLYLGYDNK-X37- GRDLYLGNPQR-X30- GRDHWDBDKSS-X32- NTDISKKONDV-X15- GTDISSKKDTS-X15-	RDXVSBLPTBVQKLKEKCX ₁₁ (3EELLERDNRSKLPKGKCX ₈ (3DXCQDKTKMIETLKVECX ₄ (3EECEEMGTEVKQLBKICX ₄ C	SDECRKKKKLENLQKQCX ₆ (SDECRKKNKKIKDVKRNCX ₁₂ C SWYCKAQSQBYDKLKKICX ₁₁ C SECABRQKKENIIKDACX ₈ C SNECKEQKKEYKLLAKCX ₁₁ C	FIG. 1	
G-X12-G-X5VCIPDRRYQLCMKEL-X47- DFCKDIRWSLGDFGDIIMGTDMEGIGYSK-X11-C-X10-C-X9VCIPDRRIQLCIVNL-X36- KFCNDLKNSELDYGHLAMGNDMDFGGYST-X17-C-X13-C-X10-VCVPPRRQELCLGNI-X36- EVCKIINKTEADIRDIIGGTDYWNDLSNR-X15-C-X11-VCGPPRRQCLCLGYI-X36- KICNAILGSXADIGDIVRGLDVWRDINTN-X17-	C-X15-C-X15-ACAPYRRLHLCDXNL-X43-QLCTVLARSEADIGDIVRGKDLYLGYDNK-X37-C-X15-C-X15-ACAPYRRLHVCDQNL-X45-QICTMLARSEADIGDIVRGRDLYLGNPQE-X30-C-X17-C-X31-VFLPPRREHMCTSNL-X55-AMCRAVRYSEADLGDIRGRDMWDBDKSS-X32-C-X10-C-X10-ACMPPRRQKLCLYYI-X52-QFLRSMMYTEGDYRDICLNTDISKKQNDV-X15-C-X10-C-X11-ACIPPRRQKLCLHYL-X51-DFKRQMFYTEADYRDICLGTDISSKKDTS-X15-	TDEKAQQRRKQMMNBSKAQIMTAMMYSV-X ₁₁ -C-X ₈ ePQIYRMIREMGRDYVSELPTEVQKLKEKCX ₁₁ C-X ₁ SEHKIKNFREMMNBFREKLMBAMLSEH-X6C-X6eLQITQMIKEMHGEELLERDNRSKLPKSKCX ₈ C-X ₀ NKKNDKLFRDEMMKVIKKDVMNVISWVF-X5C-X ₇ IPQFFRMFSEHGDDYCQDKTKMIETLKVECX ₄ C-X ₁ - KKQNDNNERNKMMEKQRNLIMSSMVKHI-X5C-X ₈ IPQFLRWLKEMGDEECEEMGTEVKQLEKICX ₄ C-X ₅	KGGDFFQLREDMMTSNRETVWKALICHA-X ₁₁ -C-X ₂₃ -VPQYLRWFEEWAEDECRKKKKKLENLQKQCX ₆ C-X _{1E} NDPEFPKLREDMMTANRETVWKAITCNA-X ₉ C-X ₂₃ -VPQYLRWFEEWAEDECRKKNKKIKDVKRNCX ₁₂ C-X ₂₂ KKPAYKKLRADMMEANRHQVWRAMKCAT-X ₄ C-X ₈ IPQRLRWTEWAEWACKAQSQBYDKLKKICX ₁₁ C-X ₆ -S-X ₆ -S-S-X ₆ -S-X ₆ -S-S-X ₆ -S-X ₆ -S-S-X ₆ -S-S-X ₆ -S-S-X ₆ -S-S-S-X ₆ -S-S-S-S-S-X ₆ -S-S-S-S-X ₆ -S-S-S-S-X ₆ -S-S-S-S-S-S-X ₆ -S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-S	MITRKN-X56CXC HIRSKP-X41-C-X7CXC HISKKK-X36-C-X20CXX-C MIKERKN-X38-C-X19CXX-C	CTNCSVWCRMYET MIDNQKK-X68-C-X30CXX-C CISCLYACNPYVD HINNQKE-X69-C-X40CXX-C CGRCKAACDKYKEBIBKHNEQWRK-X73-C-X6-C-X30-CXX-C KHRCNQACRAYQB YVENKKK-X43-C-X4CXC CVACKDQCKQXHS MIGIMID-X42-C-X8CXXXC
G-X12-G-X5VG C-X10-C-X9VC C-X13-C-X10-VC C-X12-C-X11-VC	C-X15-C-X15-AC C-X17-C-X31-VF C-X10-C-X10-AC C-X10-C-X11-AC	Tdekaoorrkohene Sehki knprkehene Nkkndkl frdehekv Kkondnnernkher	KGGDFFQLREDMMTS NDPEPFKLREDMMTA KKPAYKKLRADMMEA SKSPSGLSRQEMMT KISNSIRYRKSMMET	VPPCQNACKSYDQ RKECIDPCMKYRD DDNCKSKCNSYKE EKKCKNACSSYEK	CTNCSVWCRMYET CISCLYACNPYVD CGKCKAACDKYKBEIE KHRCNQACRAYQB CVACKDQCKQYHS
DABP F1 SABP F1 SABP F2 EBL-e1		DABP SABP F1 SABP F2 EBL-e1	EBL-e2 Proj3 F1 Proj3 F2 Proj3 F3	Dabp Sabp F1 Sabp F2 Ebl-el	ENL-e2 Proj3 F1 Proj3 F2 Proj3 F3
Family 1	Family 2	Family 1 Cont'd	Family 2 Cont'd	Family 1 Cont'd	Family 2 Cont'd

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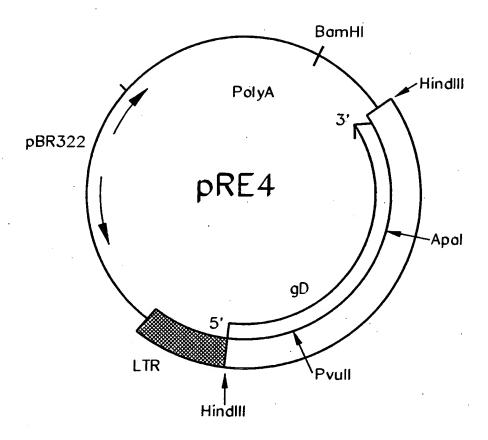


FIG. 2

3/5

FIG. 3

Concensus amino acid sequences and the synthetic oligonucleotide primers designed from them.

UNIEBP5 and 5A: PRRQ K/ELC

UNIEBP5, for A+T biased codon usage: CC(A/G)-AG(G/A)-AG(G/A)-CAA-(G/A)AA-(C/T)TA-TG

UNIEBP5A, for G+C biased codon usage: CC(C/G)-(C/A)G(C/G)-(C/A)G(C/G)-CAG-CAG-(C/T)T(C/G)-TG

UNIEBP5 B and C: F A D I/Y G/R D I

UNIEBP5B, for A+T biased codon usage: TTT-GC(A/T)-GAT-(A/T)(A/T)-(G/C)G(A/T)-GAT-AT

UNIEBP5C, for G+C biased codon usage: TTC-GC(G/C)-GAT-(A/T)(A/T)C-(G/C)G(G/C)-GAC-AT

UNIEBP3 and 3A: P Q F L/F R W

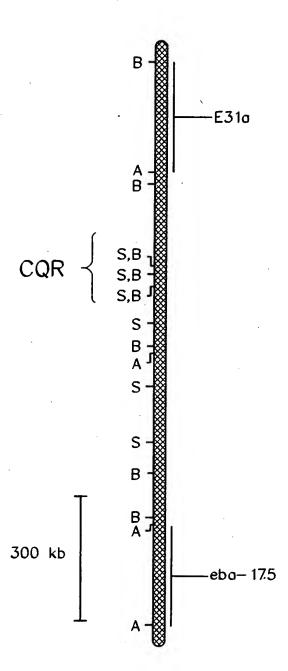
UNIEBP3, for A+T biased codon usage: CCA-(A/T)C(T/G)-(T/G)A(A/G)-(A/G)AA-TTG-(A/T)GG

UNIEBP3A, for G+C biased codon usage: CCA-(C/G)C(G/T)-G(A/T)A-GA(A/T)-CTG-(C/G)GG

UNIEBP3 B and C: E W G D/E D/E Y/F C

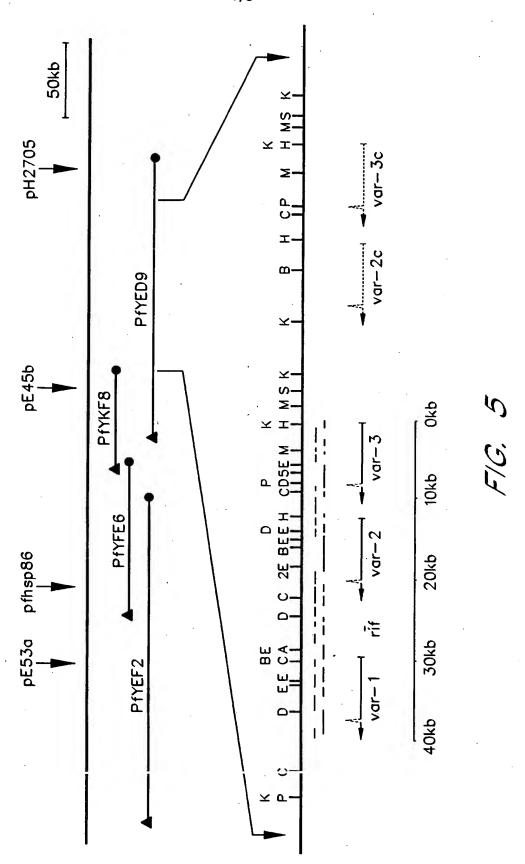
UNIEBP3B, for A+T biased codon usage: CA-A(A/T)A-(A/T)TC-(A/T)CC-CCA-TTC

UNIEBP3C, for G+C biased codon usage: CA-G(A/T)A-(G/C)TC-(G/C)TC-(G/C)CC-CCA-CTC G+C Biased



F/G. 4

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